

A Comparative study and evaluation of serum adenosine deaminase activity in the diagnosis of pulmonary tuberculosis

K. Srinivasa Rao*, **H. Anand Kumar****, **B.M.Rudresh*****, **T. Srinivas****, **K. Harish Bhat***

*Department of Biochemistry, Navodaya Medical College, Raichur, India.

**Department of Microbiology, Navodaya Medical College, Raichur, India.

***Department of Biochemistry, AIMS, B.G.Nagar, Bellur, India.

Abstract

The study was done to evaluate the ADA activity in the diagnosis of pulmonary tuberculosis and its relationship with clinical, radiological and laboratory parameters. The study was carried out on 142 patients suspected from various pulmonary disorders from July 2008 to January 2009. Forty five normal healthy individuals were included as control subjects for ADA evaluation. Four groups of patients namely A (0-20 yrs), B (21-40), C (41-60) and D (\geq 61 yrs) were formed and diagnosis was based on the x-ray Chest, Mantoux test, ESR, sputum AFB and Serum ADA level. The Sensitivity of various tests against ADA was done. In the diagnosis of Pulmonary tuberculosis, serum ADA showed high percent positivity of 88% followed by chest x-ray (76%), ESR (72%), sputum AFB (63%) and Mantoux Test (61%). Significantly high serum ADA level was reported in pulmonary tuberculosis cases compared to non tubercular pulmonary diseases. Sputum AFB negative pulmonary tuberculosis cases have also shown elevated level of serum ADA at par with sputum AFB positive cases. In the present study serum ADA level in control group was 15.3 ± 0.23 U/L and cut off value was 33 U/L for diagnosis of pulmonary tuberculosis.

As determination of ADA is not costly or time consuming, should be done routinely, particularly if the diagnosis of tuberculosis is in doubt, clinically suggestive but sputum AFB negative cases and also to differentiate pulmonary tuberculosis from non tubercular pulmonary diseases.

Key words: Serum ADA, Pulmonary Tuberculosis, Non tubercular pulmonary diseases

Accepted October 17 2009

Introduction

Tuberculosis (TB) remains the single largest infectious disease causing high mortality in humans, leading to 3 million deaths annually, about five deaths every minute. Approximately 8-10 million people are infected with *Mycobacterium tuberculosis* every year [1]. Out of the total number of cases, 40 percent of the cases are accommodated in South East Asia alone. In India there are about 500,000 deaths occurring annually due to TB [2], with the incidence and prevalence being 1.5 and 3.5 millions per year. A positive AFB smear and /or culture of *Mycobacterium* is the gold standard for the diagnosis of tuberculosis. Though smear positivity correlates well with infectivity, much of the transmission occurs before the level of bacilli reach 10^4 per ml in the sputum. Infection is a stochastic process and certainly occurs from paucibacillary

and smear negative cases which are more common in HIV seropositives [3].

Problem arises when sputum smear result is repeatedly negative for acid fast bacilli and to detect the TB bacilli microscopically, there should be at least 50,000 bacilli per ml of sputum. Culture for tubercle bacilli is a sophisticated and time consuming process. The TB bacilli grow slowly (generation time 14-15 hours) and culture becomes positive only in about 2 weeks and some times it may take up to 6-8 weeks[4]. Chest radiograph (Skiagram) provides only a probable diagnosis, they are some times difficult to differentiate from other cause of lung shadows such as pneumonia and malignancies[5]. To overcome these difficulties various workers have tried different biochemical test from time to time which may help in early diagnosis and confirmation of pulmonary tuberculosis.

Adenosine deaminase (ADA) an enzyme of purine catabolism has been shown to provide promising result. In the present study the diagnostic efficiency of serum ADA activity has been evaluated in pulmonary tuberculosis and non tubercular pulmonary diseases and its relationship with clinical, radiological and laboratory parameters.

Material and Methods:

The study was carried out on 142 patients suspected from various pulmonary disorders between July 2008 and January 2009. Forty five normal healthy individuals (Medical and paramedical staff of Navodaya Medical College Hospital and Research Centre, Raichur, Karnataka) were included as control subjects for ADA estimation. The study groups and number of patients suspected from pulmonary disorders were categorized as shown in Table 1.

Pulmonary TB was defined in two categories

1. Sputum Smear Positive pulmonary TB:

A patient with at least two sputum specimen positive for Acid Fast Bacilli by microscopy.

