

Isolation of important cultivable bacteria from the moth flies (*Clogmia albipunctata*) as a mechanical vector in Babol hospitals, north of Iran.

Mohsen Karami¹, Abazar Pournajaf², Ramazan Rajabnia², Saeid Mahdavi Omran¹, Ali Heidarpour³, Jaber Alipour¹, Aynaz Khademian², and Mojtaba Taghizadeh Armaki^{*1}

¹Department of Parasitology and Mycology, School of Medicine, Infectious Diseases and Tropical Medicine Research Center, Health Research Center, Babol University of Medical Sciences, Babol, Iran

²Department of Microbiology, School of Medicine, Infectious Diseases and Tropical Medicine Research Center, Health Research Center, Babol University of Medical Sciences, Babol, Iran

³Health Center, Babol University of Medical Sciences, Babol, Iran

Abstract

Insects have been defined as mechanical vectors of healthcare-associated infections. The study has been directed with the aim of isolation of *C. albipunctata* to transmit and transport bacterial pathogens in four teaching hospitals Babol, Iran. In this cross-sectional study, 274 adult moth flies were collected from four teaching therapeutic centers in a 5 month period. After identification of moth flies at species level using taxonomic keys, bacterial strains were detected from internal, and external body samples using microbiological and biochemical standard tests. Of 137 adult flies, 85.4% and 78.1% bacterial isolates were obtained from the gastrointestinal tract and cuticular surface, respectively. Opportunistic aerobic mesophilic *Bacillus* species, and Coagulase Negative *Staphylococci* (CoNS) were the most strains isolated, respectively. The existence of moth flies is an actual threat to human health since they can carry numerous pathogenic bacteria, which can cause a wide variety of infections.

Keywords: Cultivable bacteria, *Clogmia albipunctata*, Moth fly, Mechanical vector.

Accepted on 17 April, 2020

Introduction

Vector-borne diseases are transmitted through Arthropoda including ticks, flies, lice, bugs and mosquitoes. Insects are classified as mechanical vectors such as flies which carry infectious agents on the surface of their bodies [1]. Mechanical transmission occurs when the agents do not replicate or develop in/on the vector; it is simply transported by the vector from a host to host [2]. A Synanthropic flies, *Clogmia albipunctata* (Diptera: Psychodidae), are responsible for the mechanical transmission of a wide variety of pathogenic agents in hospitals and health facilities [3]. There is evidence that flies carry a large number of pathogens which responsible for serious infections in both humans and animals [4]. More than 130 pathogens, mainly bacteria (counting some serious and life-threatening spp) were recognized from the flies [5-8].

In various parts of the hospital, such as toilets, kitchens, emergency wards, patients' rooms, and waste disposal sites, flies are active [4]. These flies, which are considered as mechanical vectors, can transfer the infectious agents from different parts to others and patients [7]. The prevalence of these flies in hospitals depends on a variety of conditions, such as environmental condition, public health and its biology [9]. Nosocomial infections are a main safety concern for both patients and the health service providers [10]. Considering mortality, morbidity, increased length of hospitalization and the charge, efforts should be made to make the hospitals as safe as

likely by preventing such illnesses [11]. So, the aim of this study was determine of medically important cultivable bacteria isolated from moth flies (*C. albipunctata*) from four teaching hospitals, Babol, Iran.

Materials and methods

Sampling

From May 2016 to September 2017, a total of 137 adult moth flies was caught straightly from their resting places, without close contacting, using a sterilized tubes. All flies were collected from four hospital environments (A-D) and transported animated and at 25 °C to the entomology laboratories of Babol university of medical sciences for further processing. The moth flies were recognized at species level using taxonomic keys [12,13].

Bacterial testing

Each sample was placed distinctly in a sterile tube and anesthetized using freezing at 0°C for 5 minutes. After adding 2 ml of phosphate-buffered saline (PBS; pH=7.0), vortexed for 20 s to wash down bacteria from the external surface. In order to identification of the bacterial isolates in the gastrointestinal tract (GI), all *C. albipunctata* were placed in a sterile tube containing 70% ethanol (for decontamination of cuticular surface) for 3 min, and allowed to dry at room temperature

under air condition. The flies were washed to remove alcohol molecules with PBS.

