The effects of different sorghum varieties on the microbial fermentation dynamics of Huangjiu (Chinese-rice-wine).

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Abstract

Huangjiu (Chinese-rice-wine) is a fermented alcoholic beverage traditionally brewed with cereal grains (mainly wheat and rice) and wheat Qu (starter cultures), thus making it undesirable to consumers with gluten dietary restrictions. With the increased interest towards developing alternative food products for niche markets (gluten intolerant consumers), sorghum represents an ideal candidate due to the lack of gluten and important source of dietary antioxidants. Therefore, the objective of this research was to study the effect on fermentation dynamics when a gluten-free commercially available starter was used to produce Huangjiu from sorghum. Three different sorghum whole grain varieties (white, waxy, sumac) were used as substrates. Microbial and physiochemical analysis were conducted in triplicates. Taxonomy annotation and Principal Coordinates Analysis (PCoA) were used to describe the microbial profiles of the sorghum Hangjiu. By the end of the study, the three sorghum Hangjiu reached the pH range between 3.7 to 4.1, Aerobic bacteria, yeasts and molds increased progressively during the first part of the fermentation and followed a steady decline by day-10. Saccharomyces was the dominant genera detected in the starter culture and in the final fermented product. The highest fermentation yield was observed in sorghum (Table 1). This study demonstrated the possibility of using alternative ingredients in Huangjiu production by utilizing a different substrate (e.g., sorghum) for consumer's niche market.

Keywords: Sorghum-Based food, Huangjiu, Fermentation dynamics.

Introduction

Globalization has caused the uptake of Asian culture in the west for the past decades with Asian food culture entering the popular lifestyle. Recent reports showed Asian dining as the largest segment in single location full-service restaurants in America, with \$45.6 billion revenue in 2021 [1]. At the same time, also the interest in eastern-styled fermented beverages has risen and in particular for beverages derived from rice, such as Huangjiu, a fermented alcoholic beverage traditionally brewed with cereal grains (mainly wheat and rice) and wheat Qu (starter cultures) [2]. Huangjiu, or Chinese rice wine, has a long and celebrated history in the culinary culture, since it is one of the most ancient alcoholic drinks, alongside wine, and beer. Huangjiu is known for its unique and full-bodied flavor profile with relatively low alcohol content. This non-distilled alcoholic drink contains plenty of variations native to different regions [3-5].

Huangjiu production mainly consists of three steps: selection and pre-treatment of raw materials (rice, corn, wheat, or millet), fermentation, and post-process treatment [6]. The selected grains are soaked in water for 2-3 days before steaming. The starter, composed of dry rice flour, wheat and *Rhizopus* spp. is added to the steamed, cooled grains to jump start the saccharification, breaking down the grains into sugars, peptides, and amino acids that are converted during fermentation [7]. After a primary fermentation at 30°c for 5-7 days, the mash enters a secondary fermentation for up to 2 months at 15°c before being filter-pressed, clarified and pasteurized for safe consumption [8].

Since Huangjiu production traditionally involved wheat, this fermented beverage is unsuitable for people with wheat allergy, gluten sensitivity, or celiac disease. A recent survey showed that celiac disease affects about 1% of people in most populations, with an increasing trend of diagnosis rates [9]. The market demand for gluten-free products in foodservice and retail is projected to reach \$11.4 billion by 2026 in the United States of America (USA) [10]. Altering the ingredients in Huangjiu production by utilizing a different substrate (e.g., sorghum) might offer a suitable alternative for this niche market.

Sorghum is one of the top five cereal crops in the world, and the USA is the world's leading producer, with about 11.4 million metric tons of sorghum produced in 2020 [11]. This grain is primarily used for animal feed production, ethanol

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and biofuel, but it is attracting attention in the food industry because of its nutritional value and versatility in applications [12]. Compared to other cereals, sorghum has high antioxidant activities and lower digestion rate [13]. These properties offer food scientists and product development professionals' creative ways for gluten-free market applications. Nevertheless, not much is known about the microbial dynamics when alternative substrates, e.g., sorghum produced in USA, is used for Huangjiu production. Therefore, the objective of this research was to study the effects on microbial population when three different types of sorghum (namely white, waxy and sumac) grown in the USA were used for Huangjiu fermentation. The types of sorghum in this study were selected based on antioxidants levels, fiber contents and applications versatility for the food industry.

