# The role of bone morphogenetic protein 2 in the reprogramming of cancer stem cells.

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#### Abstract

Bone morphogenetic protein 2 (BMP-2) is a family member of the transforming growth factor-beta (TGF- $\beta$ ) superfamily and firstly recognized in early embryonic and postnatal development. BMP-2 has been reported to have crucial role in bone and cartilage formation, tissues and organs development, regulate cell differentiation, proliferation, angiogenesis, morphogenesis, chemotaxis, cellular survival and apoptosis. The BMPs are also identified as factors in tumor development and propagation; distinctly associated to diverse sides of carcinogenesis. The theory of cancer stem cells (CSCs) hypothesized that only a small hierarchical organization of cells is assisting tumorigenesis and inheriting cellular heterogeneity throughout long-life primary tumor. Reprogramming of CSCs using induced pluripotent stem cell (iPSC) approach possibly benefits in identifying the CSCs-related oncogenes, tumor-suppressor genes, and interactions between CSCs-related genes and the cancer microenvironment. Moreover, the reprogramming technology may provide crucial information related to cancer initiation and progression. This review will be focusing on BMP-2 signaling in modulating normal cells, human diseases, and cancer progression and suppression. Furthermore, this review will provide summary of updated reports on the role of BMP-2 in the developments of CSCs and its possible role as therapy through reprogramming technology by BMP-2 as an important regulatory factor in modulating the proliferation and aggressive properties of CSCs.

Keywords: Bone morphogenetic protein 2 (BMP-2), Reprogramming, Stem cells, Cancer stem cells (CSCs), Differentiation, Development.

#### Introduction

Bone morphogenetic proteins (BMPs) are well established as multi-functional cytokines; a family member of the transforming growth factor-beta (TGF- $\beta$ ) superfamily and firstly recognized in early embryonic and postnatal development. It was discovered in the late 1980s by Wozney et al. [1] based on the previous study which reported the activity of BMPs [2]. Following studies have reported BMPs to have crucial role in the formation of bone and cartilage. Furthermore, BMPs have been known to be part of tissues and organs development, regulate cell differentiation, proliferation, angiogenesis, morphogenesis, chemotaxis, cellular survival and apoptosis [3-5]. The BMPs are also identified as factors in tumor development and propagation; distinctly associated to diverse sides of carcinogenesis [6].

BMPs are categorized into four subgroups based on the structure, amino acid and the similarity of the nucleotide. Its

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phylogenetic comprises of BMP-2 and BMP-4, BMP-5, BMP-6, BMP-7 and BMP-8, BMP-9 and BMP-10, and BMP-12, BMP-13 and BMP-14 (GDF-5, GDF-6 and GDF-7) [7]. BMP-2 is known as osteogenic BMP which is based on its strong bone-inducing activity [8] and essential for endochondral bone formation [9]. BMP-2 is previously reported to promote the transformation process of undifferentiated cells at the beginning state [10]; induce bone and cartilage formation *in vivo* [11]; involve in cell differentiation, proliferation and apoptosis [5]; and the more recent studies regarding BMP-2 centering around regulation on tumorigenesis in several cancers [12-15].

Stem cells are undifferentiated cells, have self-renewal capability and can differentiate into specific matured cell types. Embryonic stem cells (ESCs) are pluripotent stem cells that able to differentiate to generate all types of tissues during embryonic development, whereas the adult stem cells (ASCs)

are crucial in replacing and repairing specific adult tissues [16]. Induced pluripotent stem cells (iPSCs) are reprogrammed somatic cells using specific measurements into stem cell-like cells, which have similar properties with embryonic stem cells [17]. Another important stem cell-like cell is the cancer stem cells (CSCs) which considered as a subpopulation of such stem cells. As the CSCs are found within tumors, their characteristics are similar to both stem cells and cancer cells. Uniquely, their asymmetrical cell division and alteration in gene regulations differentiate them from the normal stem cells [18,19].

The term reprogramming, in biology, refers to the reversing process of differentiated cells back into embryonic state. This biological reprogramming technology started to emerge in 2006 when Takahashi et al. demonstrated that by altering four genes, octamer 4 (*Oct4*), SRY box-containing gene 2 (*Sox2*), Kruppel-like factor 4 (*Klf4*) and oncogene c-Myc (*OSKM*) in adult mouse cells, a reprogrammed induced pluripotent stem cells (iPSCs) can be created, thus could be used in human medicine [17]. This reprogramming method has been used for reprogramming many types of cells including cancer cells due to similarity between CSCs and normal stem cells.

This review aims to provide an inclusive understanding of BMP-2 signaling in modulating normal cells, human bonerelated and non-bone diseases, and importantly cancer progression and suppression. Additionally, this review also emphasizes updated research reports on the role of BMP-2 in the developments of CSCs alongside normal stem cells and the possible future therapies utilizing reprogramming method on BMP-2 as an important regulatory tool in modulating the development of CSCs.

## **Role of BMP-2 in Normal and Disease Cellular Progression**

Signaling pathways involving BMPs progression are divided into canonical and non-canonical pathways. In general, BMPs signaling pathway is functioning when a heterotetrameric signaling dimer complexes of type 1 and type 2 receptors is formed. All receptors have a short extracellular domain, a single transmembrane domain, and an intracellular domain. Type 1 receptor consists of seven receptors, activin A receptor like type 1 (ACVRL1), activin A receptor type 1 (ACVR1), activin A receptor type 1B (ACVR1B), activin A receptor type 1C (ACVR1C), bone morphogenetic protein receptor type 1A (BMPR1A), bone morphogenetic protein receptor type 1B (BMPR1B) and transforming growth factor beta receptor 1 (TGF- $\beta$ R2), and five type 2 receptors, activin A receptor type 2A (ACVR2A), activin A receptor type 2B (ACVR2B), anti-Mullerian hormone receptor type 2 (AMHR2), bone morphogenetic protein receptor type 2 (BMPR2) and transforming growth factor beta type 2 (TGF-βR2) [20]. The heterotetrameric signaling complex mechanisms can be altered depending on what type of BMPs is initiated hence can activate different pathways. The signal transduction cascade triggered by the canonical pathway by binding to cell surface receptors and creating a heterotetrameric dimer complexes of type 1 and

type 2 receptors [21]. For example, BMP-2 and BMP-4 are preferred to bind to type 1 receptors and only enlisted type 2 receptors, while BMP-6 and BMP-7 interact with type 2 receptors and recruit type 1 receptors [22]. On the other hand, non-canonical pathways such as mitogen-activated protein kinase (MAPK) and Smad-independent signaling pathway are led to regulation of gene expression. Moreover, BMP signaling is also being regulated by intracellular (PKBP12, microRNAs, phosphatases, and I-Smads), extracellular (Noggin), and membrane (Endoglin) modulators [23]. Such example of noncanonical pathway of BMP-2 is reported in development of the dental epithelium where TGF- $\beta$  signaling initially triggered the activation of Smad1, Smad5 and Smad8 in this tissue. The report mentioned that the levels of P-Smad1, P-Smad5, and P-Smad8 are maintained in both dorsomorphin-treated dental epithelium of the tooth germs and the dental epithelium of Msx1 mutant, in which BMP-2 expression is decreased [24].