The GI of the adult moth flies was immersed in PBS and centrifuged at $\times 2000$ rpm for 5 min. For identification of the bacterial isolates, 0.1 ml of the above mentioned solutions was picked and transferred to the blood agar containing 5% sheep blood and McConkey II agar plates. All petri dishes were incubated at 37°C for 24-48h. The bacterial isolates were identified using standard microbiological and biochemical

methods. The approved strains were grown in Trypticase soy broth containing 0.3% yeast extract (TSBY) and stored at -70°C after addition of 15% glycerol (Vol/Vol).

Results

In the cross-sectional study, of 274 provided samples from 137 adult flies, 85.4% (n=117) and 78.1% (n=107) bacterial isolates were obtained from the GI and cuticular surface, respectively (Table 1).

Table 1. Bacterial isolated from *C. albipunctata* in present study.

Bacterial isolates	The body of the fly		
	GI	External	Total
Opportunistic aerobic mesophilic <i>Bacillus</i> spp.	34 (24.82)	44 (32.12)	78 (28.47)
Coagulase Negative <i>Staphylococci</i> (CoNS)	29 (21.17)	25 (18.25)	54 (19.71)
<i>Streptococcus</i> spp.	2 (1.46)	5 (3.65)	7 (2.55)
<i>Escherichia coli</i>	3 (2.19)	10 (7.3)	13 (4.74)
<i>Enterobacter cloacae</i>	8 (5.84)	0 (0.00)	8 (2.92)
Non-typhoid <i>Salmonella</i>	5 (3.65)	3 (2.19)	8 (2.92)
<i>Pseudomonas aeruginosa</i>	6 (4.38)	1 (0.73)	7 (2.55)
<i>Neisseria</i> spp.	0 (0.00)	1 (0.73)	1 (0.36)
<i>Enterococcus</i>	0 (0.00)	6 (4.38)	6 (2.19)
Diphtheroids	0 (0.00)	4 (2.92)	4 (1.46)
<i>Klebsiella pneumoniae</i>	0 (0.00)	3 (2.19)	3 (1.09)
<i>Pseudomonas</i> spp.	3 (2.19)	0 (0.00)	3 (1.09)
<i>Proteus</i> spp.	3 (2.19)	0 (0.00)	3 (1.09)
Citrobacter	3 (2.19)	0 (0.00)	3 (1.09)
<i>Corynebacterium</i> spp.	0 (0.00)	3 (2.19)	3 (1.09)
<i>Staphylococcus aureus</i>	1 (0.73)	1 (0.73)	2 (0.73)
<i>Micrococcus</i>	0 (0.00)	1 (0.73)	1 (0.36)
<i>Acinetobacter baumannii</i>	0 (0.00)	1 (0.73)	1 (0.36)
<i>Acinetobacter Iwoffii</i>	0 (0.00)	1 (0.73)	1 (0.36)
<i>Serratia marcescens</i>	0 (0.00)	1 (0.73)	1 (0.36)
<i>Shigella</i> spp.	1 (0.73)	1 (0.73)	2 (0.73)
<i>Providencia</i> spp.	1 (0.73)	0 (0.00)	1 (0.36)
Negative	38 (27.74)	26 (18.98)	64 (23.36)
Total	137 (100)	137 (100)	274 (100)

A total of 81.8% of the caught flies were positive for bacterial infestation, so that in all hospitals under study, at least 80% of the flies were carriers of bacteria. The prevalence of the isolated bacteria from the GI were 86.4%, 86.3%, 85.2%, 80%, in the D, A, B and C hospitals, respectively. So, 95%, 77%, 76.7% and 73.8% of bacterial strains were obtained from the

cuticular surface (external body) in C, D, B and A hospitals, respectively.