Materials & Methods

Whole sorghum grains and starter culture

Grain sorghum can take many shapes and sizes (tight-headed, round panicle to an open, droopy panicle, short or tall) and it comes in red, orange, bronze, tan, white, and black varieties. Three commercially available sorghum varieties (NU life market, LLC., Scott City, KS, USA) were used in this study: white whole sorghum grain (NLSC-WSWG-0010, lot# 210722-NLM10), a sustainable ancient grain, non-tannin that is grown using fewer natural resources, typically made into flour for food production; waxy burgundy whole sorghum grain (NLSC-WBBWG-0014L, Lot# 210811-NLM-20), a thinner bran layer, with lower gelatinization temperature and amylopectin starch; and sumac whole sorghum grain (NLSC-SUSWG-0018, Lot# 210811-NLM-SB), with a high-tannin, high-polyphenolics contents and excellent anti-inflammatory properties [14,15]. For analysis purposes sorghum flour was prepared on a cyclone mill with a 0.5mm and/or 0.25mm screen based on analytical request and the starter culture "Angel Yeast" (Angel Yeast co., Hubei, PRC) was purchased from an online supplier in bulk of 500g.

Huangjiu preparation

The preparation of Huangjiu was developed in our laboratory based on literature review [14,16-20]. A schematic representation of the preparation, inoculation and analysis followed in this research for the preparation of Huangjiu is shown in Figure 1 below. Briefly, white, waxy, and sumac raw whole sorghum grains were rinsed three times, cooked with a 1:2 weight ratio of dry grain to water using a pressure cooker (Instant Pot Duo, Instant Brands™ Inc., Ontario, CA) for 45 minutes (115 - 118°C and 10.2 - 11.6psi). The cooked sorghum (substrate) was covered with aluminum foil and cooled until it reached room temperature ($20 \pm 2^{\circ}$ C). The commercial starter was then added (0.8% w/w) and mixed thoroughly. The inoculated mixture was divided into portions of 100g inside sterile beakers, packed tightly, covered with aluminum foil, and incubated at 30°C for 10 days. This procedure was repeated for each sorghum type. At the end of the fermentation process two products were obtained: a liquid fraction (Huangjiu) and a solid fraction (spent grain).

Physical and Chemical Analysis

At the beginning and the end of the fermentation period (0 and 10 days), the pH of the liquid fractions obtained from the different fermented grains were recorded using a calibrated pH meter (Traceable® Digital pH/ORP pH meter, VWR, Texas, USA). Yield of the liquid fraction was calculated as a percentage of decanted liquid into a 50 ml graduated cylinder and the volume was reported. Additionally, the color of the liquids was observed using a visual scale color coded by 3 food scientists who reached consensus on grading the colors from lightest to darkest [21].



Figure 1. Schematic representation of the preparation, inoculation and analysis followed in this research for the preparation of Huangjiu from sorghum grains.

Microbiological analysis

The pattern of microbial succession throughout the fermentation process was studied. At appropriate sampling points (0, 1, 3, 5, 7, and 10 day), 25g of grain/liquid mixture was collected and homogenized in 1:10 volume of 0.1% Buffered Peptone Water (Becton, Dickinson and Company, NJ, USA). The samples were then serially diluted and enumerated to log10 CFU/g for yeasts and molds and total aerobic bacteria by spread plating onto Rose Bengal Agar (Becton, Dickinson and Company, NJ, USA), and incubated at 25°C for 48 hours, and Aerobic Count (AC) 3M Petrifilms (3M, MN, USA) and incubated at 37°C for 24 hours, respectively.

DNA extraction, shotgun sequencing and bioinformatics analysis

Total genomic DNA was extracted at day 0 from the three sorghum grains before starter addition (negative controls), and from the grain/liquid mixture at day 1 and 10. The starter alone was used as positive control. DNA was obtained using the PowerSoil Pro Kit (QIAGEN, Hilden, Germany) according to the manufacturer's instructions. DNA was quantified using Qubit 4 (Q32857, Invitrogen, MA, USA). DNA extracts were stored at -20°C until further use. DNA libraries were prepared using the Nextera DNA Flex Library Preparation Kit (Illumina, USA). The prepared library of each sample was quantified, and quality controls were performed. Libraries were pooled at equimolar amounts, and sequencing was performed with 2×150 bp chemistry on a NovaSeq 6000 platform (Illumina).

