# Prevalence of methicillin resistance *Staphylococcus aureus* (MRSA) and methicillin sensitivity *Staphylococcus aureus* (MSSA) among hospitalized Iraqi patients.

## Eptissam Younan Pirko<sup>1</sup>, Nihad Khalawe Tektook<sup>2\*</sup>, Madha Mohammed sheet Saleh<sup>2</sup>, Zeina Anwar Jaffar<sup>2</sup>

<sup>1</sup>Medical College, Dyala University, Iraq

<sup>2</sup>Middle Technical University, Collage of Medical & Health Technology, Baghdad, Iraq

#### Abstract

Background: *Staphylococcus aureus* is an important nosocomial pathogen worldwide, with two major classes; Methicillin resistant *S. aureus* (MRSA) and Methicillin sensitive *S. aureus* (MSSA).

Aim: To compare the distribution frequency and antimicrobial sensitivity of MRSA and MSSA *S. aureus* isolates in different clinical specimen from hospitalized Iraqi patients.

Materials and methods: *S. aureus* isolates from clinical specimens were investigated in 203 hospitalizes patients with wide range of ages during the period from February to May 2017. API and Vitek were used for identification and a panel of antibiotics was used to define the antimicrobial sensitivity of the isolates.

Results: The highest *S. aureus* isolates were from burn swab (35%), followed by urine specimen and blood samples with (30 and 26% respectively). MSSA isolates represents (57.5%) of the total and the rest was MRSA isolates (42.5%). MRSA isolates was higher in burns and wound specimens (485 and 13% respectively) whereas the MSSA isolates were higher in blood, urine and ear specimens (29%, 38% and 3.5% respectively). MRSA were multidrug resistance to 7 antibiotics in comparison to MSSA (only two antibiotics).

Conclusion: MSSA isolate are more common than MRSA in clinical specimens with variable proportions in different clinical specimens. Multidrug resistance was more evident among the MRSA than MSSA.

Keywords: MRSA, MSSA, Antimicrobial test.

#### Introduction

*Staphylococcus aureus* is pathogenic bacteria involved in serious infections which leading to a significant mortality and morbidity in children and adults [1,2]. *Staphylococcus aureus* is classified into two classes known as MethicIlin Resistant *S. aureus* (MRSA) and Methicillin Sensitive *S. aureus* (MSSA), and both associated with the nosocomial infections [3,4]. They cause a wide spectrum of diseases (especially in patients with compromised immune systems) as skin infections, urinary tract infections, intravenous catheters, and others [5,6]. Such pathogenicity is augmented by the existence of colonization factor and numerous other virulence factors in these bacteria [7-9].

Both MRSA and MSSA had acquired many genes that are responsible for resistance to methicillin and other beta-lactam antibiotics [10]. Such feature turns the bacteria into an Accepted on April 19, 2019

important nosocomial pathogen worldwide resistant to many common antibiotics and difficult to be treated [11-13].

#### Subjects, Materials and Methods

#### **Subjects**

203 patients with wide range of ages attending Al-Kindy Teaching Hospital during the period from February to May, 2017 were included in the study.

#### Isolation and identification

*S. aureus* were isolated by primary culture on mannitol salt agar plates (Oxoid) that were incubated for 24 hrs at 37°C and examined for the growth of bacteria and fermentation of mannitol by means of production of yellow colonies [14]. Identification was based on API and Vitek system.

#### **Biochemical test**

Catalase, Coagulase, DNase, gelatin liquefaction, mannitol fermentation, nitrate reduction, clumping factor, oxidase, protease, urease and  $\beta$ -Hemolysis were used in the biochemical tests.

#### **Detection of MRSA**

In a 0.5 equivalent McFarland standard, bacterial isolate suspension was made and lawn culture done on Muller-Hinton agar (MHA) plate. Cefoxitin disc (30 µg) [15] was placed on the culture surface and the plates were incubated at 37°C for 18 h. Growth inhibition zone diameters was measured ( $\leq$  9 mm was reported as resistant whilst  $\geq 14$  mm was considered as sensitive).