BMPs are considered as powerful stimulators for both bone formation and other related cellular functions. BMPs activities are controlled by specific molecular proteins at specific molecular levels. These activities can be either a list of BMP antagonists bind BMP ligands and inhibit BMP functions, the binding of Smad6 to type 1 BMP receptors prevents the binding and phosphorylation of Smad1 and 5 [25], or a selectively binding of tob (an anti-proliferative protein) to Smad1 and 5, thus blocks BMP signaling in osteoblasts [26], or even interaction of Smurf1, an E3 ubiquitin ligase (Smad ubiquitin regulatory factor 1) together with Smad1 and 5, mediates the degradation of the Smad proteins [27].

BMP-2 has been shown to have physiological function in both bone formation and development. This role had been demonstrated by Chen et al. by injecting BMP-2 around the calvariae of mice's surface, hence inducing the formation of periosteal bone locally without an initial cartilage step [28]. BMP-2 has been demonstrated to regulate proliferation and osteogenesis, and lacking of BMP-2 will results in serious defects in repair sites of the osteoblasts [29,30]. Moreover, BMP-2 can inhibit the differentiation of osteoprogenitor cells originated from multipotential mesenchymal cells into osteoblasts [31]. Inadequate level of BMP-2 will also slow down the process of bone healing and repair.

BMP-2 also has been associated with osteoarthritis (OA). OA is a disease which affects synovial joints, like knee, hip and hand due to degeneration of articular cartilage. Degeneration of cartilage of OA tissues [32] and chondrocyte hypertrophy might be due to dysregulation of BMP-2 response [33]. Some indicators of disease severity and joint arthroplasty is higher level of BMP-2 and BMP-4 serum, but there was no association of BMP-2 and BMP-5 in OA progression [34,35].

The BMP-2 antagonist's mutations have unveiled the importance of BMPs that is being modulated in a specific system. For instant, proximal symphalangism and multiple synostoses syndrome caused by heterozygous mutations of the human noggin gene, have the same symptoms. Proximal symphalangism, is a disorder which has an autosomal-dominant with conductive deafness, carpal and tarsal bone

fusion, and ankylosis of the proximal interphalangeal joint [36,37], while multiple synostoses syndrome is a disorder of joint morphogenesis [38]. In these disorders, noggin protein will bind and inactivated BMP-2, 4 and 7. In addition, 3D crystal structure clearly demonstrates the function of noggin which mainly targets BMP-2, 4 and 7, hence inhibits them [39]. Furthermore, BMP-induced and Smad-dependent transcription in osteoblasts were inhibited by Tob and associated with Smad1 and Smad5 proteins [40]. The Tob consists of Tob, Tob2, BTG1, BTG2 and BTG3 are belonging to an anti-proliferative protein family [41]. The study of Tob in knockout mice has demonstrated that the higher level of BMP-2 effects on osteoblast proliferation, differentiation and the local bone formation [42].

In general, BMPs have multifunctional cytokines. Not only BMPs regulate the development of both bone and cartilage, but BMPs also take part in many non-osteogenic development processes. BMPs play crucial roles in maintaining adult's tissue homeostasis and depletion of BMP production of functionality normally causes marked defects or severe pathologies. Ectodermal cell fates for example, are determined by neural induction [16] and BMPs perform as indicator of epidermal induction [43]. BMP-2 in focus, administers neuronal phenotypes developmental from neural crest cells [44]. The process of myogenesis is being inhibited by BMPs when they direct the somite development. For example in the limb bud, when BMP-2 interacts with the fibroblast growth factor 4 and sonic hedgehog, the expansion is inhibited, and the chondrocytes and osteoblast precursors formation is induced [45,46].

The BMP-2 potential in inducing bone and cartilage formation can also be used to understand the mechanism of certain diseases, hence using recombinant human BMP-2 (rhBMP-2) in disease treatments is applied. Such in cleft palate (CP), an observable birth defect that has multiple etiologies, BMP-2 is involved in palate morphogenesis in development, and syndromic CP is associated with haploinsufficiency of BMP-2 [47]. In embryogenesis, deletion of BMP-2 may results in embryonic lethality and previous report had shown that the malfunction of amnion/chorion and cardiac development happened in BMP-2 deficient mice [48]. Meanwhile, BMP-2 also modulates cartilage development, and for chondrocyte proliferation and maturation, BMP-2 is considered as a main factor in endochondral bone development [9]. BMP-2 also may results in a serious chondrodysplasia phenotype, a congenital disorder of bone and cartilage development [49]. Additionally, BMP-2 is important in homeostasis and fracture healing. BMP-2 initiates the fracture healing and limb-specific BMP-2 knockouts, a down-regulated of BMP-2 results in sudden fractures and fails to start the healing process [50].

The BMP-2 signaling is also required for normal growth and morphogenesis of the developing gastrointestinal tract [51]. Additionally, BMP-2 homozygous mutants caused abnormalities in the development of the heart, results in malformation of the amnion, chorion and embryo death [48]. BMP-2 is expressed in both extraembryonic mesoderm and myocardium, and the BMP-2 signaling in myocardium is crucial for the formation of endocardial cushion (EC). Moreover, the regulation of BMP-2 signaling triggers underlying endothelium forming ECs is depending on an epithelial-mesenchymal transformation (EMT) [52]. ECs finally produce the differentiate heart septa and valves, and allow the development of a mature heart with four chambers. Specific deletion of BMP-2 in cardiac progenitors block the formation of the four-chambered heart causing the heart valve region turns differentiated chamber myocardium. Further study has confirmed the function of BMP-2 in EC EMT when deletion of BMP-2 in atrioventricular (AV) happened, plus in development of cardiac jelly and AV myocardium [53].

