Impact of electromagnetic irradiation produced by 3G mobile phone on brain neurotransmitters in mice during growth and development period.

Fengming Li¹, Jin Chang^{2*}, Yinggang Lv³, Dianguo Xu⁴, Jianhua Chen¹, Xuewen Sun¹

Abstract

The aims of this study were to detect the levels of brain neurotransmitters in mice exposed to electromagnetic irradiation produced by 3G mobile phone (3G-EMI). The mice in the growth and development period were randomly divided into three groups: the control group (C), the low-radiation group (L), and the high-radiation group (H). Group L was irradiated 30 min × 3 times per day, and group H was irradiated 60 min × 3 times per day. The contents of Acetylcholine (Ach), Dopamine (DA), and serotonin (5-HT) in the brain were then detected by liquid chromatograph-mass spectrometer (LC-MS/MS). The contents of ach in the brain neurotransmitters of experimental groups were different, but the differences were not significant (P>0.05). The content of DA in brain of high radiation group (29.170 \pm 2.469 ng/g) was significantly lower than that of low radiation group (34.487 \pm 2.652 ng/g) (P<0.05). The Ach contents in all the experimental groups varied, but the differences were not statistical (P>0.05). The DA content in the high-radiation group (29.170 \pm 2.469 ng/g) was statistically lower than the low-radiation group (34.487 \pm 2.652 ng/g) (P<0.05). The content of 5-HT in group L (1.6648 \pm 0.8136 ng/g) was statistically lower than group C (3.4518 \pm 0.9930 ng/g) (P<0.05), while the content of 5-HT in group H (3.9240 \pm 1.2523 ng/g) was statistically higher than group L (1.6648 \pm 0.8136 ng/g) (P<0.05). 3G-EMI has certain impact on the contents of brain neurotransmitters during growth and development period.

Keywords: Mobile phone radiation, Mice in the growth and development period, Acetylcholine, Dopamine, Serotonin. *Accepted on June 08, 2017*

Introduction

As an important means of information exchange, mobile phones have more and more penetrated into people's lives nowadays. Currently, the most widely used work frequency of mobile phones is GSM 900 MHz (Global System for Mobile Communications), the electromagnetic radiation pollution of which brings harm on human health and even leads to major public health problems, so mobile phone radiation has aroused people's attention [1-3]. Scholars all over the world have carried out a large number of epidemiological investigations and experimental studies toward the impact of mobile phone radiation on biological effects, but the results are not consistent [4-6]. In recent years, the rate of mobile phone usage in children and adolescents shows an increasing trend, while whether electromagnetic interference (EMI) may cause potential harm on the growth and development of children and adolescents is rarely investigated [7]. In 2010, World Health Organization (WHO) organized seminars and emphasized the

impact of EMI on children and adolescents as one preferred project [8]. However, relevant studies were conducted among epidemiological surveys, focusing on the happiness, cognition, attention, or behavioural problems [9,10]. Certain study once reported that often using mobile phones can greatly increase the adjusting dominance ratio of headaches, migraines, or pruritus in children [11]. However, the potential harms of mobile phone radiation toward the growth and development of children and adolescents still lack clinical experiments, so it needs further studies.

The brain tissue is the closest part to the working mobile phone, so the impact of mobile phone radiation on the brain tissue has more and more become a hot spot. Choline's and monoamine neurotransmitters are important neurotransmitters secreted by the brain, and they can transmit various kinds of information so as to achieve the brain's function of regulating the body's physiological function. This study used young mice as the subjects, and detected the contents of Acetylcholine

¹Teaching and Researching Department of Pathogenic Biology, Medical College of Hebei University of Engineering, Handan, PR China

²Department of Orthopaedics, the Affiliated Hospital of Hebei University of Engineering, Handan, PR China

³Department of Radiology, the Affiliated Hospital of Hebei University of Engineering, Handan, PR China

⁴Teaching and Researching Department of Anatomy, the Affiliated Hospital of Hebei University of Engineering, Handan, PR China

(Ach), Dopamine (DA), and 5-Hydroxytryptamine (5-HT) in the brain tissue of young mice exposed to EMI, and explore the mechanism of mobile phone radiation on the growth and development and physiological function of children and adolescents, hoping to investigate the impact of mobile phone radiation on the growth, development, and physiological functions in children and adolescents.