#### Antibiotic susceptibility test

S. aureus isolates were tested for their susceptibility to antimicrobial agents by Kirby-Bauer method on MHA (Himedia) [16]. Plates were incubated at 37°C for 18 h. Following the incubation, the diameter of inhibition zone was measured according to the criteria recommended by Clinical and Laboratory Standards Institute [17]. The tested antibiotics in this study were as follows: Cefoxitin (FOX: 30 µg), Ciprofloxacin (CIP: 5 µg), Imipenem (IMP: 10 µg), Penicillin (P: 10 IU), Ceftriaxone (CRO: 30 µg), Erythromycin (E: 15µg), Gentamycin (GN: 10 µg), Methicillin (10µg) and tetracycline (TE: 30µg).

#### Statistical analysis

Data were statistically analyzed using Statistical Analysis System-version 9.1 (SAS) basing on Two-Way Analysis of Variance (ANOVA). The significant level of p-value (P<0.05) was considered for indicating the significance point.

#### **Results and Discussion**

Table 1 indicates that S. aureus isolates were with a high frequency from burn swab (35%), followed by urine specimen and blood samples with (% and 26% respectively), but with a low frequency from both wound and ear swab (7% and 2% respectively). These results correspond to those of Obajuluwa et al. [18] as they reported that 24.6% of the bacterial isolates in wound infection were caused by S aureus. The same was noticed with Srinivasan et al. [19] as they found that the high frequency (33%). of S. aureus isolates was from wounds and burns. On the contrary, Japoni et al. [20] found that S. aureus isolates were reported higher from urine samples (30.6%) and lower from both blood samples and deep wound (14, 13.5% respectively) [20,21]. Japoni et al. [20] found that the highest frequency of S. aureus isolates were in wound and burn infections (54.17%), followed by eye infections and UTIs as (20.8, 16.7% respectively), then ear infection (8.3%). While the results of Nada et al. [21] and Pirko et al. [22] study showed that the highest bacterial isolates were from UTIs infection (42%), whilst results of Sherwal et al. [23] showed that the highest rate of S. aureus isolates were from eyes infections (20.8%) (Figure 1).

Table 1. Number and percent of S. aureus and other bacteria isolated from clinical specimen.

Clinical specimen	Positive isolates				Negative isolates		<b>T</b> ( )		*D
	S aureus		Other bacteria				——— Total		*P value
	No.	%	No.	%	No.	%	No.	%	
Burn	19	35	16	16	10	20.5	45	22	S
Urine	16	30	26	26	17	35	59	29	NS
Blood	14	26	19	19	4	8	37	18	S
Wound	4	7	29	29	13	26.5	46	23	HS
Ear swab	1	2	10	10	5	10	16	8	HS
Total	54	27	100	49	49	24	203	100	NS

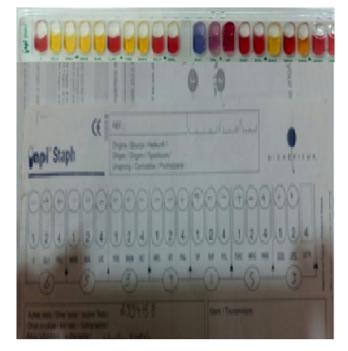
Significant, NS= Non-Significant, HS= Highly Significant

Prevalence of methicillin resistance Staphylococcus aureus (MRSA) and methicillin sensitivity Staphylococcus aureus (MSSA) among hospitalized Iraqi patients



Figure 1. MRSA on chromogen agar.

According to results of API Staph, this system can be used for the diagnosis and identification of *S. aureus* isolates (Figure 2). The API Staph system is a rapid and simple method which used for identification of S aureus, also for differentiation between *Staphylococcus, Kocuria* and *Micrococcus*.