Discussion and Conclusion

The global distribution of the moth fly has been described to be between latitudes 40°S and 42°N [9,14]. In recent years, due to climate changes, including warming global temperatures,

Citation: Karami/Pournajaf/Rajabnia/et al.. Isolation of important cultivable bacteria from the moth flies (*Clogmia albipunctata*) as a mechanical vector in Babol hospitals, north of Iran. *Allied J Med Res* 2020;4(1):40-43.

reports have been made of the presence of these flies in higher latitudes. Few studies have been conducted on the biology and ecology of this fly in Iran, but observations show a lot of infestation with the insect, especially in the warm seasons. Moth flies are commonly observed in hospital settings, such as shower cubicles, patient wards, rest rooms, hospital kitchens, toilet and laboratory rooms [4,9,14]. Faulde showed that moth flies are a potential mechanical vector of bacterial pathogens related to the healthcare-associated infections (HAIs) [4]. The bacteria carried by these flies can cause a wide range of infections, especially in immune compromised patients [15].

On the other hand, the contamination of surfaces and medical devices with insect-borne bacteria can lead to the spread and transmission of infectious agents, the dissemination of resistance factors between different strains (interspecies and intraspecies transmission) and the mobile carrier for potentially pathogenic microbes especially bacteria known to cause HAIs [16,17].

A list of microbes has been recognized by the Infectious Disease Society of America (IDSA) as being responsible for the majority of nosocomial associated drug resistant infections [18,19]. These organisms are the members of the notorious "ESKAPE" (*Enterococcus faecium*, *Staphylococcus aureus*, *Klebsiella pneumoniae*, *Acinetobacter baumannii*, *Pseudomonas aeruginosa*, and *Enterobacter* species) [20].

In the present study, *A. baumannii*, *E. coli*, *K. pneumoniae*, *P. aeruginosa*, *Enterobacter cloacae*, *Enterococcus spp.* and *S. aureus*, which were considered as a cause for HAIs, were isolated. In our study, of 274 prepared samples, 2.92% non-typhoid Salmonella (NTS) was detected. Foodborne illness caused by NTS is found to be a main public health concern worldwide. Intestinal salmonellosis is self-limiting; though, it may lead to systemic signs in children, the elderly and immunocompromised patients. Contamination of kitchens, food preparation and distribution centers with insects can help to spread Foodborne disease. So, *Shigella* is a significant cause of acute diarrhoeal illness.

Worldwide, there are approximately 164.7 million patients annually, and 1.1 million people are estimated to die from bacillary dysentery (BD). There are several ways of contracting shigellosis including (1) contaminated water, (2) contaminated foods and (3) close person-to-person contact. The pathogens can be transferred by flies, feces, fingers, fomites and food. Control of insects as a mechanical vector is plays an important role in Decreasing *Shigella*-related infections [21]. In agreement with Motazedian *C. albipunctata* is a potential mechanical vector of microbial pathogens related to HAIs [19]. The presence of *C. albipunctata* is a real hazard for human health because they can carry several pathogenic bacteria, which can cause a wide range of infections. Elimination of moth flies from our hospitals and health facilities is essential. Though, bacterial surveys of colonized *C. albipunctata* may be regarded as elegant choices for checking sources of nosocomial pathogens and contaminated healthcare environments that would then be problematic to analyze.

Acknowledgement

We would like to thank hospital personnel at all four centers for their collaboration. This study was financially supported by Vice-Chancellor for Research and Technology of Babol University of Medical Sciences No. 9542832.