Looking at molecular levels of every cells, a single individual of cell must have some very small differences in their genetic materials, DNA. Such differences might occur in either by programmed differences in specialized cells, random mutation and stability of DNA or chimerism and colonization. There are many factors can cause continuous alteration of DNA segments such as environmental damage, chemical degradation, genome instability, and small but significant errors in DNA replication and DNA repair [16]. There were so many studies of mutations of BMPs show the crucial roles of BMPs in various kinds of inherited diseases. Dysfunction of BMP-2 regulations is also being associated with the oral epithelium [54] and prostate cancer cells malignancy [55]. Moreover, further investigations have been done in the embryonic development and postnatal life, to investigate on how BMP ligands, receptors and signaling proteins are functioning by using the null mutations of these factors in animal models. For instant, inadequate BMP-2 in mice reduced their ability to survive independently after birth. Homozygous BMP-2 mutant embryos had cardiac developmental defect which demonstrated by the heart development abnormality in the exocoelomic cavity and die between 7 and 11 days period [48]. In hypertropic cartilage of BMP-2 null mutant mice, BMP-2 might functionally silent BMP-6, since BMP-2 and 6 are co-expressed in hypertropic cartilage [56,57].

## **BMP-2** as Potential Diseases and Cancer Therapies

BMP-2 in cancers can have either positive or negative effects in tumorigenesis and metastasis. BMP-2 can either act as tumor suppressors or tumor promoters through different mechanisms. BMP-2 can activate oncogenes, and initiate metastasis progression in tumor microenvironment. Evidence of BMP-2 and signaling components as a novel biomarker for cancer treatment with significant therapeutic implications remains controversial. Due to significant reduction of BMP-2 in prostate cancer compared to benign prostate tissue, it may functions as a marker of poor prognosis [58]. Moreover, the low expression of BMP-2 in epithelial ovarian cancer tissue also proposed that it probably obtain indigent prognosis of ovarian cancer patients.

Besides that, BMP-2 has negative modulation on miRNAs. miRNAs structures are short, non-coding RNAs of 18 to 25

nucleotides long that important in variety tumorigenic processes [59]. miRNAs profiling on C2L12 mesenchymal cells (a BMP-2-stimulated osteogenesis) distinguished two miRNA representatives and demonstrated miR-133 directly triggers Runx2, that essential for bone formation, and miR-135 may target SMAD5 (a signal osteogenic transducer of BMP-2) [60]. Furthermore, BMP-2 also relates to the study of drug resistance of cancer cells. For example, knockdown of BMP-2 increased chemo-resistance of the MCF-7 in breast cancer cell line [61], while BMP-2 treatment in in vivo models increased tumor development and chemotherapy resistance [62]. On the other hand, Persano et al. demonstrated that based treatment using BMP-2, escalated the temozolomide response in glioblastoma multiforme (GBM) cells with hypoxic drugresistant and this chemotherapy resistance is reported as one of the leading factors for poor GBM among the most aggressive tumor types [63].

BMP-2 has been shown to have the osteoinductive capabilities in clinical trial studies. Variety of animal models such as mice, rabbits, dogs, sheep and other laboratory animals are used to evaluate and demonstrated the capability of BMP-2 to treat bone deformity in major-sized defects [64]. Such animal models, massive bone errors are not curable without a therapeutic interference, thus eases analysis of the BMP-2 abilities in inducing bone. Among the studies are BMP-2-gene therapy studies where they showed that the implantation of transfected BMP-2-bone marrow mesenchymal stem cells with a bioresorbable polymer mixture, healed the bone defects [65]. Additionally, recombinant human BMP-2 (rhBMP-2) that has been systemically administrated in mouse models, had shown positive regulation of mesenchymal stem cell activity and overturn the loss of age-related bone and ovariectomy-induced [66]. Therefore, BMP-2 might be useful in treating osteoporosis. Moreover, rhBMP-2 also demonstrated as an enhancer of bone healing in a rat femoral bone defect model and a rabbit ulna osteotomy model by delivering rhBMP-2 using a carrier such as calcium phosphate or liposome [67,68]. More clinical studies have shown the utilization of rhBMP-2 as a complete bone graft replacement in spinal fusion surgery [69,70] and several studies have demonstrated that the induction efficacy of BMP-2 in fusion is much way better than autogenous bone graft [69,71], and very useful in intervertebral and lumbar posterolateral fusion [72]. In dentistry, BMP-2 also can induce the formation of new dentine, plus has likely to be a substitute for root canal surgery and a very effective bone inducer for periodontal reconstructive implantation [73].

## Stem Cell Differentiation and BMP-2

Stem cell biology has contributed a vantage point in addressing the problems in developmental biology. The development of human obeys the predetermined dogma from fertilized egg until it becomes a complete complex, multi-cellular organism. Stem cells are unspecialized cells, self-renewal and capable to differentiate into variety of specialized cell types. There are three types of germ layers developed from the fertilized egg: endoderm, ectoderm, and mesoderm. From these primitive cell types, they develop into all tissues of organism [74]. Several studies have showed that BMP-2 mediated osteogenesis from mesenchymal stem cell (MSC) precursors. For instance, improvement of the osteogenic differentiation of stem cells was triggered by the boost of BMP-2 binding efficiency [75,76]. Moreover, BMP-2 also has been demonstrated to activate WNT/ $\beta$ -catenin signaling and promote the differentiation of human dental pulp cells (HDPCs), which then mediate by p38 MAPK *in vitro* [77]. BMP-2 antagonist noggin has also been reported to regulate human ESCs differentiation and induce the novel cell types that give rise to neural precursors [78].

Several cancers originate from blood, brain, breast, skin, and gut, are derived from a minor group of stem cells from specific tissues, which function mainly for development, conserve their proliferation potential and to minimize DNA replication errors [79]. Adult stem cells are somatic cells that have self-renewal ability and able to differentiate into specialized cells. Normal stem cells must possess those two unique abilities as mentioned before. A normal stem cell is said to be self-renewal due to its property of producing more identical stem cells with similar replication potential and development. This ability enables mass production of the stem cells in response to intracellular and extracellular environments, hence initiate the proliferation and regulation of those cells in tissue and organs. In addition, a normal stem cell must also able to differentiate into tissue-specific specialized cells. Hematopoietic stem cells for example normally produce both myeloid and lymphoid progenitor cells which then give rise to variety of differentiated cells such as macrophages, monocytes, basophils, neutrophils, eosinophils, platelets, and erythrocytes (myeloid); T cells, B cells and natural killer cells (lymphoid) [80,81]. Such ability has brought attention for scientists to study the growth potential of stem cells in vitro and in vivo [82,83]. Stem cells also have a longer lifespan compared to matured cells. In the blood system for instance, terminally differentiated stem cells such as macrophages and basophils have a short lifespan because they generally die after normal tissue maintenance or cellular damage [84]. Therefore, stem cells are higher probability to cause the mutation rather than the matured cells.

