

## SHORT REPORT

### Mechanisms of RNAi: mRNA cleavage fragments may indicate stalled RISC

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#### ABSTRACT

The molecular mechanism of RNA interference (RNAi) is under intense investigation. We previously demonstrated the existence of inactive siRNAs and also of mRNA cleavage *in vivo* in human cells. Here it is shown that some siRNAs with low activity leave mRNA cleavage fragments while an siRNA with higher activity does not. The pattern is consistent with both short-term (4-24 hours) and long-term (1-4 days) time-series. Analysis of the putative 3' mRNA cleavage product showed high GC content immediately after the cleavage point. The cleavage fragments might indicate a stalled or slowed RNAi cleavage complex - possibly in the RISC enzyme restoration phase - and thus constitute a novel explanation for the existence of inactive siRNAs.

**KEYWORDS:** RNAi, RISC, siRNA, activity, mechanism

#### INTRODUCTION

RNA interference (RNAi) is a cellular process whereby introduced double-stranded RNA can induce degradation of target RNA with sequence similarity (Fire et al, 1998). The double-stranded RNA is broken down into small, ~21-mer RNA (Hamilton and Baulcombe, 1999), by the RNase III-like protein Dicer (Hammond et al, 2000). RNAi has become a very widely used tool for functional genomics after synthetic small interfering RNA (siRNA) was shown to be able to induce RNA degradation also in mammalian cells (Elbashir et al, 2001a).

The small RNA is incorporated into the RNA-induced silencing complex (RISC) (Bernstein et al, 2001). RISC has been shown to be a multi-turnover enzyme (Hutvagner and Zamore, 2002; Haley and Zamore, 2004). The enzyme has to be restored between enzymatic cycles (Haley and Zamore, 2004). RISC consists of several components, some of which are still uncharacterized, and genetic screens in lower organisms have unearthed a long list of candidate RNAi genes (Meister and Tuschl, 2004; Tang, 2005). Target degradation seems to be initiated by mRNA cleavage, both in *Drosophila* and in mammalian cells (Elbashir et al, 2001b; Holen et al, 2002). A central candidate for being the mRNA endonuclease "Slicer" is the Argonaute protein Ago-2 (Meister et al, 2004; Liu et al, 2004; Rivas et al, 2005).

Some siRNAs are inactive with the activity having strong sequence dependence, since a small shift in position of only 6 nucleotides can give rise to an inactive siRNA from a highly active siRNA (Holen et al, 2002). Statistics on large siRNA screening studies showed that active siRNAs had a bias towards a low GC content in the 5' end of the antisense strand (Khvorova et al, 2003). One mechanistic explanation for the existence of inactive siRNAs is that this asymmetry in the ends of the duplex leads to differential loading into RISC of one RNA strand over the other, in *Drosophila* embryo lysates (Schwarz et al, 2003).

The results presented here suggest that another mechanism for inactive siRNAs could be the slow restoration of RISC complexes – and thus the accumulation of mRNA cleavage products – perhaps due to the high GC content in the 5' end of the siRNA.

#### MATERIALS AND METHODS

##### Preparation of siRNA

The 21-mer single stranded RNAs were synthesized at 20 nmol scale by Ambion (Austin, Texas, USA). The RNA strands were resuspended in nuclease-free water (Ambion) and the concentration of the strands was verified by spectrometry using NanoDrop (Saveen Werner). Annealing of the complementary strands was performed by mixing

equal amounts of RNA and briefly heating the mixture to 65°C for 5 min and then allowed to cool to room temperature over 5 min. Annealing was verified by running siRNA on non-denaturing 12% (w/v) PAGE gels, staining with SYBRgold (Invitrogen) and visualizing bands by fluorescent scanning on a Typhoon 9410 (Amersham Biosciences).

