International Journal of Pure and Applied Zoology

Volume 2, Issue 1, pp: 61-70, 2014

©2013 Rishan Publications

ISSN (Print) : 2320-9577 ISSN (Online): 2320-9585

http://www.ijpaz.com

Research Article

CLIMATE CHANGES AND ITS IMPACT ON FISH WITH REFERENCE TO ANTIBIOTIC RESISTANT ENTERIC BACTERIA AND HEAVY METAL ACCUMULATION

* K. Shameem Rani^{1*}, M. Mumtaz¹ and M. Chandran²

¹Department of Zoology, M.S.S Wakf Board College, Madurai-625 020. ²Department of Zoology, Thiruvalluvar University, Vellore.

Article History: Received 25th November 2013; Accepted 26th March 2014; Published online 26th March 2014

ABSTRACT

Fresh fish (*Mugil cephalus*) is a very important source of protein to the population in our country. This fish may harbor *Salmonella sp.* It may be a source of pathogen to human being. A total of 20 samples (20 muscles and 20 gills) were analyzed. The isolates were exposed to 5 different antibiotics. Most of the isolated were resistances to at least one of the antibiotic. This is a clear indication that sewage effluent causes contamination of marine wildlife. Investigation on the accumulation of heavy metals (Cd, Cu and Cr) was carried out commercially important fish (*Mugil cphalus*). The accumulation was observed in tissues of muscles and gills. The result revealed that the copper and cadmium concentration were highest in the muscle and gills. In the muscle and gill of *Mugil cephalus* the order of accumulation was Copper>Cadmium>Chromium.

Key words: Salmonella sp, heavy metals, fish, muscles, gills.

INTRODUCTION

Today, humankind's activities are altering the world's climate. We are increasing the atmospheric concentration of energy-trapping gases, thereby amplifying the natural "greenhouse effect" that makes the Earth habitable. These greenhouse gases (GHGs) comprise principally, carbon dioxide (mostly from fossil fuel combustion and forest burning), plus other heat-trapping gases such as methane (from irrigated agriculture, animal husbandry and oil extraction), nitrous oxide and various humanmade halocarbons. Change in world climate would influence the functioning of many ecosystems and their member species. The majority of 1.3 billion annual cases of Salmonella – caused human gastroenteritis result from ingestion of contaminated food products such as undercooked beef, pork, eggs, milk, shell fish and fish (pang et al.,1995), (Gomez et al.,1997), (Esaki et al., 2004). Salmonella infections can also be contracted following consumption of fresh fruits or vegetables contaminated by fertilizer (Auxe, 1997). Birds and flies are important vectors for rapid widespread dissemination of Salmonella in the

1996). environment (Davies and Wray, Salmonella withstands a wider variety of stresses associated with environmental fluctuations and may persist in water environment for some time. Salmonella can be disseminated as a result of water currents, underground springs and rain runoff carrying contaminated material (Chao et al., 1987), (Abdelmonem and Dowider, 1990). Like E.coli, Salmonella is constantly released into environment from infected human, farm animals, pets and wildlife Al-Yousuf et al., (2000 a). Pathogenic and potentially pathogenic bacteria associated with fish and shell fish include Mycobacteria, Streptococcus iniae, Vibrio vulnificus, Vibro spp, Aeromonads, Salmonella spp, Shigella and the others (Lipp Rose 1997),(Zlotkin 1989),(Bhaftopadhyay,2003). Human infections by these fish pathogen are usually through contact with infected fish while handling them, water or other constituents of fish life environment (Acha and Szyfres, 2003). The initial microflora on the surface of fish is directly related to the water environment while the flora in the gastrointestinal tract corresponds to the type of food and condition of fish (Liton, 1980).

Municipal untreated sewage, run off and storm water are the most important immediate microbiological pollutants (Kayambo and Sven, 2006). The low standard of health in the gulf of mannar region is caused by a general lack of awareness of good hygiene practices, direct contamination of beach waters through bathing and washing and uncontrolled waste disposal around the shoreline. Other sectors like wildlife. agriculture, forestry, urban and rural settlements implicated have been to contribute microbiological pollution. These activities increase eutrophication process thus creating a vast conducive environment for the survival of microbes which eventually infect fish.

