

Review on antimicrobial mechanisms of spices-essential oil constituents.

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Abstract

Spices have been employed as a flavoring and coloring agents. Essential oils of spices have wide antimicrobial functions. Specific constituents of the essential oils inhibit the synthesis of bacterial cell division proteins, cell membrane depolarization, decrease adenosine triphosphate and disturbing the ionic equilibrium. In the fungal species, the constituents of essential oils decrease the ergosterol synthesis interact with delta-14-sterol reductase, affecting mitochondrial enzymes and accelerating the telomere shortening. Present review discussed the antibacterial and antifungal mechanisms of spices 'essential oil constituents'.

Keywords: Spices, *Cinnamomum cassia*, *Ocimum basilicum*, Antimicrobial, Essential oil.

Introduction

Plants contain numerous chemical components with different biological functions. Aromatically scented herbal products have been used since ancient times to flavor foods. The species of *Cinnamomum cassia* or Chinese cinnamon, *C. verum* or true cinnamon contain the spice ingredients in their inner bark. Cinnamon barks essential oils containing cinnamaldehyde as an antibiotic ingredient. The seeds of *Thymus vulgaris* or German thyme and *Trachyspermum ammi* or bishop's weed contain essential oils that possess thymol as an antibiotic molecule. Bishop's weeds are widely used in food products due to its pleasant flavor and preservative properties. Cumin or *Cuminum cyminum* contains β -pinene as an active antimicrobial monoterpene. Linalool is an acyclic monoterpene alcohol and antibiotic found in *Ocimum basilicum* plants. The essential oils of *Anethum graveolens* seeds containing carvone and limonene as antibiotic molecules. *Trans*-anethole is an antibiotic molecule found in the fruit essential oils of *Pimpinella anisum* L species. These different plant origin antibiotic molecules have potent inhibitory functions of bacterial species and fungal variants. The present review discusses the antimicrobial mechanisms alone from the selected research articles as part of our academic project, semester-V and subject code: BFP508S in B.Voc. Food processing programme.

Cinnamaldehyde

All bacterial species have a conserved cell division marker termed Filamentous temperature -sensitive protein Z (FtsZ) in the cytoplasm. Bacterial cytokinesis is mediated through the formation of a Z ring that contains the FtsZ of the cell wall component. The formed Z ring associates with other

cell division proteins and contracts gradually for the closure of septum and formation of two daughter bacteria cells [1-3]. Cinnamaldehyde inhibits the bundling of FtsZ protofilaments in the Z ring and exerts antimicrobial response in the bacterial species [4,5]. Cinnamaldehyde interfere with the mechanisms of expression of cell wall construction genes which further lead to the damage of cell wall permeability and integrity *Geotrichum citri-aurantii* [6]. Cinnamaldehyde as a component is damaging the cell wall integrity by inhibiting the ergosterol content [7], membrane invaginations, elevating the levels of reactive oxygen species, destruction of organelles and disorganization and leakage of cytoplasm in the fungal spores of *P. expansum* or blue mold decay fungi [8]. Cinnamaldehyde inhibits the spore germination, mycelial growth and fungal biomass production in *A. flavus* by reducing the ergosterol synthesis, mitochondrial membrane potential, elevating Ca^{2+} levels, reactive oxygen species, release of cytochrome c, activation of metacaspase, phosphatidylserine externalization and DNA damage [9]. *Trans* form of cinnamaldehyde changes the cell membrane permeability in *P. italicum* causing the leakage of cell material [10]. Cinnamaldehyde influences the function of fungal mitochondria by inhibiting the activity of ATPase and SDHase [11].

Thymol

Thymol 2-isopropyl-5-methylphenol is a monoterpene plant compound. Different candida species are susceptible with mean diameter of inhibition zone between 15-21 mm [12]. Thymol and carvone compounds are toxic for the growth of 12 different fungal strains at its lowest concentrations [13]. Phenols located in the ring structure are the vital cause for antifungal activity [14]. Thymol accelerates telomere shortening in the

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yeast and increases the rate of cell senescence and apoptosis [15]. Intracellular Ca²⁺ ions homeostasis, glycan-mediated post-translational modifications and the levels of endogenous ergosterol content was significantly decreased in the thymol treated *Cryptococcus neoformans* [16]. Furthermore, thymol treatment reduced endogenous ergosterol content by decreasing the expression of ergosterol biosynthesis genes in a high-osmolarity glycerol (HOG) - mitogen-Activated Protein Kinase (MAPK) pathway-dependent manner. Thymol has the ability to integrate into the Lipid layer of the cell membrane, increasing the surface curvature and also affecting the activity of internal membrane enzymes and receptors [17].