#### Cell culture and transfection

HeLa cells were maintained in Dulbecco's minimal essential medium (DMEM) supplemented with 10% (v/v) foetal calf serum (Gibco BRL). The cells were passaged at sub-confluence and plated 24 hr before transfection in 6-well-plates, where each well is approximately 10 square centimeters. Transfections were performed using 1.44 µg siRNA per well and 3.6 µl Lipofectamine2000 (Invitrogen), and thus a ratio of 2.5 of Lipofectamine2000 (in microliters to micrograms of RNA).

#### Isolation of mRNA and northern blots

The mRNA was isolated at various times after transfection using Dynabeads oligo(dT)<sub>25</sub> (Dynal) and then fractionated at 1.5 % (w/v) agarose gels containing 0.8 M formaldehyde for 3 hr at 50 V. The agarose gel was then washed for 2 x 15 min in distilled water before the RNA was blotted onto pre-wetted BrightStar-Plus nylon membranes (Ambion). The Fen1 mRNA was detected using a radioactive probe corresponding to the protein coding region (373-1515, relative to Fen1 mRNA accession number NM\_004111), using random priming with Rediprime II (Amersham Biosciences) and P-32 alpha-CTP (Amersham Biosciences). Hybridization was performed overnight using ExpressHyb (BD Biosciences), with four subsequent washes at room temperature and 50 degrees with Wash Buffer I (2 x SSC + 0.05% , w/v, SDS), and with Wash Buffer II (0.2 x SSC + 0.1%, w/v, SDS), respectively.

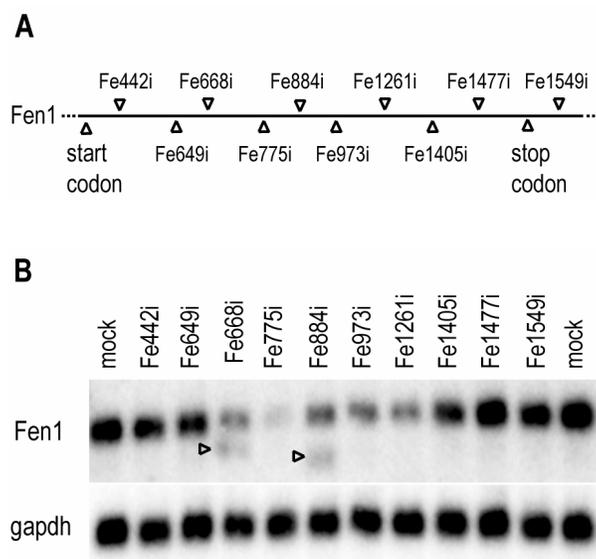
The radioactive signals were visualized with a Phosphor storage Screen and a Typhoon 9410 imager (Amersham Biosciences). Quantification of relative signal strength performed with the ImageQuant software (Nonlinear Dynamics, Amersham Biosciences).

## RESULTS AND DISCUSSION

We previously investigated inactive siRNAs using relatively sensitive and quantitative Northern blots (Holen et al, 2002; Amarzguoui et al, 2003; Holen et al, 2003). While screening for siRNAs against the DNA repair gene Flap-endonuclease 1 (Fen1) in a new study (Figure 1A), faint, 3' cleavage products were observed with two of the siRNA species when harvesting the mRNA at 24 hr post-transfection (data not shown).

The presence of these mRNA cleavage fragments was an interesting observation and, therefore, this was investigated further. When improving the quality of northern blots and accentuating the signal difference between highly active and less active siRNAs by isolating mRNA at 12 hr post-transfection, the cleavage products were clearly observed only in the lanes of two less active siRNAs, Fe668i and Fe884i, while our best candidate, Fe775i, had no such cleavage product (Figure 1B). Our previous hypothesis regarding such cleavage products proposed that cleavage products were due to lack of mRNA-degradation capacity in a down-stream rate-limiting step (Holen et al, 2002). The new data contradict this hypothesis.

