

***In vitro* antioxidant activity of *Psidium guajava* Linn. by using ethanolic extract fraction of leaves and bark.**

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Abstract

Guava is most popular and commonly available plant all over the India. This study is about the evaluation of *in vitro* antioxidant activity of *Psidium guajava* Linn. The leaves and bark of the *Psidium guajava* Linn. are used for study of the antioxidant activity. Ethanolic extract of these leaves and bark is used for this evaluation study. Nitric oxide and DPPH methods are used to evaluate free radical scavenging activity.

Keywords: Guava, Anti-oxidant activity, Ethanolic extract, Nitric oxide, DPPH method.

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Introduction

Modernisation has made many changes and modifications in the lifestyle of society, resulting in drastic increase of several disorders and diseases. Various studies have reported that consumption of good diet (vegetables, fruits) is necessary in reducing the riskfactor of many diseases [1-3].

Plants have been used for medicinal purpose from ancient times. Among many countries, India is one of the richest ancient repositories of medicinal plants. From the time of immortal Medicinal plants are used in treatment of many diseases, they are considered safe and effective. These remedies are in syn. with nature, which is biggest advantage.

Traditional medicines have deep roots in culture of ancient heritage. Ancient scholars believed that herbs are only solution for many health related problems.

The advances in various scientific field have proved that plants contain active chemical constituents.

The constituents of guava include vitamins, tannins, phenolic compounds, flavonoids, essential oils, sesquiterpene alcohols and tri terpenoid acids. Some of the therapeutic activities of Guava leaf extract are analgesic, anti-inflammatory, anti-microbial, hepatoprotective and anti-oxidant, due to the presence of phenolic group.

Plant Profile

Botanical name: *Psidium guajava* Linn.

Family: Myrtaceae.

Common names: Guava, common guava, yellow guava, apple guava.

Parts used: Fruits, leaf, flower, root and bark.

Materials and Methods

Collection and authentication

Leaves and bark were used for the ethonolic extraction to identify the antioxidant activity from the plant which was collected during the month of july 2010 from vaageswari college

of pharmacy, Karimnagar, Telangana under the guidance and authentication of Dr. Jayaraman, director of Plant aatomy centre (PARC), Tambaram, Chennai. A voucher specimen No. PARC /2010/594 has been deposited for further reference.

Extraction process

Shade dry and coarsley powder the Leaves and Bark of *Psidium guajava* Linn.. Concentrate around 300 gm of powdered medication with ethanol by cold maceration method, after 72 hrs of maceration filtration was followed [4,5]. After entire extraction, the concentrate was focused by refining off the dissolvable and afterward utilizing vaccum flash evaporator. The concentrate was dissipated to dryness under diminished weight. Using high polarity solvents like chloroform, petroleum ether, ethanol the yield was extracted. After which the fractions where evaporated. Its colour and consistencies were also observed Under vaccum. Percentage yield was calculated on air dried basis [6,7].

Phytochemical screening: the concentrate ethonolic extract was subjected to qualitative qualitative phytochemical test for identification of constituents.

Pharmacological Studies

In vitro antioxidant studies

Oxidation is one of the important biological processes for the production of energy in living organism. Living organisms uses oxidation for the production of energy to fuel biological processes [8-10]. A variety of physiological and biochemical lesions increasingly deteriorate degenerative diseases such as aging, cancer and coronary artery disease due to free radicals. Despite of anti-oxidant defence and other defence mechanism in human these systems are insufficient to prevent the damage entirely. Antioxidants are the substances that can inhibit or restrict oxidative cellular oxidizable substrates.

DPPH radical scavenging activity

The free radical scavenging activity is measured in the terms of hydrogen donating or radical scavenging ability, using the stable radical, DPPH.

Preparation of DPPH solution:

About 0.1 mM DPPH crystalline solid was taken in test tube, slowly dissolve the crystalline using organic solvent like methanol to form a solution.

The ethanolic extraction, standard (vitamin c) and control (without the test compound but with an equivalent amount of methanol) with a different concentration (50,100,200,400,800,1000 µg/ml) of about 3 ml each were taken in test tube [11,12]. To this ethanolic extract add 1 ml of DPPH solution slowly. Shake the concentrate solution and enable it to remain at room temperature for about 30 min and the absorbance was measured at 517 nm using a spectrophotometer. The IC₅₀ value (half of inhibitory focus in µg/ml) was compared with standard solution i.e., vitamin C. Free radical scavenging activity is identified by the decrease in the absorbance of the reaction mixture [13-15].