Thymol is destroyed the integrity of cell membrane resulting in leakage of intracellular material and eventual death of zoonotic bacteria *Aeromonas hydrophila* [18]. It decreases the intracellular ATP pools of *E. coli* by disrupting the cytoplasmic membrane [19]. Thymol affects the cell membrane depolarization, decrease intracellular ATP concentrations and lowers pH_i in the *Enterobacter sakazakii* [20]. Thymol promotes the leakage of proteins and nucleic acids in the vegetative and spore forms of *Alicyclobacillus acidoterrestris* [21].

Beta pinene

Beta-pinene (β -pinene) is a monoterpene, essential oil found in Cumin or *Cuminum cyminum*. Derivatives of β -pinene are antimicrobial to the bacterial strains of *Klebsiella pneumoniae*, *Enterobacter aerogenes*, *Staphylococcus aureus*, *Staphylococcus epidermidis* and fungus of *Candida albicans*. Change in substituent on the pyridine ring and benzene ring of 3-cyanopyridine derivatives is an important factor for inducing antimicrobial activity [22]. The bacterial strains of staphylococcus lost viability significantly 2-8 hours of β -pinene exposure [23]. (+)- β -pinene is an effective molecule in reducing the candida biofilm formation. The crucial antifungal activity is mediated through interference with the cell wall; through molecular interaction with Delta-14-sterol reductase and, to a lesser extent, with the 1,3- β -glucan synthase [24].

Linalool

Linalool also known as 3,7-dimethyl-1,6-octadien-3-ol is an acyclic monoterpene alcohol found in *Ocimum basilicum* with reported antimicrobial and antifungal properties [25]. Linalool affects the bacterial cells by losing the cell membrane polarization, wrinkling, glue together and broken conditions. Additionally, linalool is causing the intracellular nucleic acid leakage and decreasing the activity of bacterial respiratory dehydrogenases, ATPase's, pyruvate kinase activities and disturbing the central carbon chain metabolism [26,27]. Linalool is affecting the biofilm formation in the fungal strain of *C. albicans* ATCC 14053. Linalool is affecting the expression of adhesin genes and the genes responsible for germ tube formation which potentially inhibits the growth of *C. albicans* fungal strain [28].

Carvone and Limonene

Anethum graveolens (Dill) seeds containing essential oils of carvone and limonene possessing antimicrobial activity.

Essential oils increase relative electric conductivity, extracellular ATP concentration and cell constituent of *Campylobacter* species [29]. Dill compounds inhibiting the bacterial strains of *Staphylococcus aureus* ATCC25923, clinical *Vibrio cholerae*, *E. coli* ATCC 25922 and *Pseudomonas aeruginosa* 8821M [30]. *E. coli* and *S. aureus* strains facing alterations in hydrophobicity, surface charge and membrane integrity that subsequently promote the K⁺ ions leakage [31]. Dill seed oils also cause morphological changes in the cells of *Aspergillus flavus* and a reduction in the ergosterol quantity. Fungal cells losing the mitochondrial membrane potential, decreasing ATPase and dehydrogenase activity which indicates dill seed essential oils inhibiting fungi through induction of reactive oxygen species [32]. Derivatives of carvone effective against *E. coli* and weak antifungal over *C. tropicalis* and *C. parapsilosis* [33]. The essential oils containing 78.76% of carvone and 11.50% of limonene are effectively inhibiting the growth of gram positive bacteria [34]. Carvone of *Anethum sowa* is inhibiting the growth of human pathogenic bacteria and fungi [35]. The components of carvone and limonene significantly inhibit the growth of the *Colletotrichum gloeosporioides*, *Lasiodiplodia theobromae* and *Alternaria fungal* isolates [36]. Carvone is a versatile antifungal compound promoting the cell surface roughness, formation of pores and leakage of cellular content in the candida species [37]. Carvone is an effective inhibitor for the formation of germ tubes in the *Candida albicans* [38]. Thymol and carvone compounds are toxic for the growth of 12 different fungal strains at the lowest concentrations of 0.017% to 0.051% [13].

