Exploring the Possible Mode of Anxiolytic Action of a Polyherbal Formulation in Animal Models of Anxiety

Shalam Mohamed Hussain^{1*}, Syeda Ayesha Farhana² ¹Department of Pharmacology, College of Pharmacy, Qassim University, Qassim 51452, KSA. ²Departments of Pharmaceutics, Unaizah College of Pharmacy, Qassim University, KSA



The anxiety is managed through several synthetic molecules which come with the burden of adverse and unwanted effects [1]. On the other hand, formulations obtained from natural sources seems to have least pharmacological risks and, claim to cure much the same way as their synthetic counterparts. Among some of the natural drugs employed for this purpose are St. John's wort, Ginseng, Ginkgo Biloba, Kava, Saw Palmetto, Euphorbia Hirta, Valerian and, Convolvulus.

Ayurveda, the traditional healing system utilizing phytomedicines, considers plant as complex mixtures of several components and, assumes that these can provide desired biological action. It also considers that a feebly active plant when compounded with other components or, made pat of a group of plants, can lead to potentiated therapeutic action wherein other plant and, components reinforce the prevalent biological action and, may interact synergistically to neutralize and, counteract any potential side effects that may have occurred because of the single plant use or, which may get developed afterwards after the combination of plants usage. Therefore, several plants with desired biological activities and, as varied undesirable activities suppressants are selected, so that the final componential mixture or, the polyherbal formulation will have a concentrated desired bio-action capacity with the undesired activities being fully suppressed, diluted, under-check or, altogether absent [2,3]. The present study investigates the use of a polyherbal preparation, termed PH-1, consisting of aqueous extracts of leaves of Valeriana wallichii (60%), Plumbago zylanica (15%), Boswellia serrata (20 %) and, Acorus calamus (5%), obtained from the herbalist practitioner. The aqueous extracts were authenticated through presence of marker compounds Valerinic acid, Plumbagin and, Boswellic acid by HPLC. Herein, an attempt has been made to understand the possible mode of action of the combined extracts, PH-1, and, finding a comparison of the extent of the anxiolytic activity of this polyherbal formulation.

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*Corresponding author: Shalam Mohamed Hussain Department of Pharmacology, College of Pharmacy, Qassim University, Qassim 51452, KSA. +966550911846

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MATERIALS AND METHODS

Animals

The experiments were performed on male Swiss albino mice (25-30 g). The animals were group housed in colony cages at ambient temperature of 25 ± 10 C and 45 to 55 % relative humidity with a 12 hr/12 hr light/ dark cycle. They had free access of food and water *ad libitum*. The experiments were carried out between 0900 to 1400 hrs and animals were acclimatized for one week. All the experiments were conducted in a sound proof dimly lit room designed for psychopharmacological studies. The experimental protocols were approved by the Institutional Animal Ethics Committee and, were in accordance with the guidelines of the Committee for the Purpose of Control and Supervision on Experiments on Animals.

Drugs and Chemicals

Diazepam was prepared in water for injection through i.p administration.

Flumazenil, picrotoxin and bicuculine were purchased from Sigma Chemical Co.; St. Louis, USA and, polyherbal formulation was prepared as a concentrated mixture of aqueous extracts of the plant material as semi-solid. Flumazenil, bicuculine and, picrotoxin were suspended in 2 % DMSO-water mixture, 0.1 N HCl and, ethanol-water mixture respectively and, were administered through i.p route. Control animals were given saline at a dose of 10 ml/ kg B. W. of mice. All the other chemicals used in the study were of analytical grade.

Method of administration

In all the experimental models, the antagonists and herbal formulations were administered once daily for 15 days. On 15th day, experiment was performed one hour after the last dose of PH-1 and, half an hour in case of diazepam. In groups where antagonist was combined, antagonist drug was given half an hour before the respective treatments.

Experimental design

Acute toxicity test

Acute toxicity of the preparation was determined using female albino mice. The animals were fasted 3 hrs prior to the experiment. Animals were observed for 48 hrs for any mortality following administration of the different doses of PH-1 as per the guidelines. They were also observed for any behavioral or, gross morphological changes.