The electrophoretic migration of the cleavage fragments was measured. Using the main Fen1 band and the loading control GAPDH, with mRNA sizes at 2.3 kb and 1.3 kb respectively, the sizes of the cleavage products were calcu-



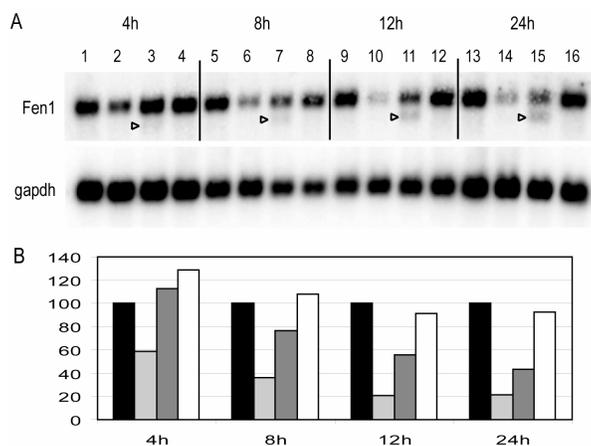
**Figure 1. (A)** Graphical representation of Fen1 mRNA and the siRNA positions. **(B)** Screening of 10 siRNAs against Fen1 at 12 hr post-transfection by northern blot assay. Upper panel: Fen1 target mRNA. Lower panel: GAPDH loading control.

lated to be consistent with primary cleavage at the siRNA-sites (data not shown).

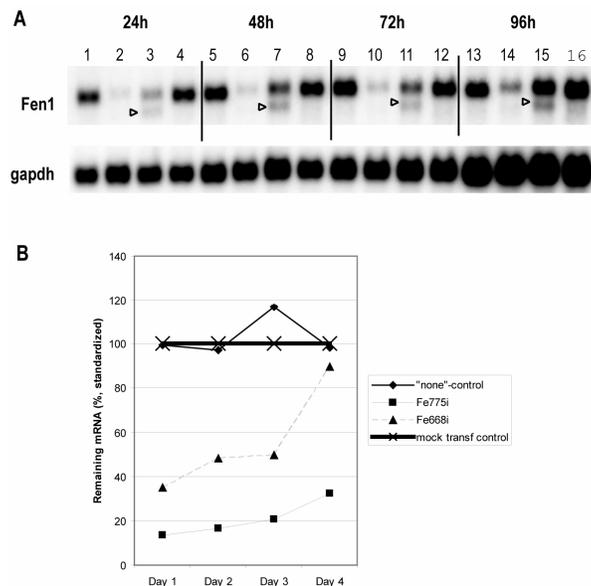
A time-series study was performed to investigate whether the cleavage fragments were of a transient nature. It seemed possible that perhaps Fe775i had produced and already lost such a cleavage fragment while depleting the Fen1 mRNA pool five-fold sometime during the first 12 hr post-transfection. The mRNA was harvested at time points 4 hr, 8 hr, 12 hr and 24 hr for Fe775i and Fe668i (Figure 2). At no time was a cleavage product observed in the

Fe775i-samples, while with Fe668i a gradual build-up of cleavage product was observed (Figure 2).

While studying the long-term silencing over several days, it was found that the main Fen1 mRNA band recovered to full strength for the weaker siRNA candidate, Fe668i, while our best siRNA candidate, Fe775i, retained more than 60% silencing even after 4 days (Figure 3). At 48 hr the cleavage product was in fact much stronger than the main band of the Fe775i-sample (Figure 3).



**Figure 2.** Short-term time-series. **(A)** Primary northern blot data. Upper panel: Fen1-probe. Lower panel: GAPDH loading control. Lanes 1-4 are mock transfection, Fe775i, Fe668i and ssRNA control, respectively, harvested at 4 hr post-transfection. Similarly for lanes 5-8, 9-12 and 13-16, for harvest at 8 hr, 12 hr and 24 hr, respectively. **(B)** Quantified and standardized results: Shown is Fen1 expression as a function of GAPDH loading control, standardized to mock transfection sample at each time point. Black columns; mock transfection control; Light grey columns; Fe775i; Dark grey columns; Fe668i; White columns; ssRNA transfection control.