A workflow for DNA library prep and metagenomics analysis performed on the samples collected in this study is given in Figure 2. Briefly, original sequence reads were pre-processed by trimming low quality bases (Q-value \leq 38), reads containing over 10bp N nucleotides, or reads containing over 15bp of overlapping adapter. Bowtie2 was used to minimize host DNA. The metagenomic reads were compared to the database of taxonomically informative gene families (microNR database) for homolog annotation. Taxonomic ranks were based on gene abundance and analyzed using Krona analysis (https://github.com/marbl/Krona/wiki). Gene prediction was carried out by

MetaGeneMark based on the Scaftigs assembling. Predicted genes were pooled together for dereplication to construct gene catalogue. The abundance information of the gene catalogue for each sample was obtained by comparing clean data of each sample from the gene catalogue. The function of the coding sequences was inferred by comparing their similarity to sequences in the databases (KEGG, eggNOG, CAZy, etc), a principal coordinate analysis (PCoA) and heat maps were used for describing the relative abundance in the metagenome.

Data Analysis

Throughout the fermentation process, assays (Microbial count, pH, color) were run in triplicates for each grain variety and sampling time. Sample unit was defined as 25g of each grain variations at the sampling interval (Total N= 21). Collected data were analyzed and plotted in Microsoft Excel and means comparison of microbial population and physiochemical traits during fermentation were determined by T-Test with a significance level of $\alpha = 0.05$.

Results and Discussion

Changes in physical characteristics during fermentation

High-pressure cooking was used to ensure the microbial safety and quality of the cooked whole sorghum grain kernels. The whole sorghum grain softens while retaining their color during cooking. White and sumac cooked grain had a fluffy appearance, whereas cooked whole waxy grain appeared to be sticky, possibly due to starch activation during stirring after the grains were cooked

After cooking, some visual physical differences based on the sorghum variety used as fermentation substrates were observed featuring different characteristics in the final product as well. Water entered sorghum kernels during high-pressure cooking and resulted in physical changes in the kernels, hydrated the chemical components, and almost doubled the volume of cooked sorghum kernels. Starch granules were hydrated, swelled, and gelatinized. During fermentation, the liquid fraction yield increased with time. White sorghum grain had the highest liquid yield after 10 days of fermentation (Table 1). Waxy sorghum grain had the earliest sign of free water release



Figure 2. DNA library preparation and metagenomic workflow analysis.

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Table 1. Visual color, pH and yield of the liquid fractions obtained from the different grain varieties after 10 days of fermentation.

Sorghum sample	Visual color	pH		
		Day 0	Day 10	rieid (%)
White whole sorghum	Light yellow	6.61 ± 0.0 ^a	4.1 ± 0.2^{a}	55 ± 1.33ª
Waxy whole sorghum	Yellow	6.65 ± 0.0ª	3.9 ± 0.1ª	51 ± 3.34 ^b
Sumac whole sorghum	Dark red	6.36 ± 0.0^{a}	3.7 ± 0.2^{a}	50 ± 0.26 ^b

Values for pH and yield are averages of triplicate readings (mean \pm standard deviation). Values followed by a different superscripts letter within the same column indicate that there is a significant difference between the sorghum types (p < 0.05).

out of the three, as early as day 1, sumac sorghum showed the lowest volume of free water release throughout the duration of the study, likely due to the high polyphenolic contents as compared to the waxy variety. The liquid fraction yield varied based on the grain variety used. Sorghum Huangjiu production from sumac had the lowest yield amongst the three varieties. The color of the three sorghum Huangjiu did not differ from their whole grain color at the end of 10-day fermentation, white, waxy, and sumac sorghum Huangjius appeared white/ yellowish, red, and dark brownish red, respectively. All three sorghum grains started with pH values above 6.30 (Table 1), as the organic acids in the fermented substrates accumulated during fermentation, the pH decreased over time.

Changes in microbial populations during fermentation

With the addition of the starter, the level of aerobic bacteria, yeasts and molds were significantly increased up to $\sim 3 \log$ CFU/g and ~ 7 log, respectively on day 0 (p<0.05). The changes of microbial population during fermentation are shown in Figure 3 below. After the boost from the inoculation starter, aerobic bacteria counts were between 1.28 to 2.11 log CFU/g until the endpoint for the three sorghum grains. Aerobic bacteria population remained steady overtime, no noticeable changes were observed when different sorghum varieties were used as substrates. This could be due to the stable environment provided by cooked sorghum grains, and the concise inoculation of the fermentation starter, providing dominance of certain microbes. Different observations were made for the yeasts and molds population. After the significant uptrend in population (p<0.05) between inoculation day and day 1 for all three grains, from the fermentation starter. No significant changes (p>0.05) in yeasts and molds population for white (Figure 3A) and waxy (Figure 3B) over time, while a significant decline (p<0.05) was reported when sumac sorghum was used as fermentation substrate (Figure 3C). The yeast and molds population reacting to different fermentation by-products differently based on the difference in chemical compositions in the sorghum varieties, the higher tannins level of this variety as compared to the other sorghum types used and the antifungal effect of these compounds might be observed here [22].