Stair case test in mice

The staircase apparatus was made up of plywood and consisted of five identical steps with 2.5 cm high, 10 cm wide and 7.5 cm deep surrounded by a glass wall of 12.5 cm height (Vogel, 2000). At the end of treatment period animals were placed singly on the floor of a transparent box facing away from the steps. The numbers of steps climbed and number of rearing over a period of 5 min were recorded. A step is considered to be climbed if the animal places all its four limbs on the step. After exploration by each animal the steps were cleaned thoroughly with 5% alcohol to eliminate potential cues such as fecal pellets and

urination left by the previous test animal. All animals were tested only once [4].

Mirror Chamber Test

The mirror chamber consisted of a mirrored cube which is open on one side and is placed inside an open top square wooden box. The mirrored cube is constructed of five pieces of mirrored glass with one mirrored side and an opposite side painted dark brown measuring 30 cm on all sides. The three mirrored side panes, a top pane, and the floor pane faced the interior of the cube. The container wooden box was 40 X 40 X 30.5 cm. The mirrored cube was placed in the center of this container so as to form a 5 cm passage around the mirror chamber. A mirror was also placed on the container wall so that it faced the single open side of the mirrored chamber. Except for this one mirrored portion on the container wall, all portions of the container walls were dark brown. Mice were placed individually in the chamber of mirrors and scored once. At the start of the experiment, the mouse was placed in a single, fixed starting point in a corner of the chamber. The animal was allowed to explore the chamber for 5 min and the following parameters were recorded, (1) Latency to enter the chamber (the time for the first entry into the chamber), (2) Number of entries, (3) Total time spent in the chamber The criterion for entry into the chamber was all four feet being placed on the floor panel of the mirrored chamber. After exploration by each animal the floor was cleaned thoroughly with 5% alcohol to eliminate potential cues such as fecal pellets and urination left by the previous test animal [5-7].

Open field test

The open field was made up of wood with dimensions 45 $cm \times 45$ cm with a wall 30 cm high, the floor was divided into equal sixteen squares, four in the centre and twelve in the periphery. The square in the periphery (adjacent to the wall) is designated as the peripheral zone (safe zone) and the squares in the centre are designated as the central zone (vulnerable zone, open space or anxiogenic zone). The time spent in the "safe" peripheral zone is thought to be proportionate to the level of anxiety in the animal [8]. This test uses the animal's repugnance for the central zone to quantify anxiety behavior. Apparatus was placed in a sound attenuated room, 48 cm above the floor and, was cleaned thoroughly with 5% alcohol before placing each animal for observation to eliminate the possible bias due the odor left by the previous animal. Animal was placed in the centre of the apparatus facing one of the sides at the start of experiment and behavior was recorded for 5 min[9]. During this period central and, peripheral crossings, vertical rearing (number of times the animal stood on its hind legs), grooming, defecation, total locomotion (number of units entered on the floor), defecation units (number of bolls) and Immobility times were recorded.

Statistical analysis

All the data obtained from the experiments were subjected

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to the statistical analysis by using Graph pad prism v 5.01 (Demo version) and Instat 3.

RESULTS

Staircase apparatus test

Mice treated with PH-1 (200, 400 and 600 mg/kg), showed significant increase in number of steps climbed and rearing dose dependency when compared to control. However, PH-1 at 800 mg/kg showed decrease in number of steps climbed as compared to control. The mice treated with PH-1 (200, 400 and 600 mg/kg), showed significant increase in number of steps climbed and, rearing dose dependently when compared to control. However, PH-1 at 800 mg/kg showed decrease in number of steps climbed and, rearing dose dependently when compared to control. However, PH-1 at 800 mg/kg showed decrease in number of steps climbed as compared to control.

to control. Flumazenil and, picrotoxin significantly blocked the effect of PH-1. However, Bicuculine pretreatment did not alter the outcome of test formulation, PH-1 (Fig 1).

Mirror chamber test

Treatment with PH-1 resulted in decrease in latency to enter the chamber and also enhanced the number of entries when compared to control and, it was comparable to standard diazepam. The mice treated with PH-1 showed significant increase in the number of mice's entry and, time spent in the chamber according to dose manipulation as compared to the control. Flumazenil and picrotoxin significantly blocked the effects of PH-1 on number of entries and time spent in the chamber (table 1).