*Figure 2. Results of API staph system of S. aureus (code number 6334 153).* 

Table 2 demonstrated the frequency of MRSA and MSSA isolates from different clinical specimens. The percentage of total MSSA isolates was (57.5%) compared to MRSA (42.5%). MRSA isolates were higher than MRSA in burns (48% vs. 26%) and wounds specimens (13% vs. 3.5%), whereas MSSA isolates where higher than MRSA in urine (38% vs. 17%), blood (29% vs. 22%) and ear specimens (1% vs. 0%). Current results were fully incompatible with the results of Peck et al. [24] and Al-Alem [25] who showed that only (48.6%) of isolates respectively MSSA. The same was true with Al-Hasani [26] and Al-Geobory [27] who found a higher frequency of the MRSA isolates among clinical specimens (83.7% and 90.9% respectively). These differences may due to the different collection site of isolates, the sources of clinical specimens, genetic background and variations in geographic area [28].

Concerning the high frequency of isolates in burns, it could be due to the skin damage as skin represent first line (protective barrier) of defense in immune system and when damaged by burn, the inner layers of skin would exposed to air, increasing the contact with the opportunity pathogenic bacteria as S. aureus and causing infections [22,29,30].

	MRSA		MSSA		Total		*P value
Clinical specimen	No.	%	No.	%	No.	%	
Burn	11	48	8	26	19	35	S
Blood	5	22	9	29	14	26	NS
Urine	4	17	12	38	16	30	HS
Wound	3	13	1	3.5	4	7	HS
Ear swab	0	0	1	3.5	1	2	HS
Total	23	42.5	31	57.5	54	100	S
*S= Significant_NS=	Non-Sid	nificant	HS= F	liahly Sid	nifican	•	

\*S= Significant, NS= Non-Significant, HS= Highly Significant

Biochemical tests include positive results as (100%) for DNase, gelatin liquefaction, mannitol fermentation, nitrate reduction, clumping factor, protease, and positive results as (90%) for  $\beta$ -hemolysis, whilst negative results as (100%) for oxidase and (77%) for coagulase (Table 3).

Table 3. Biochemical test of MRSA and MSSA.

Biochemical test	Results (%)
Catalase	+(100)
Co-agulase	-( 77)
DNase	+(100)
Gelatin liquefaction	+(100)
Mannitol fermentation	+(100)
Nitrate reduction	+(100)
Clumping factor	+(100)
Oxidase	-(100)
Protease	+(100)
Urease	+(100)
ß-Hemolysis	+(90)

As in Table 4, MRSA isolates had exhibited a 100% resistance to both methicillin and penicillin, followed by 89%, 86%, and 85% to ceftriaxon, cefoxitin and tetracycline respectively. For MSSA isolates, they exhibited resistance to both methicillin and penicillin (75% and 70% respectively) and 100% sensitivity to ciprofloxacin, gentamycin and imipenem. These results indicate that MRSA isolates were multidrug resistance (7 antibiotics) compare to MSSA isolates which were resistance to only two antibiotics. Results of Al-Geobory [27], Al-Saadi et al. [31], Tektook et al. [12] and Tektook et al. [32] studies showed similar results.

A	MRSA			MSSA			
Antibiotics	<sup>*</sup> R (%)	I (%)	S (%)	R (%)	I (%)	S (%)	
Cefoxitin	86	0	14	10	0	90	
Ceftriaxon	89	0	11	0	26	74	
Ciprofloxacin	70	25	5	0	0	100	
Erythromycin	84	0	16	10	0	90	
Gentamycin	79	0	21	0	0	100	
Imipenem	11	15	74	0	0	100	
Methicillin	100	0	0	75	0	25	
Penicillin	100	0	0	70	0	30	
Tetracycline	85	0	15	10	0	90	
No of antibiotics resistance	7	_	_	2	_	_	

Conclusion

The highest *S. aureus* isolates was obtained from burn swab followed by urine specimen and blood samples. The MSSA isolates were more frequent than MRSA. All MRSA isolates have high resistance to both methicillin and penicillin compare to MSSA isolates, so MRSA were multidrug resistance compare to MSSA.