Metagenomic sequencing and composition profiling of microbial communities

Processed samples had more than 99.9% reads passed quality control and host filtering. All samples had an average read length between 772 to 1426 bp.

While the labeling of the commercial starter culture purchased in this study listed rice flour and *Rhyzopus* spp. as the main ingredients, Krona analysis identified 70% of the starter culture as Saccharomyces spp. (Figure 4). The starter culture also contained also bacteria (~15%), most belonging of belonged to the genus Bacillus. It also contained 1% Aspergillus mold. Findings from this study were slightly different from those reported by [23,24] where Rhizopus, Aspergillus, Saccharomyces and bacteria belonging to genus Weissella, Pediococcus, Pantoea, and Lactobacillus were isolated from Huangjiu starter cultures. Rhizopus spp. was not found in the starter samples and not have much variety in the bacteria populations was observed. Possible explanations include the use of different commercially available starter culture, as compared to the one used in this study, or the use of starters not in the market or from different regions of the world. Figure 5 described the taxonomy annotation of the liquid fractions when white (A-B), waxy (C-D) and sumac (E-F) where used as substrate for fermentation. Saccharomyces spp. Continue to be taxonomically relevant with concentrations varying from 85 to 87%. No significant difference (p>0.05) among sorghum varieties or sampling days (day 1 or day 10) was observed. This may be due to the enduring property of *Saccharomyces* spp. allowing them to thrive in osmotic, temperature and ethanol stresses found in the fermentation environment [25].

Heatmaps are a useful method to explore multivariate data sets, where the response variables (in this study relative abundances) are visualized using color gradients scheme. Figure 6 represents the distribution of the dominant 35 genera of all samples displayed in the relative abundance heat map. Different colors represent different relative abundance of the genus in all seven samples tested, with red indicating the greatest abundance. The genus Aspergillus spp. was the most abundant in the starter culture (red color in Figure 6), but its presence was inconsistent during the fermentation. The blue colors indicate low relative abundance at day 1 with a slight increased at day 10. This pattern was common to all the samples tested, independently of the variety of sorghum used as substrate. Conversely the relative abundance of Saccharomyces was high and constant (purple color) over time in all samples, no differences among sampling days or grains were observed. Similar trends were observed also for Bacillus spp. This genus was persistent across all samples and varieties, but with relatively low abundance (light blue). Similar findings were reported by Liang et al. [26] that observed Aspergillus and Rhizopus dominant at the beginning of the fermentation but rapidly outcompeted by Saccharomyces and other bacteria during fermentation, similarly to our observations, likely due to ethanol metabolized by Saccharomyces increased toxicity towards populations that does not share the same metabolic pathway, suppressing their growth [26].



Figure 3. Microbial population changes in Aerobic Bacteria (APC) and Yeasts and Molds (Y&M) in white (A), waxy (B) and sumac (C) sorghum samples during fermentation.



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Figure 5. Taxonomy annotation visualization of the liquid fractions using the three different sorghum variates as substrates: white sorghum at day 1 (A) and day 10 (B); waxy sorghum at day 1 (C) and day 10 (D), sumac sorghum at day 1 (E) and day 10 (F).



Figure 6. Distribution of the dominant 35 genera of all samples displayed in the species relative abundance heat map. Different colors represent different relative abundance with red indicating the greatest abundance.



Figure 7. Principal Coordinates Analysis (PCoA) was used to visually differentiate the closeness of diversity amongst the samples. The first and second Principal Components (PCs) accounted for 99.8%, and 0.19% of the variance, respectively.

The principal coordinate analysis (PCoA) was conducted to visualize the closeness of diversity among samples (Figure 7) and to demonstrate data similarities or differences in the fungal and bacterial community. The PCoA score plot shows that all the liquid samples varied significantly in fungal and bacterial community as compared to the starter sample. The close positions among waxy samples and white samples highlight their similar principal genera composition, while sumac samples were separated in the PCoA plot, indicating a significant difference genera composition between day 1 and day 10.

Conclusion

This study explored changes in microbial population when different varieties of sorghum were used to produce a glutenfree sorghum Huangjiu, showcasing the potential of leveraging fermentation to improve the utilization of sorghum grain for human consumption. Additional research is needed, which was beyond the scope of this study, to measure the alcohol content, nutritional value, and conduct sensory evaluation of the different treatments and compare to traditional Huangjiu.

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Conflicts of Interest

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