The percentage inhibition of DPPH radical was calculated using the formula,

$$\text{Percentage inhibition (\%)} = \frac{(\text{Abs of control} - \text{Abs of test})}{\text{Abs of control}} \times 100$$

Nitric oxide scavenging activity

Nitric oxide was generated by sodium nitroprusside and measured by Griess reaction. Sodium nitroprusside (5 mM) in standard phosphate buffer saline solution (0.025 M, pH: 7.4) was incubated with different concentrations of ethanolic extract (50, 100, 200, 400, 800, 1000 µg/ml), Vitamin C as reference standard (50, 100, 200, 400, 800, 1000 µg/ml) and dissolved in phosphate buffer saline (0.025 M, pH: 7.4) and the tubes were incubated at 25°C for 5 hr [16-19]. Control experiments without the test compounds but equivalent amounts of buffer were conducted in an identical manner. After 5 hours, 0.5 ml of incubation solution is removed and diluted with 0.5 ml of Griess reagent (1% sulphanilamide, 2% O-phosphoric acid and 0.1% naphthyl ethylene diamine dihydrochloride). The absorbance of the chromophore formed during diazotization of nitrite with sulphanilamide and its subsequent coupling with naphthyl ethylene diamine was read at 546 nm. All the determinations were performed in 6 replicates. Percentage inhibition of nitric oxide radical was calculated by using the formula:

$$\text{Percentage inhibition (\%)} = \frac{(\text{Abs of control} - \text{Abs of test})}{\text{Abs of control}} \times 100$$

Results

Phytochemical screening

A positive remark for saponins, Flavonoids, Phenolic compound and Steroids were identified using phytochemical test in the *Psidium guajava* leaf (Tables 1-5, Figures 1 and 2).

Discussion

Psidium guajava Linn. is classified under family: Myrtaceae. It is a widely spread throughout the India and possess some therapeutic activities like antiseptic, antihelmintic, wound healing and in inflammatory conditions [20,21]. Some of the studies suggest that it contain of high amount of of ethanolic

Table 1. Phytochemical screening of *Psidium guajava* leaf.

Chemical Test	Petroleum ether Fraction	Chloroform Fraction	Aqueous Fraction	Ethanol Fraction
Alkaloids	-	-	-	-
Carbohydrates	-	-	-	-
Glycosides	-	-	-	-
Flavonoid	-	-	+	+
Tannin	-	-	+	+
Terpenoids	-	+	-	+
Oil and fats	-	-	-	-
Steroids	+	+	-	+

(-) indicates absent
(+) indicates present.

Table 2. Phytochemical screening of *Psidium guajava* bark.

Chemical Test	Petroleum ether fraction	Chloroform Fraction	Aqueous fraction	Ethanol Fraction
Alkaloids	-	-	-	-
Carbohydrates	-	-	-	-
Glycosides	-	-	-	-
Proteins	-	-	-	-
Amino acids	-	-	-	-
Saponins	+	-	-	+
Flavonoids	-	-	+	+
Tannin	-	-	+	+
Terpenoids	-	+	-	-
Oil and fats	-	-	-	-
Steroids	+	+	-	+

(+) indicates present.
(-) indicates absent.

Table 3. Phytochemical Screening of *Psidium guajava* Leaf and Bark (Dried powder).

Chemical Test	Dried powder (Leaf)	Dried powder (Bark)
Alkaloids	-	-
Protein	-	-
Amino acids	-	-
Saponins	+	-
Flavonoids	+	+
Tannin	+	+
Terpenoids	-	-
Oil and fats	-	-

Table 4. Free radical scavenging activity of *psidium guajava* bark and leaf of ethanolic extract by DPPH method.

S.NO	Concentration (µg/ml)	% INHIBITION		
		STANDARD (Vitamin-C)	LEE	BEE
1	50	56.438 ± 0.7557	14.94 ± 0.2474**	13.794 ± 0.2322**
2	100	65.55 ± 0.679	20.164 ± 0.4004**	19.338 ± 0.3371**
3	200	70.256 ± 0.8019	32.316 ± 0.6935**	37.298 ± 0.5346**
4	400	73.378 ± 0.7377	42.238 ± 0.7685**	44.128 ± 0.6945**
5	800	76.40 ± 0.7823	48.956 ± 0.5805**	50.186 ± 0.4745**
6	1000	82.36 ± 0.7078	59.706 ± 0.4995**	59.054 ± 0.6795**
7	IC ₅₀	540 (µg/ml)	630 (µg/ml)	590 (µg/ml)

The values are expressed as Mean ± SEM, n=6 in each group. If * P<0.05, **P<0.01 and ***P<0.001 vs. control.

extract substances. Hence the present research emphasis on isolation of ethanolic extract and its evaluation for antioxidant activity. The young leaves juice is used to treat the imbalances of the digestive function. It is also said to be a remedy for the toothache.

Phytochemical study

Various phytochemical screening were carried out for the identification of the various phytoconstituents present in the ethanolic extract fractions

In vitro antioxidant study

Mammalian cells produce nitric oxide in form of free radicals, which is involved in the regulation of various physiological

Table 5. Free radical scavenging activity of *Psidium guajava* bark and leaf of ethanolic extract by Nitric oxide method.