**Figure 3.** Long-term time-series. **(A)** Primary northern blot data. Upper panel: Fen1-probe; Lower panel: GAPDH loading control. Lanes 1-4 are samples with no treatment ("none"), Fe775i, Fe668i and mock transfection, respectively, for samples harvested at day 1 (24 hr after transfection); similar order for lanes 5-8, 9-12 and 13-16 for day 2, day 3 and day 4, respectively. **(B)** Quantified and standardized results: shown is Fen1 expression as a function of GAPDH loading control, standardized to mock transfection sample at each time point.

We also observed in a previous study that the mRNA depletion by RNAi seems to be finely balanced with mRNA production (Holen et al, 2002). The cleavage product continues to be produced even after the net mRNA silencing has been overwhelmed by mRNA transcription. As the last Fe668i-silencing is lost between 72 hr and 96 hr, cleavage product continues to build up (Figure 3). This demonstrates that putatively inactive siRNAs can also have some activity, although too low compared to the mRNA transcription rate to affect the steady-state concentration of mRNA. If the cleavage product is associated with stalled RISC – as would seem to be necessary to protect a cleaved mRNA from general degradation – this would be a novel mechanism resulting in less active siRNA species, such as Fe668i and Fe884i.

Using Martinez and Tuschl's proposed cleavage point (after the 10<sup>th</sup> nucleotide counting from the 5' end of the antisense strand) (Martinez and Tuschl, 2004), the GC-content was tabulated and analyzed in the putative 3' mRNA-cleavage products, finding that Fe668i and Fe884i (but not Fe775i, nor Fe973i and Fe1261i, two other weak siRNAs) had a high GC content in the first 8 base-pairs after the cleavage point (Figure 4).

	Cleavage point	Cleavage product?	GC-ratio (nearest 8 nt frg.)
Fe668i	CGCAGUGAG...CGGCGGGCUGAG	Yes	8/8
Fe775i	GCAGCACA...UGAUGAGUGCAA	No	3/8
Fe884i	GUCUAUGCU...CGGCUACCGAG	Yes	6/8
Fe973i	GCUGCCA...CCAGGAUUCCA	No	4/8
Fe1261i	GUGGAGCGA...GCCAAUGAAGA	No	4/8

**Figure 4.** Diagrams of putative mRNA fragment ends after siRNA cleavage. Cleavage point as defined by Martinez and Tuschl (2004). Shown are sense sequences for the five siRNAs with considerable cleavage activity at 12 hr post-transfection.

Efforts to find stable hairpin structures – an alternative hypothesis for the existence of cleavage products – in the cleaved mRNA were fruitless. One might postulate such a stabilizing hairpin structure somewhere downstream of the cleavage points of Fe668i and Fe884i, but if so, it would be reasonable to expect the upstream siRNA of Fe775i to also produce the same cleavage products. However, Fe775i does not produce such cleavage products. Furthermore, some results from our previous study can now be reinterpreted in the light of these new observations. Two central GC-pairs were mutated, with a double and single mutation, hTF167-M1 and hTF167-M2 (Holen et al, 2002). These mutations reduced the mRNA activity but the single and double mutations still retained approximately 80 and 70 percent mRNA depletion, respectively. Both the single and the double mutation were situated 5' of the putative cleavage point. Interestingly, the cleavage product was partly lost with M1, and completely lost with M2.

At the time we interpreted the loss of cleavage product as a consequence of the lower silencing capacity and in the light of the hypothesis of a saturated downstream rate-limiting step in the RNAi mRNA depletion process. How-

ever, now this loss of cleavage products can be reinterpreted to support the hypothesis of RISC restoration being dependent on the binding strength of the antisense siRNA-mRNA duplex after cleavage.

## CONCLUSIONS

In conclusion, these observations suggest that the strand-selection hypothesis (Schwarz et al, 2003) may not be the sole explanation for the inactivity of siRNAs. The cleavage fragments observed with a series of siRNAs indicate that the restoration of RISC complexes after cleavage of mRNA can also be a rate-limiting process, which seems to be affected by the siRNA sequence, possibly the GC-content of the 5' end of the antisense siRNA strand.