Values are expressed as mean \pm S.E.M (n = 06). ^{ns}- statistically non-significant, *<0.05, **p < 0.01, when compared to controld group. #p<0.05, ##p < 0.01, when compared to respective drug alone treated group.

lowing treatment for 15 days							
Groups	Latency in sec	No. of entries	Time spent in the chamber				
Control	247 ±12.5	2.85±0.031	13 ± 1.2				
Diazepam	88±11.2	16 ±1.4	124 ±11.1				
PH-1 100 mg/kg	125±13.3	07 ±0.5*	98 ±10.3*				
PH-1 200 mg/kg	109±11.7	11 ±1.1*	$102 \pm 10.5^*$				
PH-1 200 mg/kg PH-1 400 mg/kg	90±09.1	15 ±1.7**	$114 \pm 11.11^*$				
PH-1 800 mg/kg	189±12.6	3.2 ± 0.2 ns	53.6 ±05.5				
Flumazenil	251±22.3	03 ± 0.1	23 ± 1.5				
Flumazenil + Diazepam	210±21.2###	$04.5 \pm 0.12 \# \# \#$	51.2±2.5##				
Flumazenil + PH-1	231±21.3###	$3.1 \pm 0.01 \# \# \#$	42.4±3.9##				
Bicuculine 1 mg/kg b.w. i.p	211 ± 11.6	3.45 ± 0.02	14 ± 1.6				
Bicuculine + Diazepam	83±11.7 ^{ns}	14 ± 2.5 ns	$131 \pm 8.6^{\text{ns}}$				
Bicuculine + PH-1 400	97±5.6 ns	$17 \pm 1.6^{\text{ns}}$	111 ± 11.51 ns				
Picrotoxin 1 mg/kg b.w, i.p	263 ±22.1	2.4±0.04	10 ± 3.12				
Picrotoxin + Diazepam	222 ±18.5###	5.81±0.041###	16 ± 1.7 ###				
Picrotoxin + PH-1 400	231 ±14.6###	3.34±0.045###	$15 \pm 2.1 \# \#$				

Values are expressed as mean \pm S.E.M (n = 06). ^{ns}- statistically non significant, *<0.05, **p < 0.01, ***p < 0.001 when compared to controld group. #p<0.05, ##p < 0.01, ###p < 0.001 when compared to respective drug alone treated group.

Groups	Rearing	No. of squares Crossed	No. of fecal pellets	Grooming
Control	0.6 ± 0.04	23.60±3.28	5.1 ± 0.64	0.5 ± 0.22
Diazepam	3 ± 0.64	64.60±7.14	2.7 ± 0.26	3 ± 0.051
PH-1 100 mg/kg	0.9 ± 0.03 ns	$25.8 \pm 8.3^{3n}s$	5 ± 0.66 ns	1.1 ± 0.3^{7n} s
PH-1 200 mg/kg	$2 \pm 0.44^{*}$	49.6 ± 8.05 ns	4.5 ± 0.61^{ns}	$2.7 \pm 0.96^{*}$
PH-1 400 mg/kg	$3 \pm 0.40^{*}$	$57.5 \pm 6.95^{*}$	$3.3\pm0.8^{4n}s$	$2.9 \pm 0.1^{6*}$
PH-1 800 mg/kg	1 ± 0.32	33 ± 0.40	4 ± 0.060	1.9 ± 0.040
Flumazenil	2 ± 0 . 05	2 3 ± 2 . 40	3 ± 0.040	1.6 ± 0.060
Flumazenil + Diazepam	1 ± 0.03##	34.33 ± 3.30#	4.3 ± 0.03	0.6 ± 0.04###
Flumazenil + PH-1 400	1.2 ± 0.0 2	27.4 ± 2.43**	5.4 ± 0.40	1.2 ± 0.0 2
Bicuculine 1 mg/kg b.w, i.p	2.33 ± 0.04	21.15±3.5 2	3.5±0.15	2.31±0.04 1
Bicuculine + Diazepam	$3.45 \pm 0.17^{\text{ns}}$	55.55±11.4 3	3.12±0.06	2.9±0.0 3
Bicuculine + PH-1 400	2.91 ±0.05 ^{ns}	49.14±6.21 ⁿ S	4 . 1 ± 0 . 1 4 ⁿ S	2.8±0.01 ⁿ S
Picrotoxin 1 mg/kg b.w, i.p	1.93 ± 0.06	21.15±2.5 2	1.77±0.015	1 . 9 2 ± 0 . 0 4 1
Picrotoxin + Diazepam	2.11 ± 0.012 ^{ns}	31.22±1.33#	1.93±0.06 ⁿ S	0.93±0.03##
Picrotoxin + PH-1 400	1.95 ± 0.06 ⁿ S	41.59±5.55#	2.22±0.09	2.55±0.0 2