Mutations resulted from aberrant mitoses in the regulatory systems that suppress abnormal proliferation. This might happen during mitotic division when a parent stem cell is selfrenewing itself continuously. Majority of the mutations are benign because the abnormal cells are usually eradicated from the normal pool of dividing cells. However, at several phases, these abnormal cells could get accumulated and might trigger the development of cancer. Most mutations are affecting protein regulations of cell division, DNA damages and repair mechanisms as well as signaling pathways. Stem cells are said to be the target for mutations because they are the only longlived cells in nearly the entire tissues that are vulnerable to genotoxic stresses compared to their specialized progeny [85]. From stem cell and cancer studies, CSCs emerged and believed to originate from normal stem cells or progenitor cells, which promote tumors when encountering specific genetic mutation or environmental alterations [86]. The study of genetic alterations in differentiation of stem cells is a crucial approach for regeneration of defective tissue in stem cells therapy. Therefore, adding BMP-2 as a key factor on differentiation of stem cells is something that is worth to be investigated.

## Possible Role of BMP-2 in CSCs Reprogramming

Stem cells role in cancer was discovered in 1994, reported by Lapidot et al. [87] followed by identification of CSC proposed by Bonnet and Dick in their research involving human acute myeloid leukemia (AML) [88]. After sample transplantation from patients with AML into severe combined immunedeficient (SCID) mice, they were able to identify an AMLinitiating cell population. CSCs were later identified in many common solid tumors, including leukemia [87-89], breast cancer [90], colorectal cancer [91-93], and brain cancer [94].

The theory of CSCs hypothesized that only a small hierarchical organization of cells is assisting tumorigenesis and inheriting cellular heterogeneity throughout long-life primary tumor. CSCs do not really emerge from normal tissue stem cells modification even though they possess unique stem cell properties. Moreover, several observations have shown that cancers are resistant to both chemotherapy and radiation treatment, hence explains the tumor dormancy and metastasis phenomenon [95]. The studies of CSCs have encouraged the advanced treatment strategies for cancers focusing on eliminating CSCs, and not diminishing tumor size [96]. The origin of CSCs has speculated that, are they either really emerged from normal stem cells or normal somatic cells that gone mad over their regulatory and growth mechanisms? Perhaps progenitor or differentiated cells can obtain stem cell properties through mutations and cancer? Either theory speculated another important question is how CSC is functioning? CSCs require specific regulatory networks to exert their carcinogenic functions, such as cytokines from the cancer cell microenvironments. Therefore, novel cancer therapies might be developed through the elucidation of these pathways [97]. Because of the same properties of self-renewal and differentiation shared by both normal and CSCs, they may also share similar regulatory mechanisms relating to cell function stemness. Several pathways including Wnt pathways, Bmi-1, c-myc, Notch and Hedgehog (Hh) are examples of shared pathways in both normal and CSCs [98].

CSC studies have contributed to the search of novel cancer targeted therapies. By targeting the distinct functional and molecular properties of CSCs, it would be improving the efficacy of cancer therapies. Clinically, deciphering mechanisms of chemo- and radioresistance that control in CSCs is vital. Many targeting strategies are being explored within the different aspects of CSCs such as self-renewal pathways, quiescence, radio-resistance and CSC-specific cells surface molecules as reviewed by Batlle et al. [99]. In addition, the discrete molecular and functional properties of CSCs may also represent therapeutic liabilities that could be utilized for other novel combination strategies development. For BMP-2 and TGF- $\beta$  family in general, there are few strategies of treatment under development that combines both anti-CSCs

with chemotherapy. For instance, the self-renewal ability of triple-negative breast CSCs have been improvably inhibited using an anti-proliferative agent named paclitaxel by implicating TGF- $\beta$  type 1 receptor in *in vivo* models [100]. Besides, the cronic myeloid leukemia (CML) TGF- $\beta$ -Akt signaling inhibition suppressed imatinib cytotoxicity and apoptosis in CSCs, which sequentially regulated the nuclear localization of FOXO3a [101]. BMP-2 in specific has been demonstrated to sensitize glioblastoma stem-like cells to temozolomide (TMZ) by influencing the stability of hypoxia-inducible factor-1 $\alpha$  (HIF-1 $\alpha$ ) and O6-methylguanine-DNA-methyltransferase (MGMT) expression [102].

Elucidating the problems arise as mentioned before about CSCs may be benefiting for the development of novel cancer therapies. Such problems may require a new methodology. One of the most recent technologies is the use of cancer cellreprogramming approach using induced pluripotent stem cell (iPSC) technology. The reprogramming technology allows for the discovery of CSC-related oncogenes, anti-oncogenes, tumor-suppressor genes and epigenomes. This method also benefits for studying the associations between CSC microenvironment and its related genes, plus the mechanisms of cancer stem initiation and progression. However, this reprogramming method still faces so many challenges such as the chromosomal aberrations, genetic mutations, and cancer-The fundamental knowledge of specific epigenetic. reprogramming introduced by Takahashi and Yamanaka of using four specific transcription factors OSKM by generating stem cells-like cells provided a stepping stone for more researches to study the functional mutations of cancerassociated genes and genome epigenetic alterations, hence understanding the molecular mechanisms of tumorigenesis in humans [17].

Previously, BMP-2 has been reported as a key regulator in several normal and CSCs [103,104]. The reprogramming of *BMP-2* gene modification of iPSC-MSCs for bone tissue engineering had been done previously. Liu et al. demonstrated BMP-2-iPSC-MSC on Arg-Gly-Asp-calcium phosphate cement (RGD-CPC) enhanced differentiation and bone mineral production [105]. Unfortunately, there is no report about BMP-2 in the reprogramming of CSCs. Therefore, BMP-2 modulation in iPSC-CSCs is worth to be investigated.