S.NO	Concentration (µg/ml)	% Inhibition		
		Standard (Vitamin-C)	LEE	BEE
1	50 (µg/ml)	8.30 ± 0.45	38.496 ± 0.0534**	41.83 ± 0.0365**
2	100 (µg/ml)	16.66 ± 0.90	41.748 ± 0.1393**	42.72 ± 0.0634**
3	200 (µg/ml)	27.77 ± 1.08	45.18 ± 0.024**	45.34 ± 0.033**
4	400 (µg/ml)	52.77 ± 0.60	46.178 ± 0.030**	46.196 ± 0.025**
5	800 (µg/ml)	61.11 ± 0.51	46.472 ± 0.081**	46.662 ± 0.0316**
6	1000 (µg/ml)	65.88 ± 0.84	47.294 ± 0.048**	47.494 ± 0.0285**
7	IC ₅₀	380 (µg/ml)	560 (µg/ml)	540 (µg/ml)

The values are expressed as Mean ± SEM, n=6 in each group. If * P<0.05, **P<0.01 and ***P<0.001 vs. control.

reactions [22]. Under aerobic conditions nitric oxide is very unstable species. Nitric oxide when undergoes metabolism, produce nitrate and nitrite as an end product through intermediates. It is estimated by using the Griess reagent and in presence of test compound which was the scavenger.

In the present research, sodium nitroprusside in standard phosphate saline buffer at 25°C was incubated to produce nitrite. It was estimated that due to the presence of ethanol, free radical scavenging property can be produced using the above method.

DPPH assay is considered as a valid method to evaluate scavenging activity of antioxidants, since the radical compound is very stable and do not have to generate as in other radical assays. DPPH radicals react with suitable reducing agents and then electrons become paired off and the solutions loses colour stoichiometrically with the number of electrons taken up. such reactivity has been widely used to test the ability of plant extract to act as free radical scavengers. DPPH assay of ethanolic extract showed a dose dependent increase in the percentage of inhibition of free radicals [23,24]. The ethanolic extract fraction was found to show a good antioxidant potential.

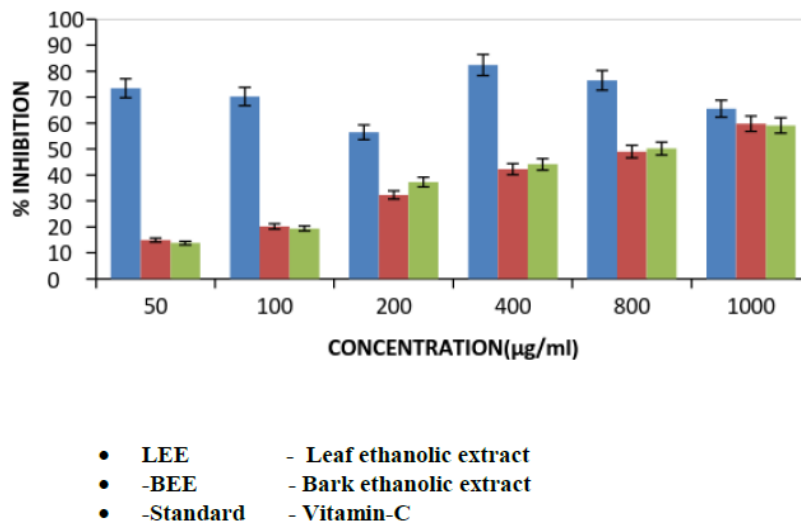


Figure 1: DPPH free radical scavenging activity of STANDARD, LEE and BEE.

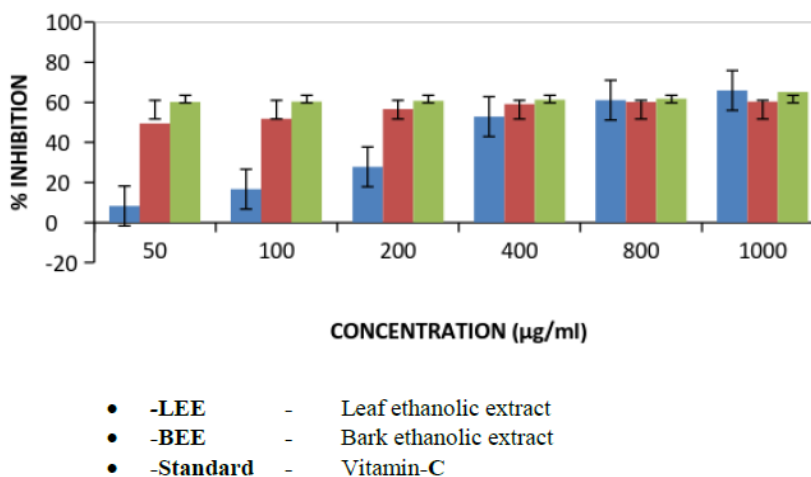


Figure 2: Nitric oxide scavenging activity of STANDARD (vit-C), LEE and BEE.

Conclusion

From this, we can infer that ethanolic extract from leaves and bark of *Psidium guajava* Linn. have huge significant antioxidant activity in all *In vitro* models based on free radical scavenging property. The antioxidant activity, is most likely due to the presence of ethanolic extract.

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