Trans-anethole

Pimpinella anisum L fruits containing *trans*-anethole as the major essential oil component [39,40]. Aqueous decoction of aniseed is 18.1% effective against 176 oral bacteria isolated from 200 individuals [41]. Aromatic compounds could potentially inhibit bacterial growth by precipitating cell wall proteins of *Staphylococcus aureus* NCTC6571 and *Escherichia coli* NCTC 5933 [42]. Seed aqueous extracts inhibiting the strains of *Bacillus cereus*, *Staphylococcus aureus*, *Salmonella typhimurium*, and *Escherichia coli* [43]. Additionally, anise seed oil extracts inhibiting the fungal mycelial growth of *Alternaria alternata*, *Aspergillus niger* and *Aspergillus parasiticus* [40]. Anethole is exhibiting the strong synergism as bacteriostatic with polygodia, an aldehyde phytocompound [44]. The fungal strains of *Candida albicans*, *C. parapsilosis*, *C. tropicalis*, *C. pseudotropicalis* and *C. krusei* are effectively inhibiting by anise seed components [45]. Anethole is blocking the recovery of *S. cerevisiae* and *C. albicans* from fungistatic effects of dodecanol [46]. Plant pathogenic fungal strains significantly inhibited by the *trans*-anethole contained essential oil [47]. *Trans*-anethole disrupts the cell wall integrity of *Bacillus cereus* and promotes the leakage of nucleic acid components [48]. Anethole, an antivirulence compound- inhibiting the *in vitro* expression of cholera toxin and toxin coregulated pilus through by over expressing *crp* genes of cyclic AMP receptor signaling system [49,50].

Conclusion

Essential oil extracts of spices effectively inhibiting or killing the microbial flora of bacterial and fungal species. Hence, food industry is more beneficial to use spices as condiments and its essential components as controlling agents for bacteria and fungi. Active components of spices-essential oils significantly disrupting the biological systems of bacteria and fungi, hence suggested as preservatives in the food industry. Further studies about the antimicrobial mechanisms and their molecular events of spices-essential oil active components were needful.