A patient with at least one sputum specimen positive for Acid Fast Bacilli by microscopy, and radiographic abnormalities consistent with pulmonary TB

A patient with at least one sputum smear positive for Acid Fast Bacilli by microscopy, which is culture positive for *M. tuberculosis*

A decision by physician to treat with a full course of anti TB chemotherapy.

2. Sputum smear negative pulmonary TB:

Two sets (taken at least 2 weeks apart) of at least two sputum specimens negative for Acid Fast Bacilli and radiographic abnormalities consistent with pulmonary TB.

Lack of clinical response despite one week of a broad spectrum antibiotic administration and a decision by physician to treat with a full curative course of anti- TB chemotherapy.

Inclusion and exclusion criteria followed in the present study are as follows:

Inclusion criteria for pulmonary TB

Cases diagnosed as a “new case” of tuberculosis; Possessing at least two positive sputum smear test positive for Acid Fast Bacilli; Radiographic abnormalities consistent with pulmonary tuberculosis, A decision by physician to treat with a full course of anti-TB Chemotherapy, non tubercular pulmonary diseases: a decision by physician.

Exclusion criteria for pulmonary TB

Patients with extra pulmonary TB and/or patients requiring surgical intervention, Chronic pulmonary TB (receiving at least two courses of anti-TB treatment for more than six months), Presence of secondary immunodeficiency states: HIV, organ transplantation, diabetes mellitus, treatment with corticosteroids.

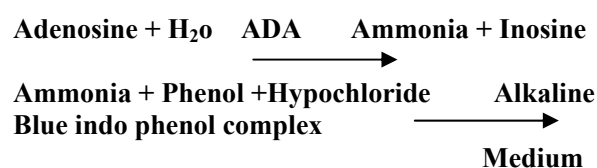
A detailed clinical history was taken and thorough physical examination was carried out in every subject. A set of investigations including three consecutive (spot-early mornings-spot) sputum samples examination, Mantoux test, Posteroanterior chest x-ray and ESR by Westergren method were carried out on study group. ADA estimation was done by sensitive colorimetric method described by Guisti [6]. Serum ADA estimation was done in healthy controls, pulmonary tuberculosis patients and other common respiratory diseases.

Locus of study

The study was conducted in the Dept. of Biochemistry and Microbiology Navodaya Medical College, Raichur and central clinical laboratory Navodaya Medical College Hospital and Research Centre, Raichur, Karnataka.

Principle

Adenosine deaminase hydrolyses adenosine to ammonia & inosine. The ammonia formed further reacts with a phenol as hypochloride in an alkaline medium to form a blue color indophenol complex with sodium nitroprusside acting as a catalyst. Intensity of the blue colored indophenol complex formed is directly proportional to the amount of ADA present in the sample.



This method of serum ADA estimation is based on the principle of Guisti G Galanti methods of enzymatic analyses 1974 [6]. For the determination of Adenosine deaminase activity in serum, plasma, and biological fluids, ADA MTB diagnostic kit from Microexpress a division of Tulip diagnostics (P) Ltd has been used.

Result

Control Group

Forty five normal individuals aged between 18 years - 60 years [30 Males (66.6%) and 15 females (33.3%)] were included as controls. In this group the serum ADA active-

Table 1: Study groups of pulmonary tuberculosis and non tubercular pulmonary disease patients

S.No	Study Group	No. of Patients
1	Group-I Pulmonary tuberculosis	101
	A-Sputum positive patients (AFB +ve)	64
	i) Moderate	39
	ii) Advanced.	25
	B- Sputum negative patients (AFB -ve)	37
	i) Initial	11
	ii) Moderate	11
	iii) Advanced.	15
	2	Group-II Supparative lung diseases
A- lung abscess		3
B- Bronchiectasis.		5
3	Group-III Malignancy of lung.	2
4	Group-IV Empyema	4
5	Group-V Tubercular pleural effusion	27
Total		142
Healthy Controls		45

ity level ranged from 14.48 to 16.02 U/L with a mean value of 15.50 ± 0.52 U/L as mentioned in Table 2.