Table: 2. PH-1 and, antagonists effects on behavioral patterns in open field test after 15 days of treatment in mice

Values are expressed as mean \pm S.E.M (n = 06). ns- statistically non significant , *<0.05, **p < 0.01, ***p < 0.001 when compared to controld group. #p<0.05, ##p < 0.01, ###p < 0.001 when compared to respective drug alone treated group

Open field test

PH-1 caused a dose dependent increase the number of crossings, rearing and, grooming in the open field paradigm which reflected the enhanced exploratory activity and, the condition of reduced fear when compared to control. The biological effects produced by flumazenil and, picrotoxin were in line with the observations made in the study (Table 2).

DISCUSSION

Several central receptor systems have been suggested in the modulation of fear and, anxiety. The GABA receptor is one of the prominent anxiolytic activities modulating receptor site [2]. The antagonists bicuculine, flumazenil and, picrotoxin working on different sites of this receptor were chosen to find out comparative binding and, study the detailed biological interaction outcomes in comparison with the investigational polyherbal formulation, PH-1.

The results obtained with the investigational drug, PH-1, clearly indicated the anxiolytic activity profile. A dose response effect curve was constructed by using different doses of PH-1 (100, 200, 400, 600 and 800 mg/kg BW). The dose which showed prominent anxiolytic effect (400 mg/kg) was selected for further study to explore the mechanism.

The mechanisms underlying the anxiolysis produced by any drug is multifaceted, as several reports which claim to be involved in the activity, to name a few, are GABAergic, serotonergic and noradrenergic systems which are prominent ones in the brain. Add to the problem of complexity, neural regulatory mechanisms are abundant, regulating one another in the brain, or in some cases themselves. The mechanism involved in the anxiolytic-like effect of PH-1 was studied by using various antagonists for GABA receptor system e.g., flumazenil (GABAAbenzodiazepine binding site antagonist), picrotoxin (GABAA-chloride channel complex antagonist) and bicuculine (GABAA-GABA binding site antagonist).

The flumazenil produces no marked behavioral effects, but can reverse almost all the pharmacological actions of anxiolytics belonging to benzodiazepine group. In our study too Flumazenil per se caused no obvious effect in mice submitted to the staircase, mirror chamber and OFT tests. However it showed a significant antagonistic effect on the anxiolytic outcome induced by diazepam as well as by the PH-1 (400 mg/kg). This might suggest that anxiolytic effect produced by PH-1 due to in part by interaction with central benzodiazepine binding site on GABAA-receptors. Furthermore, pretreatment with the bicuculine, did not alter the effect of either of the antagonists and, thus ruling out the possibility of interaction with the GABA binding site on the GABAA receptors.

However, picrotoxin resulted in the complete blockade of effects of PH-1, supplementing the possibility of involvement of interaction with GABAA chloride channel complex along with benzodiazepine binding site. Thus it is likely that formulation's anxiolytic effect is due to the blockade of benzodiazepine site/chloride channel complex of GABA receptor system. However, a cursor glance at the literature available on some of the components of this formulation could help to uncover the possible mechanism of anxiolytic activity of PH-1. Valeriana was reported to have anxiolytic, tranquilizing, and sleep inducing effects. These effects were claimed to be produced by an interaction with central GABA receptors [10-14]. In an *in vitro* binding study to GABA receptors, Valerian was found to displace the agonist, muscimol, suggesting its binding

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to these receptors. The Valerian extract was also found to contain GABA and other amino acids. Thus these studies coupled with the present findings suggested the possibility of interaction of PH-1 with the benzodiazepine binding site or, GABA-chloride channel receptor complex excluding GABA-b in its anxiolytic effect [15-18].

CONCLUSION

It can be concluded that PH-1 has promising anxiolytic activity and, works through occupation of benzodiazepine binding site and, facilitation of interdependent chloride channel complex. However, further studies are required to ascertain the action of each marker component and, polyherbal extract with varying composition of plants and marker products in these plants.