## Conclusions

In summary, BMP-2 is one of the most important factors in regulating bone and cartilage formation. Additionally, BMP-2 also regulates tumorigenesis in several cancers. The studies of BMP-2 may provide more views and understanding about the signaling pathways and molecular properties of CSCs hence utilizing BMP-2 as a modulating factor. Moreover, a precise approach is needed to deliver the BMP-2 into targeted cells such as reprogramming technology. Better results are expected from using the suitable approach thus providing more conclusive insights on CSCs progression and suppression to benefit cancer therapy.

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#### References

- Wozney JM, Rosen V, Celeste AJ, Mitsock LM, Whitters MJ, Kriz RW, Hewick RM, Wang EA. Novel regulators of bone formation: molecular clones and activities. Science 1988; 242: 1528-1534.
- 2. Urist MR. Bone: formation by autoinduction. Science 1965; 150: 893-899.
- 3. Hogan BL. Bone morphogenetic proteins in development. Curr Opin Genet Dev 1996; 6: 432-438.
- Hogan BL. Bone morphogenetic proteins: multifunctional regulators of vertebrate development. Genes Dev 1996; 10: 1580-1594.
- Hruska KA, Mathew S, Saab G. Bone morphogenetic proteins in vascular calcification. Circ Res 2005; 97: 105-114.
- 6. Hardwick JC, Kodach LL, Offerhaus GJ, van den Brink GR. Bone morphogenetic protein signalling in colorectal cancer. Nat Rev Cancer 2008; 8: 806-812.
- 7. Mueller TD, Nickel J. Promiscuity and specificity in BMP receptor activation. FEBS Lett 2012; 586: 1846-1859.
- Luu HH, Song WX, Luo X, Manning D, Luo J, Deng ZL, Sharff KA, Montag AG, Haydon RC, He TC. Distinct roles of bone morphogenetic proteins in osteogenic differentiation of mesenchymal stem cells. J Orthop Res 2007; 25: 665-677.
- Shu B, Zhang M, Xie R, Wang M, Jin H, Hou W, Tang D, Harris SE, Mishina Y, OKeefe RJ, Hilton MJ, Wang Y, Chen D. BMP2, but not BMP4, is crucial for chondrocyte proliferation and maturation during endochondral bone development. J Cell Sci 2011; 124: 3428-3440.
- Chapellier M, Bachelard-Cascales E, Schmidt X, Clement F, Treilleux I, Delay E, Jammot A, Menetrier-Caux C, Pochon G, Besancon R, Voeltzel T, Caron de Fromentel C, Caux C, Blay JY, Iggo R, Maguer-Satta V. Disequilibrium of BMP2 levels in the breast stem cell niche launches epithelial transformation by overamplifying BMPR1B cell response. Stem Cell Reports 2015; 4: 239-254.
- 11. Barboza E, Caula A, Machado F. Potential of recombinant human bone morphogenetic protein-2 in bone regeneration. Implant Dent 1999; 8: 360-367.
- Beck SE, Jung BH, Fiorino A, Gomez J, Rosario ED, Cabrera BL, Huang SC, Chow JY, Carethers JM. Bone morphogenetic protein signaling and growth suppression in colon cancer. Am J Physiol Gastrointest Liver Physiol 2006; 291: 135-145.
- 13. Kawamura C, Kizaki M, Yamato K, Uchida H, Fukuchi Y, Hattori Y, Koseki T, Nishihara T, Ikeda Y. Bone morphogenetic protein-2 induces apoptosis in human

myeloma cells with modulation of STAT3. Blood 2000; 96: 2005-2011.

- Piccirillo SG, Reynolds BA, Zanetti N, Lamorte G, Binda E, Broggi G, Brem H, Olivi A, Dimeco F, Vescovi AL. Bone morphogenetic proteins inhibit the tumorigenic potential of human brain tumour-initiating cells. Nature 2006; 444: 761-765.
- 15. Piccirillo SG, Vescovi AL. Bone morphogenetic proteins regulate tumorigenicity in human glioblastoma stem cells. Ernst Schering Found Symp Proc 2006; 59-81.
- Strachan T, Read A. Human Molecular Genetics (4th Edn.). Garland Sciences, Taylor & Francis Group LLC 2011.
- 17. Takahashi K, Yamanaka S. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. Cell 2006; 126: 663-676.
- Al-Hajj M, Clarke MF. Self-renewal and solid tumor stem cells. Oncogene 2004; 23: 7274-7282.
- Beckmann J, Scheitza S, Wernet P, Fischer JC, Giebel B. Asymmetric cell division within the human hematopoietic stem and progenitor cell compartment: identification of asymmetrically segregating proteins. Blood 2007; 109: 5494-5501.
- 20. Hinck AP, Mueller TD, Springer TA. Structural biology and evolution of the TGF-beta family. Cold Spring Harb Perspect Biol 2016; 8.
- 21. Heldin CH, Miyazono K, ten Dijke P. TGF-beta signalling from cell membrane to nucleus through SMAD proteins. Nature 1997; 390: 465-471.
- 22. de Caestecker M. The transforming growth factor-beta superfamily of receptors. Cytokine Growth Factor Rev 2004; 15: 1-11.
- 23. Corradini E, Babitt JL, Lin HY. The RGM/DRAGON family of BMP co-receptors. Cytokine Growth Factor Rev 2009; 20: 389-398.
- 24. Yuan G, Yang G, Zheng Y, Zhu X, Chen Z, Zhang Z, Chen Y. The non-canonical BMP and Wnt/beta-catenin signaling pathways orchestrate early tooth development. Development 2015; 142: 128-139.
- 25. Hata A, Lagna G, Massague J, Hemmati-Brivanlou A. Smad6 inhibits BMP/Smad1 signaling by specifically competing with the Smad4 tumor suppressor. Genes Dev 1998; 12: 186-197.
- Wang M, Jin H, Tang D, Huang S, Zuscik MJ, Chen D. Smad1 plays an essential role in bone development and postnatal bone formation. Osteoarthritis Cartilage 2011; 19: 751-762.
- 27. Zhang Y, Chang C, Gehling DJ, Hemmati-Brivanlou A, Derynck R. Regulation of Smad degradation and activity by Smurf2, an E3 ubiquitin ligase. Proc Natl Acad Sci USA 2001; 98: 974-979.
- Chen D, Harris MA, Rossini G, Dunstan CR, Dallas SL, Feng JQ, Mundy GR, Harris SE. Bone morphogenetic protein 2 (BMP-2) enhances BMP-3, BMP-4, and bone cell differentiation marker gene expression during the induction of mineralized bone matrix formation in

cultures of fetal rat calvarial osteoblasts. Calcif Tissue Int 1997; 60: 283-290.