Out of 142 patients suspected from various pulmonary disorders, 101 (71.1%) were found to be positive for pulmonary tuberculosis. Among 101 pulmonary tuberculosis cases 64 (63.3%) were sputum AFB positive and 37 (36.6%) were Sputum AFB negative. The mean serum ADA activity in the study group was 36.35 ± 12.15 U/L as compared to 15.50 ± 0.52 U/L in control group.

None of the patients of non tubercular pulmonary diseases showed serum ADA level above 33 U/L, though their values were significantly high compared to control group. Hence serum ADA of 33 U/L is taken as cut off value. The mean serum ADA activity level in pulmonary tuberculosis (sputum AFB positive as well as in AFB negative cases) was 40.48 ± 8.02 U/L.

While the same in non tubercular respiratory diseases, taken together was 28.4 ± 4.2 U/L. [Lung abscess 26.6 ± 2.4 U/L, Bronchiectasis 27.65 ± 2.35 U/L malignancy

lung 25.22 ± 2.22 and empyemas 28.55 ± 4.05 U/L] as depicted in Table 3. The difference in ADA level between patients of Pulmonary tuberculosis and Non tubercular pulmonary disease was significant ($p < 0.001$).

In the present study, specificity and sensitivity of serum ADA level in the diagnosis of pulmonary TB was found to be 95.35% and 98.06% respectively.

No significant difference was observed in mean serum ADA activity in different sex and age groups among these patients. However the mean serum ADA activity level in sputum AFB positive as well as in AFB negative cases was 40.48 ± 8.02 and 41.6 ± 6.4 U/L respectively.

Four groups of patients namely A (0-20 yrs) B (21-40 yrs) C (41-60 yrs) and D (≥ 61 yrs) was formed, as shown in Table 4. The diagnosis of pulmonary tuberculosis was based on ESR, Sputum AFB, Serum ADA, Mantoux test and chest x-ray. Serum ADA level was positive in 89 (88.1%) cases followed by 76 (75.3%) in Mantoux test, 73 (72.2%) in chest x-ray, 62 (61.3%) in sputum AFB and 63 (62.3%) in ESR.

Table 2 Serum Adenosine Deaminase level in control groups,patients with pulmonary tuberculosis and non tubercular pulmonary diseases

Group Range	Study Population	No. of Patients	Mean	Serum ADA levels U/L	P-Value
I	Healthy Controls	38	(15.50± 0.52)	(14.98 - 16.02)	P <0.001
II	Pulmonary Tuberculosis (Sputum P.S -VE Culture +ve)	36	(40.48± 8.02)	(32.46 - 48.50)	
III	Non Tubercular	37	(28.40± 4.20)	(24.20 - 32.60)	

Table 3: Serum Adenosine Deaminase levels in various study groups & Health controls

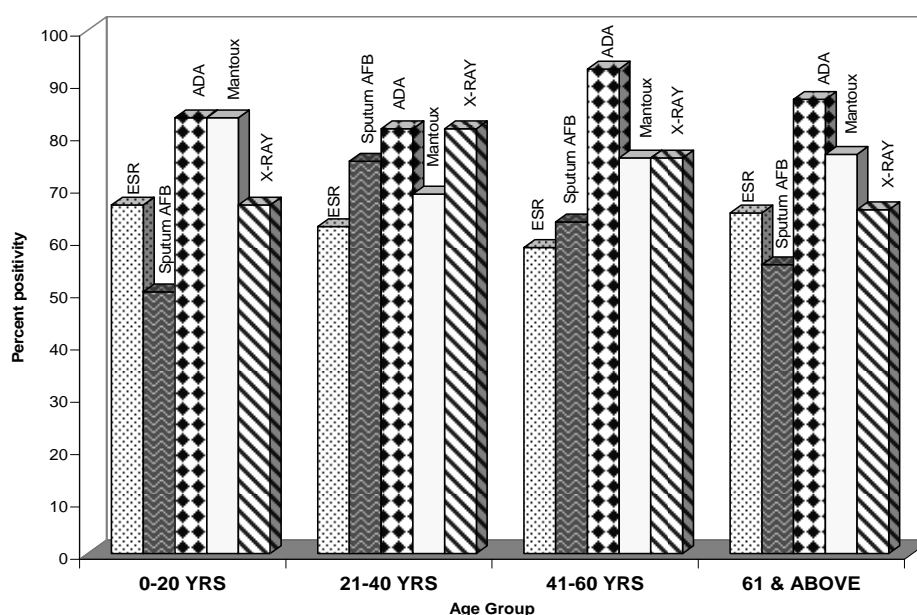
S. No	Study Group	Serum ADA Activity	
		Range U/L	Mean U/L
1	I - Pulmonary Tuberculosis		
	A-Sputum positive patients (AFB +ve)		
	i) Moderate	35.20 - 47.50	41.35
	ii) Advanced	38.50 - 48.00	43.25
	B-Sputum negative patients (AFB -ve)		
	i) Initial	36.60 - 48.50	42.55
	ii) Moderate	32.46 - 43.22	37.84
	iii) Advanced	34.80 - 42.50	38.85
2	II - Supparative Lung diseases		
	A-Lung Abscess	24.20 - 29.00	26.6
	B-Bronchiectasis	25.30 - 30.00	27.65
3	III - Malignancy lung	23.00 - 27-44	25.22
4	IV- Emphyema	24.50 - 32.60	28.55
5	V- TB Pleural effussion	38.50 - 44.20	41.35
6	Healthy Controls	13.40 - 21.40	17.4