Materials and Methods

Animals

2-week-old C57BI/6J young mice (weighing 6-8 g, male: female=1:1, certificate No.5CXX (Jing) 2007-0003) were provided by Beijing Vital River Laboratory Animal Co. Ltd. This study was carried out in strict accordance with the recommendations in the Guide for the Care and Use of Laboratory Animals of the National Institutes of Health. The animal use protocol has been reviewed and approved by the Institutional Animal Care and Use Committee (IACUC) of Hebei University of Engineering. The distance between mobile phone and base station was not measured. The laboratory electric field intensity range is between $1.3 \times 10^{-4} \text{ W/m}^2 \sim 2.2 \times 10^{-4} \text{ W/m}^2$ 10-2 W/m² by the German Narda company's NBM-500 type electromagnetic field strength measuring instrument, which has met GB 8702-1988 "electromagnetic radiation protection regulations" requirements (less than 0.4 W/m², the threshold value for public irradiation).

Animal grouping and treatment

A total of 30 young mice were randomly divided into group C, group L, and group H, with 10 mice in each group. Each mouse was housed alone in one cylindrical cage with free access to diet and water (environmental temperature 25°C, humidity 40%~45%). During the experiment, we use an external partition board to restrict the activity space of the single mouse in the cylindrical rat cage. After the mobile phone signal is connected, the external suspension type is adopted to ensure that the antenna part of the mobile phone is close to the ear of the rat. The mobile phone (a certain brand of commercial mobile phone with the frequency as GSM 1800 MHz) was used to simulate the status of making phone calls. Meanwhile, the antenna of the mobile phone was ensured to be close to the ear of each mouse for the irradiation. Group L was irradiated 30 min \times 3 times a day (8:00, 12:00, and 20:00), while group H was irradiated 60 min × 3 times a day (8:00, 12:00, and 20:00). The three groups were treated in the same way, while the phone used in group C was not connected. After 4 weeks of radiation, all the rats were killed by ether, and the brain tissues were collected to detect the contents of Ach, DA, and 5-HT by LC-MS/MS.

High performance liquid chromatography (HPLC) conditions

Column: Syncronis HILIC, 150×4.6 mm, 5 µm; LC: Mobile phase: acetonitrile (A)-(0.01% formic acid+2 mM ammonium acetate) (B), gradient elution program: 68-35 B% in 2 min; 35

B% for 4 min; 35-68 B% in 2 min; flow rate: 800 μ L/min; oven temperature: 30°C.

MS conditions

One+ESI source was used with the following parameters: IS 5500V, pulverization temperature 650°C, pulverization gas pressure (GS1, N2) 60 psi, auxiliary gas pressure (GS2, N2) 60 psi, air curtain pressure (N2) 25 psi, and scanning mode MRM.

Preparation of reference solution and internal standard solution

Certain amounts of Ach standard (Alfa Aesar Chemical Co., Ltd., Tianjin, China), DA standard (Alfa Aesar Chemical Co., Ltd., Tianjin, China), and 5-HT standard (Sigma, Shanghai, China) were dissolved using 50% acetonitrile (Dima Technology Co., Ltd., Beijing, China) and shaken evenly to prepared the standard stock solutions, respectively. Certain amounts of the above stock solutions measured precisely, prepared the series standard solution of DAQ/5-HT using 50% acetonitrile, and stored at 4°C. The Glu-Glu standard (Sigma, Shanghai, China) was dissolved in 50% acetonitrile to prepared the Glu-Glu stock solution, which was then precisely measured certain amount and diluted with 50% acetonitrile (containing 0.1% formic acid) to prepare the internal standard solution, which was then stored at 4°C for future use.

Sample preparation

Each brain tissue sample was weighed, and homogenized in 0.1% formic acid solution (v/w=5 mL/g). After 5 min ultrasound on ice and 10 min centrifugation at 14000 r/min and 4°C (LG16-B, Beijing Lab Centrifuge Co., Ltd.), the supernatant was placed into one sample vial, freeze-dried, and stored at -20°C. The lyophilized sample was then added 20 μL of internal standard, 20 μL of 50% acetonitrile (containing 0.1% formic acid), and 50 μL of 50% acetonitrile (containing 0.1% formic acid), and vortexed for 2 min (XW80A, Shanghai Medical Instrument Factory). After 6 min centrifugation (14000 rpm), the supernatant was transferred into an auto sampler vial, and 50 μL of the sample was injected into one LC-MS/MS for the analysis.

Statistical analysis

SPSS12.0 software was used for the analysis of variance; the data were expressed as $\bar{x} \pm s$, and the pairwise comparison used the q-test, with P<0.05 considered as statistical significance.