Analysis of fish tissue slurry indicated that fish harvested from landing beaches along gulf are infested with; Salmonella, Shigella and E. coli (Onyango et al., 2008b). Given the prevalence of water and food borne disease; Salmonellosis in marine fishes, it was important that all possible infection routes of the pathogens be investigated and prevention measures recommended. Fishes are major part of the human diet and it is therefore not surprising that numerous studies have been carried out on metal pollution in different species of edible fish. Predominantly, fish toxicological environmental studies have prompted interest in the determination of toxic elements in seafood.

This study aimed to isolate and characterize *Salmonella* from marine water and fish muscle and gills and to determine the concentrations of heavy metals such as Cadmium, Chromium and Copper in fish muscle, collected from Rameswaram and Tuticorin. It is expected that the results of this research will assist in acquiring information about the level of toxic metals in this region.

MATERIALS AND METHODS

1. Collection of sample

Marine fish samples (*Mugil cephalus*) were collected from markets located in two different cities (Rameswaran and Tuticorin). The samples were collected in the month (batch wise) of January-March-I, April-June-II, July-September-III, October -December-IV. An average of 5 fishes was bought and transported to laboratory in plastic container within 4 hours. Then marine water samples were also collected in the same area.

2. Sample processing for microbial analysis

With gloved hands and sterilized knife, the fish was severed into parts (Gill and whole body). 20 grams of each part was grinded with 225ml Buffered Peptone Water (BPW) for 3min. Pellet were obtained by centrifugation at 20°C, 10,000 x g RPM, for 15minutes for fish sample. The pellet was then dissolved into 10ml of BPW.

The inoculums were later streaked on to Salmonella Shigella Agar (SSA Difco), Xylose-Lysine Deoxycholate (XLD Difico) and Bismuth sulfite Agar (BSA – Difco) and were incubated at 37°C for 48hrs. In SSA Salmonella spp. were seen as white or yellow with black spot centrally, in XLD, Salmonella spp. grew as pink color with black centre while in BSA salmonella colony grew as grey black with metallic sheen color.

3. Antimicrobial sensitivity test

The agar diffusion method according to Kirby Bauer guidelines was applied for antimicrobial susceptibility testing of *Salmonella* isolates. Mueller Hinton Agar was used in order to perform this test. Suspension of 0.5ml density was used for inoculation. Total of eight isolates from Rameswaram and Tuticorin fishes were tested for antimicrobial susceptibility. The following antimicrobial agents were used: Amoxycillin, Tetracycline, Penicillin, Streptomycin and Chloramphenicol. The results were tabulated.

4. Quantitative analysis of heavy metals

Sample preparation for heavy metal analysis: About 20.0 g of fresh fish samples were weighed accurately and homogenized then poured in an iodine flask separately; 25 ml of concentrated HNO3 was added into each flask. The iodine flasks were refluxed for 1 hr. at $95^{\circ}\text{C} \pm 5^{\circ}\text{C}$. The sample solutions were cooled and 10 ml of Concentrated HNO3 was added into each flask. The flasks were again were refluxed for about 1 hr. at 95° C $\pm 5^{\circ}$ C. Repeated the process until the digestion was completed. Evaporated the solution to 5 ml. Solutions were cooled and 10 ml of Concentrated HCl was added into each flasks. The solution was kept for refluxed for about 15 min to remove the nitrous fumes. Cooled the digested sample solutions, 20 ml of HPLC grade water was added into each flask. Filtered the digested solution through Whatman filter paper no. 41 into 50 ml volumetric flask and made up to the volume using HPLC grade water. Recovery study was carried out by

fortifying known concentration of standards into pre analyzed sample.

To determine the metals (cadmium, chromium and copper) concentration in the samples, a atomic absorption spectrometer (AAS) was used. All chemical regents were from analytical reagent grade (Merck). All solutions were prepared in deionized water. Calibration standards of each heavy metal were prepared by appropriate dilution of the stock solutions (1000 ppm, Merck). The glassware and plastic containers were acid washed with nitric acid 10% for 24 h and rinsed with double distilled water before use.