- 29. Rosen V. BMP2 signaling in bone development and repair. Cytokine Growth Factor Rev 2009; 20: 475-480.
- Bais MV, Wigner N, Young M, Toholka R, Graves DT, Morgan EF, Gerstenfeld LC, Einhorn TA. BMP2 is essential for post natal osteogenesis but not for recruitment of osteogenic stem cells. Bone 2009; 45: 254-266.
- 31. Yin T, Li L. The stem cell niches in bone. J Clin Invest 2006; 116: 1195-1201.
- 32. Nakase T, Miyaji T, Tomita T, Kaneko M, Kuriyama K, Myoui A, Sugamoto K, Ochi T, Yoshikawa H. Localization of bone morphogenetic protein-2 in human osteoarthritic cartilage and osteophyte. Osteoarthritis Cartilage 2003; 11: 278-284.
- 33. Papathanasiou I, Malizos KN, Tsezou A. Bone morphogenetic protein-2-induced Wnt/beta-catenin signaling pathway activation through enhanced lowdensity-lipoprotein receptor-related protein 5 catabolic activity contributes to hypertrophy in osteoarthritic chondrocytes. Arthritis Res Ther 2012; 14: 82.
- Bijsterbosch J, Kloppenburg M, Reijnierse M, Rosendaal FR, Huizinga TW, Slagboom PE, Meulenbelt I. Association study of candidate genes for the progression of hand osteoarthritis. Osteoarthritis Cartilage 2013; 21: 565-569.
- 35. Albilia JB, Tenenbaum HC, Clokie CM, Walt DR, Baker GI, Psutka DJ, Backstein D, Peel SA. Serum levels of BMP-2, 4, 7 and AHSG in patients with degenerative joint disease requiring total arthroplasty of the hip and temporomandibular joints. J Orthop Res 2013; 31: 44-52.
- Shaw L, McCaul J, Irwin GJ, Huntley JS. Foot anomalies and proximal symphalangism. Clin Anat 2012; 25: 781-784.
- Cushing H. Hereditary Anchylosis of the proximal Phalan-Geal joints (Symphalangism). Genetics 1916; 1: 90-106.
- Krakow D, Reinker K, Powell B, Cantor R, Priore MA, Garber A, Lachman RS, Rimoin DL, Cohn DH. Localization of a multiple synostoses-syndrome disease gene to chromosome 17q21-22. Am J Hum Genet 1998; 63: 120-124.
- 39. Groppe J, Greenwald J, Wiater E, Rodriguez-Leon J, Economides AN, Kwiatkowski W, Affolter M, Vale WW, Izpisua Belmonte JC, Choe S. Structural basis of BMP signalling inhibition by the cystine knot protein Noggin. Nature 2002; 420: 636-642.
- Yoshida Y, Tanaka S, Umemori H, Minowa O, Usui M, Ikematsu N, Hosoda E, Imamura T, Kuno J, Yamashita T, Miyazono K, Noda M, Noda T, Yamamoto T. Negative regulation of BMP/Smad signaling by Tob in osteoblasts. Cell 2000; 103: 1085-1097.
- 41. Baranzini SE. The role of antiproliferative gene Tob1 in the immune system. Clin Exp Neuroimmunol 2014; 5: 132-136.

- 42. Usui M, Yoshida Y, Yamashita T, Tsuji K, Isao I, Yamamoto T, Nifuji A, Noda M. Enhancing effect of Tob deficiency on bone formation is specific to bone morphogenetic protein-induced osteogenesis. J Bone Miner Res 2002; 17: 1026-1033.
- 43. Munoz-Sanjuan I, Brivanlou AH. Neural induction, the default model and embryonic stem cells. Nat Rev Neurosci 2002; 3: 271-280.
- 44. Christiansen JH, Coles EG, Wilkinson DG. Molecular control of neural crest formation, migration and differentiation. Curr Opin Cell Biol 2000; 12: 719-724.
- 45. Niswander L, Martin GR. FGF-4 regulates expression of Evx-1 in the developing mouse limb. Development 1993; 119: 287-294.
- 46. Wall NA, Hogan BL. TGF-beta related genes in development. Curr Opin Genet Dev 1994; 4: 517-522.
- 47. Williams ES, Uhas KA, Bunke BP, Garber KB, Martin CL. Cleft palate in a multigenerational family with a microdeletion of 20p12.3 involving BMP2. Am J Med Genet A 2012; 158: 2616-2620.
- 48. Zhang H, Bradley A. Mice deficient for BMP2 are nonviable and have defects in amnion/chorion and cardiac development. Development 1996; 122: 2977-2986.
- 49. Wu M, Chen G, Li YP. TGF-beta and BMP signaling in osteoblast, skeletal development, and bone formation, homeostasis and disease. Bone Res 2016; 4: 16009.
- Fassbender M, Minkwitz S, Strobel C, Schmidmaier G, Wildemann B. Stimulation of bone healing by sustained bone morphogenetic protein 2 (BMP-2) delivery. Int J Mol Sci 2014; 15: 8539-8552.
- Torihashi S, Hattori T, Hasegawa H, Kurahashi M, Ogaeri T, Fujimoto T. The expression and crucial roles of BMP signaling in development of smooth muscle progenitor cells in the mouse embryonic gut. Differentiation 2009; 77: 277-289.
- 52. Rivera-Feliciano J, Tabin CJ. Bmp2 instructs cardiac progenitors to form the heart-valve-inducing field. Dev Biol 2006; 295: 580-588.
- Ma L, Lu MF, Schwartz RJ, Martin JF. Bmp2 is essential for cardiac cushion epithelial-mesenchymal transition and myocardial patterning. Development 2005; 132: 5601-5611.
- 54. Jin Y, Tipoe GL, Liong EC, Lau TY, Fung PC, Leung KM. Overexpression of BMP-2/4, -5 and BMPR-IA associated with malignancy of oral epithelium. Oral Oncol 2001; 37: 225-233.
- 55. Harris SE, Harris MA, Mahy P, Wozney J, Feng JQ, Mundy GR. Expression of bone morphogenetic protein messenger RNAs by normal rat and human prostate and prostate cancer cells. Prostate 1994; 24: 204-211.
- 56. Chen D, Zhao M, Mundy GR. Bone morphogenetic proteins. Growth Factors 2004; 22: 233-241.
- 57. Solloway MJ, Dudley AT, Bikoff EK, Lyons KM, Hogan BL, Robertson EJ. Mice lacking Bmp6 function. Dev Genet 1998; 22: 321-339.