REFERENCES

1. Lundegaard PR, Anastasaki C, Grant NJ, Sillito RR, Zich J, Zeng Z, Paranthaman K, Larsen AP, Armstrong JD, Porteous DJ, Patton EE. MEK Inhibitors Reverse cAMP-Mediated Anxiety in Zebrafish. Chem Biol. 2015 S1074-5521(15)

2. Hamann J, Linde K, Schweiger HD, Kusmakow O, Förstl H. Overthe-counter-drugs for the treatment of mood and anxiety disorders--the views of German pharmacists. Pharmacopsychiatry. 2014;47 (3):84-8

3. Shalam Md, Shantakumar SM, Laxmi Narasu M. Possible anxiolytic activity of Trans-01, a polyherbal formulation in mice. Pharmacologyonline 1: 138-145 (2007).

4. Shalam Md, Shantakumar SM, Laxmi Narasu M. Pharmacological and biochemical evidence for the antidepressant activity of a herbal preparation, Trans-01. Ind. journal of Pharmacology 39(5) 2007 231-234

5. Vandenbogaerde A, Zanoli P, Puia G, Truzzi C, Kamuhabwa A, De Witte P, Merlevede W, Baraldi M. Evidence that total extract of Hypericum perforatum affects exploratory behaviour and exerts Anxiolytic effects in rats. Pharmacol. Biochem. Behav.2000, 65: 637-633.

6. Gopala Krishna HN, Sangha RB, Misra N, Pai MRSM. Antianxiety activity of NR-ANX-C, a polyherbal preparation in rats. Ind. J. Pharmacol. 2006; 38:330-5.

7. Vogel H, Gerhard, Vogel Wolf gang H. (Eds) Drug discovery and

evaluation pharmacological assays (Springer). 2008.

8. Joshi D, Naidu PS, Singh A, Kulkarni SK. Reversal of triazolam tolerance and withdrawal-induced hyperlocomotor activity and anxiety by bupropion inmice. Pharmacology. 2005 Oct; 75(2):93-7.

9. Lang AP, de Angelis L. Experimental anxiety and antiepileptics: the effects of valproate and vigabatrin in the mirrored chamber test. Methods Find Exp Clin Pharmacol. 2003; 25(4):265-71.

10. Rajesh K Goel, Amanpreet Singh, Pattipati S Naidu, Mohinder P Mahajan, Shrinivas K Kulkarni. Pass assisted search and evaluation of some azetidine-2-ones as CNS active agents. J Pharm Pharmaceut Sci. 2005; 8(2):182-189.

11. Sollozo-Dupont I, Estrada-Camarena E, Carro-Juárez M, López-Rubalcava CJ. GABAA/benzodiazepine receptor complex mediates the anxiolytic-like effect of Montanoa tomentosa. Ethnopharmacol. 2015 Mar 13;162:278-86..

12. Ene HM, Kara NZ, Barak N, Reshef Ben-Mordechai T, Einat H[.] Effects of repeated asenapine in a battery of tests for anxiety-like behaviours in mice. Acta Neuropsychiatr. 2015 Sep 11:1-7.

 Kakehashi A, Kato A, Ishii N, Wei M, Morimura K, Fukushima S, Wanibuchi H Valerian inhibits rat hepatocarcinogenesis by activating GABA(A) receptor-mediated signaling. PLoS One. 2014; 9(11):e113610.
Becker A, Felgentreff F, Schröder H, Meier B, Brattström A The anxiolytic effects of a Valerian extract is based on valerenic acid.. BMC Complement Altern Med. 2014 28;14:267.

15. Bussmann RW. The globalization of traditional medicine in northern peru: from shamanism to molecules. Evid Based Complement Alternat Med. 2013;2013:291903.

16. Trauner G, Khom S, Baburin I, Benedek B, Hering S, Kopp BP. Modulation of GABAA receptors by valerian extracts is related to the content of valerenic acid. Planta Med. 2008 Jan;74 (1):19-24.

17. Khom S, Baburin I, Timin E, Hohaus A, Trauner G, Kopp B, Hering S. Valerenic acid potentiates and inhibits GABA(A) receptors: molecular mechanism and subunit specificity. Neuropharmacology. 2007 Jul; 53(1):178-87.

18. Neuhaus W, Trauner G, Gruber D, Oelzant S, Klepal W, Kopp B, Noe CR.. Transport of a GABAA receptor modulator and its derivatives from Valeriana officinalis L. s. l. across an in vitro cell culture model of the blood-brain barrier. Planta Med. 2008 Sep; 74(11):1338-44.

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