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References

- Li X, Ma S. Advances in the discovery of novel antimicrobials targeting the assembly of bacterial cell division protein FtsZ. *EJMECH*. 2015;95:1-5.
- Panda D, Bhattacharya D, Gao QH, et al. Identification of agents targeting FtsZ assembly. *Future Med Chem*. 2016;8(10):1111-32.
- Haranahalli K, Tong S, Ojima I. Recent advances in the discovery and development of antibacterial agents targeting the cell-division protein FtsZ. *Bioorg Med Chem*. 2016;24(24):6354-69.
- Domadia P, Swarup S, Bhunia A, et al. Inhibition of bacterial cell division protein FtsZ by cinnamaldehyde. *Biochem Pharmacol*. 2007;74(6):831-40.
- Doyle AA, Stephens JC. A review of cinnamaldehyde and its derivatives as antibacterial agents. *Fitoterapia*. 2019;139:104405.
- OuYang Q, Duan X, Li L, et al. Cinnamaldehyde exerts its antifungal activity by disrupting the cell wall integrity of *Geotrichum citri-aurantii*. *Front Microbiol*. 2019;10:55.
- Wei J, Bi Y, Xue H, et al. Antifungal activity of cinnamaldehyde against *Fusarium sambucinum* involves inhibition of ergosterol biosynthesis. *J Appl Microbiol*. 2020;129(2):256-65.
- Wang Y, Feng K, Yang H, et al. Antifungal mechanism of cinnamaldehyde and citral combination against *Penicillium expansum* based on FT-IR fingerprint, plasma membrane, oxidative stress and volatile profile. *RSC advances*. 2018;8(11):5806-15.
- Qu S, Yang K, Chen L, et al. Cinnamaldehyde, a promising natural preservative against *Aspergillus flavus*. *Front Microbiol*. 2019;10:2895.
- Huang F, Kong J, Ju J, et al. Membrane damage mechanism contributes to inhibition of trans-cinnamaldehyde on *Penicillium italicum* using Surface-Enhanced Raman Spectroscopy (SERS). *Sci Rep*. 2019;9(1):1-0.
- Niu A, Wu H, Ma F, et al. The antifungal activity of cinnamaldehyde in vapor phase against *Aspergillus niger* isolated from spoiled paddy. *LWT*. 2022;159:113181.
- Sharifzadeh A, Shokri H, Katirae F. Anti-adherence and anti-fungal abilities of Thymol and Carvacrol against *Candida* species isolated from patients with Oral candidiasis in comparison with fluconazole and Voriconazole. *JNPP*. 2021;16(1).
- Morcia C, Malnati M, Terzi V. In vitro antifungal activity of terpinen-4-ol, eugenol, carvone, 1, 8-cineole (eucalyptol) and thymol against mycotoxigenic plant pathogens. *Food Addit Contam: Part A*. 2012;29(3):415-22.
- Ahmad A, Khan A, Akhtar F, et al. Fungicidal activity of thymol and carvacrol by disrupting ergosterol biosynthesis and membrane integrity against *Candida*. *EJCMID*. 2011;30(1):41-50.
- Darvishi E, Omid M, Bushehri AA, et al. Thymol antifungal mode of action involves telomerase inhibition. *Med Mycol*. 2013;51(8):826-34.
- Jung KW, Chung MS, Bai HW, et al. Investigation of Antifungal Mechanisms of Thymol in the Human Fungal Pathogen, *Cryptococcus Neoformans*. *Molecules*. 2021;26(11):3476.
- Kowalczyk A, Przychodna M, Sopata S, et al. Thymol and thyme essential oil—new insights into selected therapeutic applications. *Molecules*. 2020;25(18):4125.
- Liang C, Huang S, Geng Y, et al. A Study on the antibacterial mechanism of thymol against *Aeromonas hydrophila* in vitro. *Aquac Int*. 2022;30(1):115-29.
- Helander IM, Alakomi HL, Latva-Kala K, et al. Characterization of the action of selected essential oil components on Gram-negative bacteria. *J Agric Food Chem*. 1998;46(9):3590-5.
- Tian L, Wang X, Liu R, et al. Antibacterial mechanism of thymol against *Enterobacter sakazakii*. *Food Control*. 2021;123:107716.
- Cai R, Zhang M, Cui L, et al. Antibacterial activity and mechanism of thymol against *Alicyclobacillus acidoterrestris* vegetative cells and spores. *LWT*. 2019;105:377-84.
- Liao S, Shang S, Shen M, et al. One-pot synthesis and antimicrobial evaluation of novel 3-cyanopyridine derivatives of (-)- β -pinene. *Bioorg Med Chem Lett*. 2016;26(6):1512-5.
- Leite AM, Lima ED, Souza EL, et al. Inhibitory effect of beta-pinene, alpha-pinene and eugenol on the growth of potential infectious endocarditis causing Gram-positive bacteria. *Brazilian J Pharm Sci*. 2007;43:121-6.
- Macedo DE, Andrade AC, Rosalen PL, et al. Antifungal activity, mode of action, docking prediction and anti-biofilm effects of (+)- β -pinene enantiomers against *Candida* spp. *Curr Top Med Chem*. 2018;18(29):2481-90.