RESULT AND DISCUSSION

The method of isolation and media used contributed to the effect of Salmonella sp significance. This proves that different media produce different results and performance. The result obtained of the gills and whole body fish differ because of the sensitivity of different media that was used. From this study, SSA media was more sensitive than BSA (Plate 1). Salmonella spp. was present in all parts of the fish especially the whole body, gills and marine water represented in Table 1 and 2. In the above study, SSA gave more bacterial isolates than the other two XLD and BSA. Dutch et al., (1995) reported that the sensitivity of SSA and BSA were 76.6% and 50.0% respectively. (Michael et al., 2003) showed that SSA presented better conditions for isolation of salmonella sp. colonies, hence eliminating the volume of false positives.

As a result, the better selectivity of the media is responsible for the greater detection of Salmonella sp: majorly when streaked from a selective enrichment that eliminates overgrowth of competitors the method of isolation was largely responsible for the significant difference on Salmonella sp. From this study, Salmonella sp. was found to contaminate different parts of the body. This was supported by the finding (Haltha et al., 1997). That these bacteria would exist on fish's skin, gills and intestine and the most potential reservoir of salmonella spp. was the intestine. Hence, it is highly recommended that cross – contamination of other tissue notably digestive tract during handling or preparation be avoided. This is important for future study in order to know the route of salmonella species transmission from pond to the next food chain supply. The study showed that salmonella was more on the gills than whole body of fish. Salmonella shigella agar proved to be a better selective media than the other two media.

Determinations of Salmonellae by specific media (SSA) in water sample were detected. The highest numbers during one year (January 2012-December 2012) was 11 x 10² in Rameswaram in Table 1. The highest Salmonella sp was detected in muscle and gills of Rameswarm fishes 3.0 x 10^2 and $4.3X10^2$ respectively. The maximum number of Salmonella sp occurred in Tuticorin fish muscle and gills region were 5.2X10² and 7.2X10³ respectively. From this result we concluded that the density of Salmonellae sp the minimum value was recorded in month of January to march (Table 3). Similar results were obtained by Hunter and McDonald number was significantly higher in STE and polluted (1991), Tian et al. (2002) and Hyland et al. (2003) who stated that seawater over unpolluted. Enterobacteriaceae were the faecal indicator bacteria populations normally peak in isolated from gills, skin, and muscles.

In our study, the isolated Salmonella sp are more sensitivity to Sreptomycin, Amoxycillin and Pencillin. But highly resistant pattern was observed in Chloramphenicol and Tetracycline drugs (Table 4, 5 and Plate 2). Antimicrobial agents and their metabolites entering the aquatic environment become highly diluted and therefore of these compounds detection becomes extremely difficult (Kümmerer, 2009). Overuse of antibiotics has led to the emergence of resistant bacteria and consequently caused an imbalance between susceptible and resistant bacteria. This eventually has sub-grouped them into susceptible and resistant variants (Levy, 1994). In addition, the potent killing and growth inhibition of bacteria have increased the number of resistant strains which have ultimately evolved into prominent populations of the microbial flora (Levy, 1992).

Hence, the presence of antibiotic resistant bacteria have been used as bio-indicators of polluted effluents since resistant bacteria can be easily isolated and detected (Al-Bahry *et al.*, 2009). Based on the present results, the coastal area of Gulf of Mannar and Tuticorin its fish are continuously exposed to contaminated discharge. Various types of plastics ,bottles, waste clothes, papers, rusted materials and wasted or unused pharmaceutical compounds were dumped at the area of Gulf of mannar through the ship from

other countries (Srilanka, Malaysia and Myanmar). The microbes of human origin are affecting the marine environment and antibiotic resistant determinants are being transferred to other bacteria in the area. Further studies on the effects of sewage discharge and human contamination and bacterial contamination on the environment and public health are urgently needed in different regions of Indian coastal.