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## STATEMENT OF COMPETING INTERESTS

The author declared no competing interests.

## REFERENCES

- Amarzguioui M, Holen T, Babaie E and Prydz H. 2003. Tolerance for mutations and chemical modifications in a siRNA. *Nucleic Acids Res*, 31, 589-595.
- Bernstein E, Caudy AA, Hammond SM and Hannon GJ. 2001. Role for a bidentate ribonuclease in the initiation step of RNA interference. *Nature*, 409, 363-366.
- Elbashir SM, Harborth J, Lendeckel W, Yalcin A, Weber K and Tuschl T. 2001a. Duplexes of 21-nucleotide RNAs mediate RNA interference in cultured mammalian cells. *Nature*, 411, 494-498.
- Elbashir SM, Lendeckel W and Tuschl T. 2001b. RNA interference is mediated by 21- and 22-nucleotide RNAs. *Genes Dev*, 15, 188-200.
- Fire A, Xu S, Montgomery MK, Kostas SA, Driver SE and Mello CC. 1998. Potent and specific genetic interference by double-stranded RNA in *Caenorhabditis elegans*. *Nature*, 391, 806-811.
- Haley B and Zamore PD. 2004. Kinetic analysis of the RNAi enzyme complex. *Nat Struct Mol Biol*, 11, 599-606.
- Hamilton AJ and Baulcombe DC. 1999. A species of small antisense RNA in posttranscriptional gene silencing in plants. *Science*, 286, 950-952.
- Hammond SM, Bernstein E, Beach D and Hannon GJ. 2000. An RNA-directed nuclease mediates post-transcriptional gene silencing in *Drosophila* cells. *Nature*, 404, 293-296.
- Holen T, Amarzguioui M, Babaie E and Prydz H. 2003. Similar behaviour of single-strand and double-strand siRNAs suggests they act through a common RNAi pathway. *Nucleic Acids Res*, 31, 2401-2407.
- Holen T, Amarzguioui M, Wiiger MT, Babaie E and Prydz H. 2002. Positional effects of short interfering RNAs targeting the human coagulation trigger Tissue Factor. *Nucleic Acids Res*, 30, 1757-1766.
- Hutvagner G and Zamore PD. 2002. A microRNA in a multiple-turnover RNAi enzyme complex. *Science*, 297, 2056-2060.
- Khvorova A, Reynolds A and Jayasena SD. 2003. Functional siRNAs and miRNAs exhibit strand bias. *Cell*, 115, 209-216.
- Liu J, Carmell MA, Rivas FV, Marsden CG, Thomson JM, Song JJ, Hammond SM, Joshua-Tor L and Hannon GJ. 2004. Argo-

- naut2 is the catalytic engine of mammalian RNAi. *Science*, 305, 1437-1441.
- Martinez J and Tuschl T. 2004. RISC is a 5' phosphomonoester-producing RNA endonuclease. *Genes Dev*, 18, 975-980.
- Meister G, Landthaler M, Patkaniowska A, Dorsett Y, Teng G, and Tuschl T. 2004. Human Argonaute2 mediates RNA cleavage targeted by miRNAs and siRNAs. *Mol Cell*, 15, 185-197.
- Meister G and Tuschl T. 2004. Mechanisms of gene silencing by double-stranded RNA. *Nature*, 431, 343-349.
- Rivas FV, Tolia NH, Song JJ, Aragon JP, Liu J, Hannon GJ and Joshua-Tor L. 2005. Purified Argonaute2 and an siRNA form recombinant human RISC. *Nat Struct Mol Biol*, 12, 340-349.
- Schwarz DS, Hutvagner G, Du T, Xu Z, Aronin N and Zamore PD. 2003. Asymmetry in the assembly of the RNAi enzyme complex. *Cell*, 115, 199-208.
- Tang G. 2005. siRNA and miRNA: an insight into RISCs. *Trends Biochem Sci*, 30, 106-114.
- Tomari Y and Zamore PD. 2005. Perspective: machines for RNAi. *Genes Dev*, 19, 517-529.

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