- 58. Horvath LG, Henshall SM, Kench JG, Turner JJ, Golovsky D, Brenner PC, ONeill GF, Kooner R, Stricker PD, Grygiel JJ, Sutherland RL. Loss of BMP2, Smad8, and Smad4 expression in prostate cancer progression. Prostate 2004; 59: 234-242.
- 59. Calin GA, Sevignani C, Dumitru CD, Hyslop T, Noch E, Yendamuri S, Shimizu M, Rattan S, Bullrich F, Negrini M, Croce CM. Human microRNA genes are frequently located at fragile sites and genomic regions involved in cancers. Proc Natl Acad Sci USA 2004; 101: 2999-3004.
- Li Z, Hassan MQ, Volinia S, van Wijnen AJ, Stein JL, Croce CM, Lian JB, Stein GS. A microRNA signature for a BMP2-induced osteoblast lineage commitment program. Proc Natl Acad Sci USA 2008; 105: 13906-13911.
- 61. Du M, Su XM, Zhang T, Xing YJ. Aberrant promoter DNA methylation inhibits bone morphogenetic protein 2 expression and contributes to drug resistance in breast cancer. Mol Med Rep 2014; 10: 1051-1055.
- Choi YJ, Ingram PN, Yang K, Coffman L, Iyengar M, Bai S, Thomas DG, Yoon E, Buckanovich RJ. Identifying an ovarian cancer cell hierarchy regulated by bone morphogenetic protein 2. Proc Natl Acad Sci USA 2015; 112: 6882-6888.
- 63. Eramo A, Ricci-Vitiani L, Zeuner A, Pallini R, Lotti F, Sette G, Pilozzi E, Larocca LM, Peschle C, De Maria R. Chemotherapy resistance of glioblastoma stem cells. Cell Death Differ 2006; 13: 1238-1241.
- 64. Murakami N, Saito N, Horiuchi H, Okada T, Nozaki K, Takaoka K. Repair of segmental defects in rabbit humeri with titanium fiber mesh cylinders containing recombinant human bone morphogenetic protein-2 (rhBMP-2) and a synthetic polymer. J Biomed Mater Res 2002; 62: 169-174.
- 65. Chang SC, Chuang HL, Chen YR, Chen JK, Chung HY, Lu YL, Lin HY, Tai CL, Lou J. Ex vivo gene therapy in autologous bone marrow stromal stem cells for tissueengineered maxillofacial bone regeneration. Gene Ther 2003; 10: 2013-2019.
- Turgeman G, Zilberman Y, Zhou S, Kelly P, Moutsatsos IK, Kharode YP, Borella LE, Bex FJ, Komm BS, Bodine PV, Gazit D. Systemically administered rhBMP-2 promotes MSC activity and reverses bone and cartilage loss in osteopenic mice. J Cell Biochem 2002; 86: 461-474.
- Matsuo T, Sugita T, Kubo T, Yasunaga Y, Ochi M, Murakami T. Injectable magnetic liposomes as a novel carrier of recombinant human BMP-2 for bone formation in a rat bone-defect model. J Biomed Mater Res A 2003; 66: 747-754.
- Li RH, Bouxsein ML, Blake CA, DAugusta D, Kim H, Li XJ, Wozney JM, Seeherman HJ. rhBMP-2 injected in a calcium phosphate paste (alpha-BSM) accelerates healing in the rabbit ulnar osteotomy model. J Orthop Res 2003; 21: 997-1004.

- 69. Lykissas M, Gkiatas I. Use of recombinant human bone morphogenetic protein-2 in spine surgery. World J Orthop 2017; 8: 531-535.
- Deyo RA, Ching A, Matsen L, Martin BI, Kreuter W, Jarvik JG, Angier H, Mirza SK. Use of bone morphogenetic proteins in spinal fusion surgery for older adults with lumbar stenosis: trends, complications, repeat surgery, and charges. Spine (Phila Pa 1976) 2012; 37: 222-230.
- Gupta A, Kukkar N, Sharif K, Main BJ, Albers CE, El-Amin Iii SF. Bone graft substitutes for spine fusion: A brief review. World J Orthop 2015; 6: 449-456.
- 72. Sandhu HS. Bone morphogenetic proteins and spinal surgery. Spine (Phila Pa 1976). 2003; 28: 64-73.
- 73. Cochran DL, Wozney JM. Biological mediators for periodontal regeneration. Periodontol 1999; 19: 40-58.
- 74. Gilbert SF. The morphogenesis of evolutionary developmental biology. Int J Dev Biol 2003; 47: 467-477.
- 75. Jung T, Lee JH, Park S, Kim YJ, Seo J, Shim HE, Kim KS, Jang HS, Chung HM, Oh SG, Moon SH, Kang SW. Effect of BMP-2 delivery mode on osteogenic differentiation of stem cells. Stem Cells Int 2017; 2017: 7859184.
- 76. Park SH, Kwon JS, Lee BS, Park JH, Lee BK, Yun JH, Lee BY, Kim JH, Min BH, Yoo TH, Kim MS. BMP2modified injectable hydrogel for osteogenic differentiation of human periodontal ligament stem cells. Sci Rep 2017; 7: 6603.
- 77. Yang J, Ye L, Hui TQ, Yang DM, Huang DM, Zhou XD, Mao JJ, Wang CL. Bone morphogenetic protein 2-induced human dental pulp cell differentiation involves p38 mitogen-activated protein kinase-activated canonical WNT pathway. Int J Oral Sci 2015; 7: 95-102.
- Pera MF, Andrade J, Houssami S, Reubinoff B, Trounson A, Stanley EG, Ward-van Oostwaard D, Mummery C. Regulation of human embryonic stem cell differentiation by BMP-2 and its antagonist noggin. J Cell Sci 2004; 117: 1269-1280.
- 79. Morrison SJ, Qian D, Jerabek L, Thiel BA, Park IK, Ford PS, Kiel MJ, Schork NJ, Weissman IL, Clarke MF. A genetic determinant that specifically regulates the frequency of hematopoietic stem cells. J Immunol 2002; 168: 635-642.
- Singer JW, Fialkow PJ. The use of genetic markers in studies of myeloid neoplasia. Leuk Lymphoma 1990; 3: 165-172.
- Fialkow PJ. Stem cell origin of human myeloid blood cell neoplasms. Verh Dtsch Ges Pathol 1990; 74: 43-47.
- 82. Lee KS, Kim EY, Jeon K, Cho SG, Han YJ, Yang BC, Lee SS, Ko MS, Riu KJ, Lee HT, Park SP. 3,4-Dihydroxyflavone acts as an antioxidant and antiapoptotic agent to support bovine embryo development in vitro. J Reprod Dev 2011; 57: 127-134.
- Lin W, Srajer G, Evrard YA, Phan HM, Furuta Y, Dent SY. Developmental potential of Gcn5(-/-) embryonic stem cells in vivo and in vitro. Dev Dyn 2007; 236: 1547-1557.