Results

Methodology of LC-MS/MS

The detection of the contents of Ach, DA, and 5-HT in the brain tissue of the mice exhibited good accuracy and precision under the experimental conditions. The MS peaks of each neurotransmitter were intact without tailing and should-peak (Figure 1).

Impact of electromagnetic irradiation produced by 3G mobile phone on brain neurotransmitters in mice during growth and development period

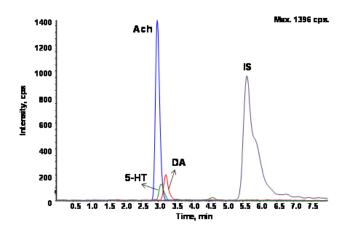


Figure 1. Typical MS peaks.

Standard curves

20 μ L of internal standard, 20 μ L of standard solutions, and 50 μ L of 50% acetonitrile (containing 0.1% formic acid) were added into centrifuge tubes to prepare the standard curves referring to the procedures in "1.6". The Analyst 1.5.2 data system (AB, USA) was used for the analysis with the concentration of the analyte as the abscissa (x) and the ratio of peak areas of the standards to the internal reference as the ordinate (y). The typical linear regression equations of the standards were (weight: $1/(x \times x)$): Ach: y=0.207x+0.0392 (r=0.9987); DA: y=0.00553x+0.0116; 5-HT: y=0.00397x+0.0054 (r=0.0059). The results showed that the three neurotransmitters exhibited good linear relationship.

Impact of EMI on brain neurotransmitters

As shown in Table 1, the DA content in group H is lower than group C, and the difference is statistically significant (P<0.05). The 5-HT content in group L is statistically lower than group C (P<0.05), but the 5-HT content in group H is statistically higher than group L (P<0.05). The contents of Ach among different groups are different, but the differences are not significant (P>0.05).

Table 1. Impact of EMI on brain neurotransmitters in mice during growth and development period.

Group	Ach (ng/g)	DA (ng/g)	5-HT (ng/g)
С	2.2238 ± 1.1312	34.487 ± 2.652	3.4518 ± 0.9930
L	2.6878 ± 0.6122	30.047 ± 1.920	1.6648 ± 0.8136 ^a
Н	1.3235 ± 0.6468	29.170 ± 2.469 ^a	3.9240 ± 1.2523 ^b

Note: aCompared with group C, P<0.05; bCompared with group L, P<0.05.

It can be seen from Table 1 that the content of the brain neurotransmitter DA in the high-radiation group is statistically lower than the control group (P<0.05). The 5-HT content in the low-radiation group is statistically lower than the control group (P<0.05), and the 5-HT content in the high-radiation group is statistically higher than the low-radiation group (P<0.05). Compared with the control group, the Ach content in the low-

radiation group is increased while decreased in the high-radiation group, but the differences are not statistically significant (P>0.05).

Discussion

Mobile phones emit electric waves when in communication, so the electromagnetic field generated around increases the electromagnetic pollution, if these waves are propagated in vivo in the right direction, they can be totally reflected and form standing waves, which is the maximum energy absorption in vivo and can cause certain biological effects, thus having impact on human health [12,13]. When persons are making phone calls, the brain is the body tissue nearest to the mobile phone, so the impact of mobile phone radiation on the brain tissue has become the focus of attention [14]. Cells, especially the brain cells, in children and adolescents are in the productive development period, and childhood and adolescent stages are important growth and development periods. Some scholars believe that the brain tissue in children has more adsorbent while thinner skull and relatively smaller proportion, so children may absorb EMI much more easily than adults [15].

In the present study, the contents of Ach, DA, and 5-HT in the brain tissue of mice were measured so as to explore the impact of mobile phone radiation on brain neurotransmitters in the growth and development period. The results indicated that EMI can cause metabolic disorders of DA and 5-HT in the brain tissue. Aboul Ezz et al. reported that the monoamine neurotransmitters in adult mice exposed to EMI can be interfered [16]. Ji et al. reported in 2011 that exposing to highintensity EMI can lead to neurotransmitter metabolic disorders in foetal rats [17]. Eris et al. showed that short-term EMI can increase the content of blood 5-HT, which can lead to learning slowness and spatial memory deficiency [18]. The results of this study suggested that there was no statistical significance in the Ach content among different experimental groups; however, certain studies also reported that 800 MHZ electromagnetic radiation can increase the release of Ach in mice, the different results of which may be caused by different experimental animals or experimental methods [19].