Table 4: Showing different parameters with positive findings in the diagnosis of Pulmonary Tuberculosis

AGE GROUP	ESR	AFB	ADA	MT	X-RAY CHEST
A [0-20 Yrs]					
n=6	04 (H)	3	5 (H)	5	4
B [21-40 Yrs]					
n=16	10 (H)	12	13 (H)	11	13
C [41-60 Yrs]					
n=41	24 (H)	26	38 (H)	31	31
D [\geq 61Yrs]					
n=38	25 (H)	21	33 (H)	29	25
TOTAL	63	62	89	76	73

	(62.3%)	(61.3%)	(88.1%)	(75.2%)	(72.2%)
MT=	Mantoux test				
MT +VE =	More than 10 MM Erythema and induration				
ESR =	Erythrocyte sedimentation rate by Westergren Method				
Serum ADA =	Serum Adenosine Deaminase (Cut off value >33 U/L)				
Sputum AFB=	Sputum acid fast bacilli by Z-N staining technique				
(H)=	High (values above normal range)				
n=	number of subjects				

Graph showing percent positivity of different parameters in the diagnosis of Pulmonary Tuberculosis



Discussion

Various workers have found different values for normal human serum ADA level at 37⁰ C. Low serum ADA levels have been reported by Schwartz et.al [7] (12.49 ± 2.50 U/L) and krawczynski et.al[8] (13.14 ± 4.28U/L).Higher serum ADA level was reported by Giusti[6] (17.05 ±3.75 U/L) and Jhamaria JP et.al [9] (19.09 ± 2.99 U/L).However serum ADA level in control group reported by Guisti et.al [10] (15.8 ± 3.7 U/L) was almost at par with our study.

In our study, serum ADA levels were found to be significantly high in patients with pulmonary tuberculosis and patients with non tubercular pulmonary diseases compared to control group. Agarwal MK et.al [11] and Jhamaria JP et.al [9] also found increased serum ADA level in patients with pulmonary tuberculosis and patients with non tubercular pulmonary diseases. However the rise

was much higher in patients of pulmonary tuberculosis. The significant finding in our study was, clinically suspected and sputum AFB negative cases have also shown high serum ADA level at par with sputum AFB positive cases. Alatas.F et.al[12] in their studies determined the role of serum ADA activity in the diagnosis and follow up of pulmonary tuberculosis and monitoring the efficiency of therapy. A Significant difference was observed in ADA activity before and after treatment , also from old TB patients and healthy control subjects. This shows that serum ADA activity is increased in pulmonary tuberculosis patients which is a help full parameter for monitoring therapy.

Paliwal.R et.al [13] evaluated the efficiency and usefulness of serum ADA activity for diagnosis of pulmonary tuberculosis and other non tuberculosi respiratory condition, serum ADA levels were determined in 10 healthy, 90 patients with 65 pulmonary tuberculosis,

15 patients with suppurative lung disease and 10 with lung carcinoma. The sensitivity and specificity of serum ADA, as a diagnostic test for pulmonary tuberculosis were found to be 100 % and 88.6 % respectively, which can be considered as an important investigation to arrive at a diagnosis.