The results of heavy metals in fish sample from Rameswaram and Tuticorin are presented in Figure 1 and 2. The concentration of cadmium was found to be higher in fish gills (1.32µg/g) least in the muscles (0.09µg/g). Chromium had an overall mean value of (1.04 µg/g) in the fish parts. Copper also had its highest value (5.22µg/g) in muscles at Rm3. It however had its lowest concentration in the fish gills (0.09µg/g) followed by in the fish muscles (0.13µg/g). It was generally observed that amongst the fish parts, gills have the highest concentration of heavy metals while the muscles had the lowest concentrations. Heavy metals entering the fish have a possibility to get accumulated in different parts of the body and the residual amount can build up to a toxic level. The fish, Mugil cephalus is economically important and they form a large part of the fish catch in the study

Metal elimination routes are more than uptake routes, however metal accumulation is more rapid than metal elimination probably due to the presence of metal binding proteins in tissues (Kendrick *et al.*, 1992). The accumulation of the metals in liver could be based on the greater tendency of the elements to react with the

oxygen, carboxylate, amino group, nitrogen or sulphur of the mercapto group in the metallothionein protein, which was at highest concentration in the liver (Kendrick et al., 1992). These complexes are slowly redistributed to the renal cortex. Liver has also an important role in storage, redistribution, detoxification or transformation and also serve as an active site of pathological effects induced by contaminants (El-Shahawi, 1996). This study revealed that metal accumulation in gills and liver occurs in higher magnitude than what appeared in the muscle. This is a common finding that is also reported by several investigations (Usero et al., 2003; Dural et al., 2007; Al-Yousuf et al., 2000 b). Because of the presence of high levels of metallothionein protein, liver tissue acts as a target organ for heavy metal detoxification (Yilmaz, 2003; Kraemer et al., 2006; Canli and Atli, 2003; Al-Yousuf et al., 2000 b; Romeo et al., 1999). Gills act as the main site for entry of different kinds of contaminants such as heavy metals due to its continuous contact with the external medium. This organ serves a variety of physiological functions such as respiratory gas exchange, osmoregulation and nitrogen excretion (Hoar and Randall, 1984; Altindag and Yigit, 2005). Therefore, heavy metals may appear in a high level in liver and gill tissues compared to what occurs in muscle. Muscle tissue has lower tendency to accumulate heavy metals (Huang, 2003, Al-Saleh, and N. Shinwari, 2002, Altındag, and S. Yig, 2005, Aucoin et al., 1996). Altindag and Yigit, (2005), Romeo et al., (1999) and Huang, (2003) found higher concentration of heavy metals in liver and gill than in muscle.

Table 1. The number of *salmonella spp*. Occurred in marine water sample.

Water sample	No. of CFU/mL
R1	11 X10 ²
R2	$29 \text{ X} 10^1$
R3	15×10^{1}
R4	12×10^{1}
T1	21×10^{1}
T2	$19 \text{ X} 10^1$
Т3	$5 \text{ X} 10^{1}$
T4	$9 ext{ X} 10^1$

R1, R2, R3, R4-Rameswaram water sample

T1, T2, T3, T4-Tuticorin water sample.

Table 2. Antibiotic sensitivity (mm) patterns of *Salmonella spp*. isolated from water sample.

Antibiotic Compound	R1	R2	R3	R4	T1	T2	Т3	T4
Amoxycillin	5.0	-	5.0	6.1	12	5.0	8.2	-
Pencilline	7.4	15.2	11.2	-	-	-	-	-
Tetracycline	-	12.4	8.4	12	10.2	-	10.8	8.4
Streptomycin	-	10.2	7.2	9.4	-	9.5	-	12.8
Chloramphenicol	5.2	-	8.4	-	-	-	-	7.6

Table 3. The number of *Salmonella* bacteria present in marine fish sample.

Sample month/Number -	Ram	eswaram	Tuticorin			
	Muscle	Gills	Muscle	Gills		
Jan-Mar-I	$3.0 \text{X} 10^2$	2.8×10^3	$4.6 \mathrm{X} 10^2$	7.2×10^3		
Apr-Jun-II	2.5×10^{2}	3.1×10^2	5.0×10^2	6.4×10^3		
Jul-Sep-III	2.0×10^{2}	3.6×10^2	5.2×10^2	5.9×10^3		
Oct-Dec-IV	2.8×10^{2}	4.3×10^2	$4.0 \text{ X} 10^2$	$6.0 \text{ X} 10^3$		

Table 4. Antibiotic sensitivity (mm) patterns of *Salmonella spp*. Isolated from Rameswaram fish (Muscle and gills) sample.