References

1. Sarwar M. Insect vectors involving in mechanical transmission of human pathogens for serious diseases. *Int J Bioinform Biomed Eng.* 2015;1:300-306.
2. Foil L.D, Gorham J.R. Mechanical transmission of disease agents by arthropods. *Med Entomol.* 2000;461-514.
3. García-Solache M, Jaeger J, Akam M. A systematic analysis of the gap gene system in the moth midge *Clogmia albipunctata*. *Dev Biol.* 2010;344:306-318.
4. Faulde M, Spiesberger M. Role of the moth fly *Clogmia albipunctata* (Diptera: Psychodinae) as a mechanical vector of bacterial pathogens in German hospitals. *J Hosp Infect.* 2013;83:51-60.
5. Kassiri H, Zarrin M, Veys-Behbahani R, et al. Isolation and identification of pathogenic filamentous fungi and yeasts from adult house fly (Diptera: Muscidae) captured from the hospital environments in Ahvaz City, southwestern Iran. *J Med Entomol.* 2015;52:1351-1356.
6. Khamesipour F, Lankarani KB, Honarvar B, et al. A systematic review of human pathogens carried by the housefly (*Musca domestica* L.). *BMC.* 2018;18:1049.
7. Kassiri H, K. Akbarzadeh, and A. Ghaderi, Isolation of pathogenic bacteria on the house fly, *Musca domestica* L. (Diptera: Muscidae), body surface in Ahwaz hospitals, Southwestern Iran. *Asian Pac J Trop Biomed.* 2012;12:1116-1119.
8. Barin A, Arabkhazaeli F, Rahbari S, et al. The housefly, *Musca domestica*, as a possible mechanical vector of Newcastle disease virus in the laboratory and field. *Med Vet Entomol.* 2010;24:88-90.
9. Faulde M, Spiesberger M, Hospital infestations by the moth fly, *Clogmia albipunctata* (Diptera: Psychodinae), in Germany. *J Hos Inf.* 2012;81:134-136.
10. Wiener-Well Y, Galuty M, Rudensky B, et al. Nursing and physician attire as possible source of nosocomial infections. *American J Infec Cont.* 2011;39:555-559.
11. Mehta Y, Gupta A, Todi S, et al. Guidelines for prevention of hospital acquired infections. *Indian J Crit Care Med.* 2014;18:149-163.
12. Boumans L, Zimmer JY, Verheggen F. First record of the bathroom mothmidge *Clogmia albipunctata*, a conspicuous element of the Belgian fauna that went unnoticed (Diptera: Psychodidae). *Phegea.* 2009;37:153-160.
13. Kvitte G.M. Biodiversity studies in Afrotropical moth flies (Diptera: Psychodidae). *The Uni Ber.* 2011.
14. Kvitte G.M, Ivković M, Klaric A, New records of moth flies (Diptera: Psychodidae) from Croatia, with the

- description of *Berdeniella keroveci* sp. nov. *Zootaxa*. 2013;3737:57-67.
15. Forster M, Sievert K, Messler S, et al. Comprehensive study on the occurrence and distribution of pathogenic microorganisms carried by synanthropic flies caught at different rural locations in Germany. *J Med Entomol*. 2009;46:164-1166.
 16. Mullen GR, Durden LA, Medical and veterinary entomology. Academic press. 2009.
 17. Fotedar R, Banerjee U, Samantray JC, et al. Vector potential of hospital houseflies with special reference to *Klebsiella* species. *Epidemiol Infect*. 1992;109:143-147.
 18. Spellberg B, Guidos R, Gilbert D, et al. The epidemic of antibiotic-resistant infections: a call to action for the medical community from the Infectious Diseases Society of America. *Clin Infect Dis*. 2008;46:155-164.
 19. Kalantari M, Motazedian MH, Qasem Asgari, et al. Bionomics of phlebotomine sand flies species (Diptera: Psychodidae) and their natural infection with *Leishmania* and *Crithidia* in Fars province, southern Iran. *J Para Dis*. 2018;42:511-518.
 20. Boucher HW, Talbot GH, Bradley JS, et al. Bad bugs, no drugs: no ESKAPE! An update from the Infectious Diseases Society of America. *Clin Infect Dis*. 2009;48:1-12.
 21. Tajbakhsh M, Migura LG, Rahbar M, et al. Antimicrobial-resistant *Shigella* infections from Iran: an overlooked problem. *J Antimicro chemo*. 2012;67:1128-1133.

***Correspondence to**

Mojtaba Taghizadeh Armaki

Department of Parasitology and Mycology

School of Medicine, Babol University of Medical Sciences

Babol

Iran

E-mail: mojtabataghizade@yahoo.com