The brain neurotransmitters are the molecular mechanism that can impact the functions of the central nervous system, mainly including choline's and monoamines. The central cholinergic system is closely related to learning and memory, among which Ach is one important neurotransmitter in the central cholinergic system, mainly functioning in such cognitive activities as learning and memory, spatial working memory, attention maintenance, spontaneous movement, or exploring behaviours. Among the monoamine neurotransmitters, DA is the key neurotransmitter in the hypothalamus and pituitary gland. The DA-energy nervous system plays an important role in pleasure and behaviour motivation. 5-HT is mainly distributed in the pineal gland and hypothalamus, and its functional activity reduction is closely related to depressive mood, anorexia, sex disorder, or endocrine disorders in depression patients [20].

Neurotransmitters are synthetized by neurons and then transported into the synaptic vesicles in the pre-protruding cells, which are then converted by the Ca²⁺ channel and released at the nerve ends. After diffusing through the synaptic cleft, neurotransmitters act on the postsynaptic neurons or the receptors of effector cells, thus realizing the roles of transferring, amplifying, and regulating information among neurons. Hocking et al. considered that EMI may change the neurotransmitter system, including the neurotransmitter release and the receptor coagulation [21]. Mugunthan et al. reported that long-term exposing to 900-1800WHZ EMI can significantly reduce the density and diameter of the hippocampal neurons in foetal mice [22]. Anderson et al. reported that a small amount of EMI toward the embryonic mouse brain tissue can cause acute morphological changes in the mature brain tissue [23]. The glial cells can support and neutralize the neurons. Lu et al. reported that EMI can lead to inflammation in the microglia and astrocytes [24]. Other similarities also reported that EMI can result in the death of neurons and glial cells [25,26]. In our experiments, the mice received EMI in their growth and development period exhibited the content changes of DA and 5-HT in the brain tissue. The mechanism is not clear yet, maybe due to the disturbances of electromagnetic radiation of mobile phone to the weak and stability electromagnetic field of brain, which cause morphological or functional changes of the brain cells and lead to metabolic change. The content changes of DA and 5-HT in the brain tissue were caused by the injury of neurons and glial cells, or by the synthetic and metabolic imbalance of neurotransmitters still needs further experiments foe the verification.

In summary, EMI caused by mobile phones can cause metabolic disorders of brain neurotransmitters in mice during growth and development period. Therefore, we suggest that children and adolescents in the growth and development period should be more cautious about using mobile phones, and should not use it for long period.

Acknowledgement

This study was supported by the Projects of Hebei Health Department.

Conflict of Interest

The authors declare no conflict of interest.

References

- 1. Guo J, Wang XW, Sheng J, Tang JT. Biological effect of electromagnetic radiation on the nervous system. J Clin Rehab Tissue Eng Res 2009; 13: 5939-5942.
- 2. Kumar S, Nirala JP, Behari J, Paulraj R. Effects of electromagnetic irradiation produced by 3G mobile phone on male rat reproductive system in a simulated scenario. Indian J Exp Biol 2014; 52: 890-897.