### References

- Chittawatanarat K, Jaikriengkrai K, Permpikul C. Thai Society of Critical Care Study group. Survey of respiratory support for intensive care patients in 10 Tertiary hospital of Thailand. J Med Assoc Thai 2014; 97:S8–S14.
- 2. Pirko EY, Tektook NK, Salman HK. Biofilm formation and antibiotic resistance patterns of *Pseudomonas aerogenosa* isolated from different clinical sources. Int J PharmTech Res 2017; 10:725-730.
- Beilman GJ, Sandifer G, Skarda D, Jensen B, McAllister S, Killgore G, Srinivasan A. Emerging infections with community-associated methicillin-resistant *Staphylococcus aureus* in outpatients at an Army Community Hospital. Surg Infect (Larchmt) 2005; 6:87-92.
- 4. Tektook NK. Study the virulence factors and patterns of antibiotics resistance in *Acinetobacter bumannii* isolated from hospitalization patients in Baghdad city. Pak J Biotechnol 2018; 15:23-27.
- 5. Miling Y, Sünje P, Fukuyama J, Hwang J, Peter H, Do-Yeon C, Relman S, David A. Nasal microenvironments and interspecific interactions influence nasal microbiota

complexity and *S. aureus* carriage. Cell Host & Microbe 2014; 14:631-640.

- 6. Abbas MF, Latif R, Tektook NK. Prevalence of gram negative and gram positive pathogenic bacteria in different isolate of patients in Baghdad hospitals. WJPR 2015; 2:123.
- Holmes A, Ganner M, Guane S. *Staphylococcus aureus* isolates carrying Panton-Valentine leukocidin genes in England and Wales: frequency, characterization and association with clinical disease. J Clin Microbiol 2005; 43:2384-2390.
- Essa RA, Hussain SS, Hussain ND, Tektook NK. Prevalence of Staph epidermidis in some hospital of Baghdad city. Bayat Al-Hikma 2015.
- Tektook NK, Al-Lehibi KI, Al-Husseinei RK. Prevalence of some pathogenic bacteria causing UTI in diabetic patients in specialized center for endocrinology and diabetes of Baghdad City–Iraq. Med J Babylon 2017; 14:260-266.
- 10. JH. Methicillin (Oxacillin)-resistant *Staphylococcus aureus* strains. Appl. Environ. Microbiol 2003; 69:116489-116494.
- Lipsky BA, Tabak YP, Johannes RS, Vo L, Hyde L, Weigelt JA. Skin and soft tissue infections in hospitalised patients with diabetes: culture isolates and risk factors associated with mortality, length of stay and cost. US NIH 2010; 53:914-923.
- 12. Hussain SS, Tektook NK, Essa RH. Study of some immunological aspects in patients infected by *Staphylococcus epidermidis* form biofilm. European J Biomed Pharm Sci 2016; 3:153-161.
- Pirko EY, Razooqi BM, Salman HK, Tektook NK. Bacterial isolate isolated from Mobile Medical Staff of Immam Ali Hospital-Baghdad. 2nd international conference in Al-Samawah Technical institute 2018.
- 14. Warren DK, Liao RS, Merz LR, Eveland M, Dunne WM. Detection of methicillin-resistant *Staphylococcus aureus* directly from nasal swab specimens by a real-time PCR assay. J Clin Microbiol 2004; 42:5578-5581.
- 15. Clinical and Laboratory Standards Institute (CLSI). Performance standards for antimicrobial susceptibility testing: Twenty-second Informational Supplement. CLSI document 2012; 100:S22.
- Baur AW, Kirby WM, Scherris JC, and Torch M. Antibiotic susceptibility testing by standardized single methods. AM J Path 1966; 45:493-496.
- 17. Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing: Twentieth Informational Supplement 2010; M100-S20.
- Obajuluwa AF, Onaolapo JA, Oyi AR, Olayinka BO. Susceptibility profile of methicillin-resistant *Staphylococcus aureus* (MRSA) isolates to antibiotics and methanolic extracts of Parkiabiglobosa (Jacq) Benth. BJPR 2013; 3:587-596.
- 19. Srinivasan A, Dick JD, Perl TM. Vancomycin resistance in Staphylococci. Clin Microbiol 2002; 15:430-438.