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25. Duman AD, Telci I, Dayisoğlu KS, et al. Evaluation of bioactivity of linalool-rich essential oils from *Ocimum basilicum* and *Coriandrum sativum* varieties. *Nat Prod Commun.* 2010;5(6):1934578X1000500634.
26. Guo F, Chen Q, Liang Q, et al. Antimicrobial activity and proposed action mechanism of linalool against *Pseudomonas fluorescens*. *Front Microbiol.* 2021;12:562094.
27. He R, Chen W, Chen H, et al. Antibacterial mechanism of linalool against *L. monocytogenes*, a metabolomic study. *Food Control.* 2022;132:108533.
28. Hsu CC, Lai WL, Chuang KC, et al. The inhibitory activity of linalool against the filamentous growth and biofilm formation in *Candida albicans*. *Med Mycol.* 2013;51(5):473-82.
29. Mutlu-İngök A, Karbancıoğlu-Güler F. Cardamom, cumin, and dill weed essential oils: Chemical compositions, antimicrobial activities, and mechanisms of action against *Campylobacter* spp. *Molecules.* 2017;22(7):1191.
30. Derakhshan S, Navidinia M, Ahmadi A. Antibacterial activity of Dill (*Anethum graveolens*) essential oil and antibiofilm activity of Cumin (*Cuminum cyminum*) alcoholic extract. *Infect Epidemiol Microbiol.* 2017;3(4):122-6.
31. Lopez-Romero JC, Gonzalez-Rios H, Borges A, et al. Antibacterial effects and mode of action of selected essential oils components against *Escherichia coli* and *Staphylococcus aureus*. *ECAM.* 2015;2015.
32. Tian J, Ban X, Zeng H, et al. The mechanism of antifungal action of essential oil from dill (*Anethum graveolens* L.) on *Aspergillus flavus*. *PloS one.* 2012;7(1):30147.
33. Moro IJ, Gondo GD, Pierri EG, et al. Evaluation of antimicrobial, cytotoxic and chemopreventive activities of carvone and its derivatives. *Brazilian J Pharm Sci.* 2018;53(4).
34. Shahbazi Y. Chemical composition and in vitro antibacterial activity of *Mentha spicata* essential oil against common food-borne pathogenic bacteria. *J Pathog.* 2015;2015.
35. Aggarwal KK, Khanuja SP, Ahmad A, et al. Antimicrobial activity profiles of the two enantiomers of limonene and carvone isolated from the oils of *Mentha spicata* and *Anethum sowa*. *J Flavour Fragr.* 2002;17(1):59-63.
36. Regnier T, Combrinck S, Du Plooy W, et al. Evaluation of *Lippia scaberrima* essential oil and some pure terpenoid constituents as postharvest mycobiocides for avocado fruit. *Postharvest Biol Technol.* 2010;57(3):176-82.
37. Giovana CB, Simone NB, Priscilla DL, et al. Antifungal and cytotoxic activity of purified biocomponents as carvone, menthone, menthofuran and pulegone from *Mentha* spp. *Afr J Plant Sci.* 2016;10(10):203-10.
38. Piras A, Porcedda S, Falconieri D, et al. Antifungal activity of essential oil from *Mentha spicata* L. and *Mentha pulegium* L. growing wild in Sardinia island (Italy). *Nat Prod Res.* 2021;35(6):993-9.
39. Özcan MM, Chalchat JC. Chemical composition and antifungal effect of anise (*Pimpinella anisum* L.) fruit oil at ripening stage. *Ann Microbiol.* 2006;56(4):353-8.
40. Orav A, Raal A, Arak E. *Essential oil composition of Pimpinella anisum* L. fruits from various European countries. *Nat Prod Res.* 2008;22(3):227-32.
41. Chaudhry NM, Tariq P. Bactericidal activity of black pepper, bay leaf, aniseed and coriander against oral isolates. *Pak J Pharm Sci.* 2006;19(3):214-8.
42. Fatima SS, Bushra AM, Zainab SA. Extraction and identification of oil extract from anise (*Pimpinella anisum* L.) seeds and study of its antimicrobial activity. *GJPACR.* 2015;3:1-6.
43. Amer AM, Aly UI. Antioxidant and antibacterial properties of anise (*Pimpinella anisum* L.). *J Egypt Pharm* 2019;18(1):68.
44. Kubo I, Fujita KI. Naturally occurring anti-Salmonella agents. *J Agric Food Chem.* 2001;49(12):5750-4.
45. Kosalec I, Pepeljnjak S, Kustrak D. Antifungal activity of fluid extract and essential oil from anise fruits (*Pimpinella anisum* L, Apiaceae). *Acta Pharm.* 2005;55(4):377-85.
46. Fujita KI, Fujita T, Kubo I. Anethole, a potential antimicrobial synergist, converts a fungistatic dodecanol to a fungicidal agent. *Phytother Res: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives.* 2007;21(1):47-51.
47. Huang Y, Zhao J, Zhou L, et al. Antifungal activity of the essential oil of *Illicium verum* fruit and its main component trans-anethole. *Molecules.* 2010;15(11):7558-69.
48. Naksang P, Tongchitpakdee S, Thumanu K, et al. Assessment of antimicrobial activity, mode of action and volatile compounds of *Etligeria paviana* essential oil. *Molecules.* 2020;25(14):3245.
49. Zahid MS, Awasthi SP, Asakura M, et al. Suppression of virulence of toxigenic *Vibrio cholerae* by anethole through the cyclic AMP (cAMP)-cAMP receptor protein signaling system. *PloS one.* 2015;10(9):e0137529.
50. Zahid MS, Awasthi SP, Hinenoya A, et al. Anethole inhibits growth of recently emerged multidrug resistant toxigenic *Vibrio cholerae* O1 El Tor variant strains in vitro. *J Vet Sci.* 2015:14-0664.

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