- 84. Mukai K, Matsuoka K, Taya C, Suzuki H, Yokozeki H, Nishioka K, Hirokawa K, Etori M, Yamashita M, Kubota T, Minegishi Y, Yonekawa H, Karasuyama H. Basophils play a critical role in the development of IgE-mediated chronic allergic inflammation independently of T cells and mast cells. Immunity 2005; 23: 191-202.
- Serrano L, Liang L, Chang Y, Deng L, Maulion C, Nguyen S, Tischfield JA. Homologous recombination conserves DNA sequence integrity throughout the cell cycle in embryonic stem cells. Stem Cells Dev 2011; 20: 363-374.
- Cozzio A, Passegue E, Ayton PM, Karsunky H, Cleary ML, Weissman IL. Similar MLL-associated leukemias arising from self-renewing stem cells and short-lived myeloid progenitors. Genes Dev 2003; 17: 3029-3035.
- Lapidot T, Sirard C, Vormoor J, Murdoch B, Hoang T, Caceres-Cortes J, Minden M, Paterson B, Caligiuri MA, Dick JE. A cell initiating human acute myeloid leukaemia after transplantation into SCID mice. Nature 1994; 367: 645-648.
- 88. Bonnet D, Dick JE. Human acute myeloid leukemia is organized as a hierarchy that originates from a primitive hematopoietic cell. Nat Med 1997; 3: 730-737.
- Uckun FM, Sather H, Reaman G, Shuster J, Land V, Trigg M, Gunther R, Chelstrom L, Bleyer A, Gaynon P, Crist W. Leukemic cell growth in SCID mice as a predictor of relapse in high-risk B-lineage acute lymphoblastic leukemia. Blood 1995; 85: 873-878.
- Al-Hajj M, Wicha MS, Benito-Hernandez A, Morrison SJ, Clarke MF. Prospective identification of tumorigenic breast cancer cells. Proc Natl Acad Sci USA 2003; 100: 3983-3988.
- 91. Dalerba P, Dylla SJ, Park IK, Liu R, Wang X, Cho RW, Hoey T, Gurney A, Huang EH, Simeone DM, Shelton AA, Parmiani G, Castelli C, Clarke MF. Phenotypic characterization of human colorectal cancer stem cells. Proc Natl Acad Sci USA 2007; 104: 10158-10163.
- 92. OBrien CA, Pollett A, Gallinger S, Dick JE. A human colon cancer cell capable of initiating tumour growth in immunodeficient mice. Nature 2007; 445: 106-110.
- Ricci-Vitiani L, Lombardi DG, Pilozzi E, Biffoni M, Todaro M, Peschle C, De Maria R. Identification and expansion of human colon-cancer-initiating cells. Nature 2007; 445: 111-115.
- 94. Singh SK, Hawkins C, Clarke ID, Squire JA, Bayani J, Hide T, Henkelman RM, Cusimano MD, Dirks PB. Identification of human brain tumour initiating cells. Nature 2004; 432: 396-401.
- 95. Chen J, Li Y, Yu TS, McKay RM, Burns DK, Kernie SG, Parada LF. A restricted cell population propagates

glioblastoma growth after chemotherapy. Nature 2012; 488: 522-526.

- 96. Pascual G, Avgustinova A, Mejetta S, Martin M, Castellanos A, Attolini CS, Berenguer A, Prats N, Toll A, Hueto JA, Bescos C, Di Croce L, Benitah SA. Targeting metastasis-initiating cells through the fatty acid receptor CD36. Nature 2017; 541: 41-45.
- 97. Kahn M. Can we safely target the WNT pathway? Nat Rev Drug Discov 2014; 13: 513-532.
- 98. Takebe N, Miele L, Harris PJ, Jeong W, Bando H, Kahn M, Yang SX, Ivy SP. Targeting Notch, Hedgehog, and Wnt pathways in cancer stem cells: clinical update. Nat Rev Clin Oncol 2015; 12: 445-464.
- 99. Batlle E, Clevers H. Cancer stem cells revisited. Nat Med 2017; 23: 1124-1134.
- 100. Bhola NE, Balko JM, Dugger TC, Kuba MG, Sanchez V, Sanders M, Stanford J, Cook RS, Arteaga CL. TGF-beta inhibition enhances chemotherapy action against triplenegative breast cancer. J Clin Invest 2013; 123: 1348-1358.
- 101. Naka K, Hoshii T, Muraguchi T, Tadokoro Y, Ooshio T, Kondo Y, Nakao S, Motoyama N, Hirao A. TGF-beta-FOXO signalling maintains leukaemia-initiating cells in chronic myeloid leukaemia. Nature 2010; 463: 676-680.
- 102. Persano L, Pistollato F, Rampazzo E, Della Puppa A, Abbadi S, Frasson C, Volpin F, Indraccolo S, Scienza R, Basso G. BMP2 sensitizes glioblastoma stem-like cells to Temozolomide by affecting HIF-1alpha stability and MGMT expression. Cell Death Dis 2012; 3: 412.
- 103. Huang P, Chen A, He W, Li Z, Zhang G, Liu Z, Liu G, Liu X, He S, Xiao G, Huang F, Stenvang J, Brunner N, Hong A, Wang J. BMP-2 induces EMT and breast cancer stemness through Rb and CD44. Cell Death Discov 2017; 3: 17039.
- 104. Wang L, Park P, Zhang H, La Marca F, Claeson A, Valdivia J, Lin CY. BMP-2 inhibits the tumorigenicity of cancer stem cells in human osteosarcoma OS99-1 cell line. Cancer Biol Ther 2011; 11: 457-463.
- 105. Liu J, Chen W, Zhao Z, Xu HH. Reprogramming of mesenchymal stem cells derived from iPSCs seeded on biofunctionalized calcium phosphate scaffold for bone engineering. Biomaterials 2013; 34: 7862-7872.

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