Prevalence of methicillin resistance Staphylococcus aureus (MRSA) and methicillin sensitivity Staphylococcus aureus (MSSA) among hospitalized Iraqi patients

- Japoni A, Ziyaeyan M, Jmalidoust M, Farshad S, Alborzi A, Rafaatpour N, Badiee P. Antibacterial susceptibility patterns and cross-resistance of methicillin resistant and sensitive *Staphyloccus aureus* isolated from the hospitalized patients in Shiraz, Iran. Braz. J. Microbiol 2010; 41.
- Nada KK, Shaimaa OH, Shatha KK. Bacteriological study of urinary tract infections with antibiotics susceptibility to bacterial isolates among honeymoon women in Al-Qassim hospital, Babylon Province, Iraq. Br Biotechnol J 2013; 3:332-340.
- 22. Pirko EY, Salman HK, Tektook NK. Study of some virulence factors of Coagulase negative *Staphylococci* (CONS) isolated from urinary tract infections (UTI). WJPR 2016; 5:68-78.
- 23. Sherwal BL, Verma AK. Epidemiology of ocular infection due to bacteria and fungus- A prospective study. Lady Hardinge Medical College India 2008; 10.
- 24. Peck KR, Baek JY, Song JH, and Ko KS. Comparison of genotypes and enterotoxin genes between *Staphylococcus aureus* isolates from blood and nasal colonizers in Korean hospitals. J Korean Med Sci 2009; 24:585-591.
- 25. Al-Alem AH. Antibiotic resistant *Staphylococcus aureus* infection studies in hospitals. Middle East Technical University 2008.
- 26. Al-Hasani HM. Comparative study between methicillinresistant coagulase positive and negative Staphylococci. University of Baghdad 2011.
- 27. Al-Geobory HA. Comparative study between Methicillin resistant *Staphylococcus aureus* (MRSA) and Methicillin sensitive *Staphylococcus aureus* (MSSA), and detect the

antimicrobial effects of some plant extracts on them. University of Baghdad 2011.

- 28. Fayad RJ, AL Saade RE, Tektook NK, Mahammed AN. Antimicrobial agent study of bacterial isolates isolated from acute ear infection In Iraqi patients, Al-Mustansiriyah J. Sci 2013; 24:19-24.
- 29. Wong TH, Tan BJ, Ling ML, Song C. Multi-resistant *Acinetobacter baumannii* on burns unit-clinical risk factors and progenosis. Burns 2002; 28.
- Tektook NK, Essa RA, Hussain SS. Evaluation of genotype and phenotype methods for detection of biofilm formed by Staph epidermidis isolated from patients in some hospitals of Baghdad city. WJPR 2015; 4:1835-1854.
- 31. Al-Saadi NT, Kadhim TA. The incidence of methicillin resistant *staphylococcus aureus* in Al-Hussain teaching hospital of Al-samawa city, Iraq and its antibiotics susceptibility. WJPPS 2014, 3.
- 32. Tektook NK. Biochemical study in Diabetic retinopathy patients with and without urinary tract infections. First conference of application research 2018.

#### \*Correspondence to

Nihad Khalawe Tektook

College of Medical & Health Technology

Middle Technical University

Baghdad

Iraq