- 3. Kesari KK, Siddiqui MH, Meena R, Verma HN, Kumar S. Cell phone radiation exposure on brain and associated biological systems. Indian J Exp Biol 2013; 51: 187-200.
- 4. Irmak MK, Oztas E, Yagmurca M, Fadillioglu E, Bakir B. Effects of electromagnetic radiation from a cellular telephone on epidermal Merkel cells. J Cutan Pathol 2003; 30: 135-138.
- 5. Tsurita G, Nagawa H, Ueno S, Watanabe S, Taki M. Biological and morphological effects on the brain after exposure of rats to a 1439MHZ TDMA field. Bioelectromagnetics 2000; 21: 364-371.
- Fritze K, Sommer C, Schmitz B, Mies G, Hossmann KA, Kiessling M, Wiessner C. Effect of global system for mobile communication (GSM) microwave exposure on blood-brain barrier permeability in rat. Acta Neuropathol 1997; 94: 465-470.
- 7. Feychtin M. Mobile phones, radiofrequency fields, and health effects in children-epidemiological studies. Prog Biophys Mol Biol 2011; 107: 343-348.
- 8. WHO Research Agenda for Radiofrequency Fields. World Health Organization 2010.
- 9. Heinrich S, Thomas S, Heumann C, von Kries R, Radon K. The impact of exposure to radio frequency electromagnetic fields on chronic well-being in young people-a cross-sectional study based on personal dosimetry. Environ Int 2011; 37: 26-30.
- 10. Feychting M. Mobile phones, radiofrequency fields, and health effects in children epidemiological studies. Prog Biophys Mol Biol 2011; 107: 343-348.
- 11. Chiu CT, Chang YH, Chen CC, Ko MC, Li CY. Mobile phone use and health symptoms in children. J Formos Med Assoc 2015; 114: 598-604.
- 12. Kayabasoglu G, Sezen OS, Eraslan G, Aydin E, Coskuner T, Unver S. Effect of chronic exposure to cellular telephone electromagnetic fields on hearing in rats. J Laryngol Otol 2011; 125: 348-353.
- 13. Dabholkar YG, Pusalkar AG, Velankar HK. Effects of cell phone EMF radiations on the auditory system-a review. IJHSR 2016; 6: 506-515.
- 14. Buckus R, Strukcinskiene B, Raistenskis J, Stukas R. Modelling and assessment of the electric field strength caused by mobile phone to the human head. Vojnosanit Pregl 2016; 73: 538-543.
- 15. Morgan LL, Kesari S, Davis DL. Why children absorb more microwave radiation than adults: consequences. J Micro Ultra 2014; 2: 197-204.
- 16. Aboul Ezz HS, Khadrawy YA, Ahmed NA, Radwan NM, El Bakry MM. The effect of pulsed electromagnetic radiation from mobile phone on the levels of monoamine neurotransmitters in four different areas of rat brain. Eur Rev Med Pharmacol Sci 2013; 17: 1782-1788.
- 17. Jing J, Yuhua Z, Xiao-qian Y, Rongping J, Dong-mei G, Xi C. The influence of microwave radiation from cellular phone on foetal rat brain. Electromagn Biol Med 2012; 31: 57-66.

Impact of electromagnetic irradiation produced by 3G mobile phone on brain neurotransmitters in mice during growth and development period

- 18. Eris AH, Kiziltan HS, Meral I, Genc H, Trabzon M, Seyithanoglu H, Yagci B, Uysal O. Effect of Short-term 900 MHz low level electromagnetic radiation exposure on blood serotonin and glutamate levels. Bratisl Lek Listy 2015; 116: 101-103.
- 19. Testylier G, Tonduli L, Malabiau R, Debouzy JC. Effects of exposure to low level radiofrequency fields on acetylcholine release in hippocampus of freely moving rats. Bioelectromagnetics 2002; 23: 249-255.
- 20. Pankova NB, Vetrilé LA, Basharova LA, Krupina NA, Khlebnikova NN, Rodina VI, Kryzhanovskii GN. Immunization of rats with conjugates of dopamine and serotonin with bovine serum albumin prevents the development of experimental MPTP-induced depressive syndrome (electrophysiological parameters). Neurosci Behav Physiol 2004; 34: 131-138.
- 21. Hocking B, Westerman R. Neurological effects of radiofrequency radiation. Occup Med 2003; 53: 123-127.
- 22. Mugunthan N, Shanmugasamy K, Anbalagan J, Rajanarayanan S, Meenachi S. Effects of long term exposure of 900-1800 MHz radiation emitted from 2G mobile phone on mice hippocampus- a Histomorphometric study. J Clin Diagn Res 2016; 10: 1-6.
- 23. Anderson RE, Berthrong M, Fajardo LF. Radiation injury. Anderson's Pathology. Von Hoffman Press 1996; 1: 484-512.

- 24. Lu Y, He M, Zhang Y, Xu S, Zhang L, He Y, Chen C, Liu C, Pi H, Yu Z, Zhou Z. Differential pro-inflammatory responses of astrocytes and microglia involve STAT3 activation in response to 1800 MHz radiofrequency fields. PLoS One 2014; 9: e108318.
- 25. Joubert V, Bourthoumieu S, Leveque P, Yardin C. Apoptosis is induced by radiofrequency fields through the caspase-independent mitochondrial pathway in cortical neurons. Radiat Res 2008; 169: 38-45.
- 26. Liu YX, Tai JL, Li GQ, Zhang ZW, Xue JH, Liu HS, Zhu H, Cheng JD, Liu YL, Li AM, Zhang Y. Exposure to 1950 MHz TD-SCDMA electromagnetic fields affects the apoptosis of astrocytes via caspase-3-dependent pathway. PLoS One 2012; 7: e42332.

*Correspondence to

Jin Chang

Department of Orthopaedics

The Affiliated Hospital of Hebei University of Engineering

Handan

PR China