In our study, taking 33 U/L as cut off point, the specificity and sensitivity of serum ADA level was 95.35% and 98.06% respectively. However Jhamaria P et al[9] using 33U/L as cut off limit had found 100% specificity and 98% sensitivity of serum ADA level for diagnosis of pulmonary tuberculosis. Agarwal MK et al[11] reported high specificity and sensitivity of serum ADA in the diagnosis of smear negative, culture positive pulmonary tuberculosis taking 33 U/L as cut off point.

In the present study, a high occurrence of pulmonary tuberculosis was found in the age group 41-60 years with Serum ADA positivity of 92.6% (38/41) cases. Percent positivity of serum ADA was more, compared to other diagnostic tests like Mantoux test, sputum AFB, chest x-ray and ESR. However elevated levels of ADA have been reported in effusions due to peritoneal, meningeal, pleural, pericardial and in several diseases like Typhoid fever, infectious mononucleosis [14], brucellosis and bronchogenic carcinoma [15] involving stimulation of cell mediated immunity. According to Giblett et.al [16], a fully functioning cell mediated immune response is dependent on normal lymphocyte metabolism which is, in part, regulated by the purine salvage enzyme, Adenosine deaminase. Therefore increased serum ADA activity is also found in other diseases involving stimulation of cell mediated immunity.

As determination of ADA is not costly or time consuming, should be done routinely, particularly if the diagnosis of tuberculosis is in doubt, sputum AFB negative pulmonary tuberculosis cases and also to differentiate pulmonary tuberculosis from non tubercular pulmonary diseases.

References

1. World Health Organization. The World Health Report: making a difference. 1999: 110.
2. Revised National Tuberculosis Control Programme: Key factor and Concepts. New Delhi: Central TB Division, Directorate General of Health Services, Ministry of Health and Family Welfare; 1999.
3. Lalit Kant. Improving detection of infectious cases. Indian J Tuberculosis. 2001; 48: 115-6.
4. Ananthnarayan R, Jayaram Panikar CK. Text book of Microbiology. 7th edition. Universities press; 2005:351-364.
5. Stead WW, Lofgren JP, Warren E, Thomas C. Tuberculosis as an endemic and nosocomial infection among the elderly in nursing homes. N. Engl. J Med. 1985 ; 312 : 1483-1487.
6. Giusti G. Adenosine deaminase. In Bergmeyer H.V.Ed. Methods of enzymatic analysis. Vol-2, New York Academic press, 1974:1092.
7. Schwartz and Bodansky . Proc .Soc Exptt.Biol.Med :1959, 101,560.
8. Krawczynski J, Raczynska J, Zonas S, Wencel J and Ilowiecka, K. Clin. Chem, ACTA. 1965; 11: 227.
9. Jhamaria JP, Jena, RK, Lutada, SK, Mathur, D.K Parihar H.L and sharma, SK. Serum ADA in differential diagnosis of Pulmonary Tuberculosis and common non Tuberculosis respiratory diseases. Indian J.Tuberculosis. 1988; 35: 25-29.
10. Giusti,G & Gakis.B. Enzyme. 1971; 12: 417.
11. Agarwal MK, Jitendra Nath, Mukerji PK, V.M.LSrivastava. A study of serum Adenosine Deaminase activity in Sputum Negtive patients of Pulmonary Tuberculosis. Ind J.Tub. 1991; 38: 139-141.
12. Alatas F, Uslu S, Moral H, Alatas O, Metints M.Erginel S Uegun I. Serum adenosine deamise activity in Pulmonary tuberculosis. Tuberk Toraks 2003; 51(3) ;277-81.
13. Paliwal R, Shah K.V. Serum adenosine deaminase estimation in patients with Pulmonary Tuberculosis and other non tuberculosis respiratory conditions. Indian J of Tuberculosis. 1998; 45(3): 174.
14. Piras MA. Immunological Studies in Mediterranean Spotted fever. Lancet. 1982; 1 (8283): 1249.
15. Latnansky and Seflich. Klin Wschr. 1958; 36: 826.
16. Giblett E R , Anderson I E, Cohen F. Pollara B and Meucuissen J.ADA deficiency in two patients with severely impaired cellular immunity. Lancet ;1972, 22; 1067.

Correspondence

K. Srinivasa Rao
 Dept.of Biochemistry
 Navodaya Medical college
 Mantralayam Raod
 Raichur-584103.
 India
 e-mail: srini.nmc@gmail.com