Antibiotic	Rm1	Rm2	Rm3	Rm4	RG1	RG2	RG3	RG4
compound								
Amoxycillin	-	8.5	8.0	-	7.0	6.3	-	12.2
Pencillin	10	-	-	8.4	-	6.8	-	11.0
Tetracycline	8.3	6.5	-	5.3	-	-	12.5	9.0
Streptomycin	9.0	4.8	-	8.0	11	-	-	8.8
Chloramphenicol	8.0	-	6.5	-	-	-	-	-

Rm1, Rm2, Rm3, Rm4- Fish muscle of Rameswaram.

RG1, RG2, RG3, RG4-Fish gills of Rameswaram.

Table 5. Antibiotic sensitivity (mm) patterns of *Salmonella* sp isolated from Tutucorin fish (Muscle and gills) sample.

Antibiotic compound	Tm1	Tm2	Tm3	Tm4	TG1	TG2	TG3	TG4
Amoxycillin	-	-	-	6.7	8.7	-	-	5.4
Pencilline	6.9	-	4.0	7.3	-	4.2	-	-
Tetracycline	7.8	-	10.0	-	-	-	-	-
Streptomycin	12.5	8.5	-	9.3	9.2	6.2	-	8.4
Chloramphenicol	-	-	-	12.6	4.0	-	8.8	7.9

Tm, Tm2, Tm3, Tm4-Fish muscle of Tuticorin.

TG1, TG2, TG3, TG4-Fish gills of Tuticorin.



Plate 1: Isolation of Salmonella sp from fish by using SS agar medium.

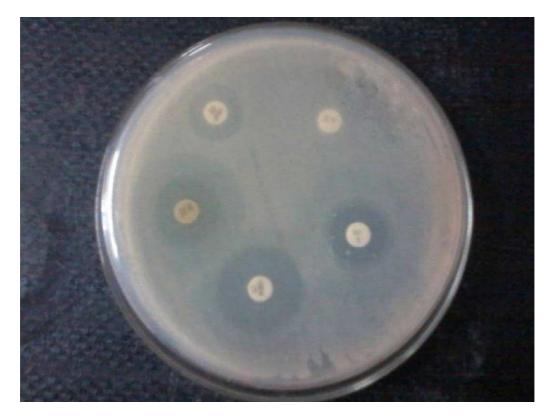


Plate 2: Antibiotic sensitivity (mm) patterns of *Salmonella sp* isolated from fish (Muscle and gills) sample.

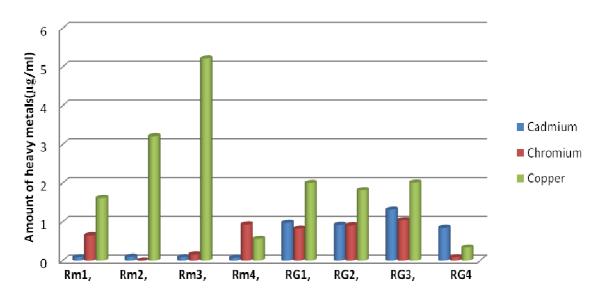


Figure 2. Heavy metals concentration in the muscles and gills region of Rameswaram fish.

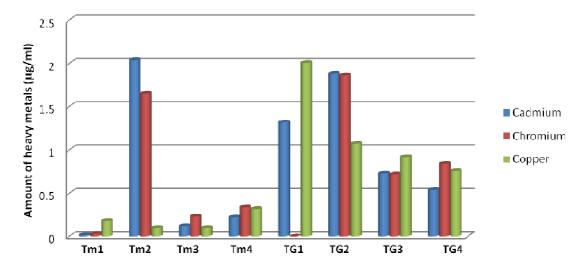


Figure 2. Heavy metals concentration in the muscles and gills region of Tuticorin fish.

CONCLUSION

In the present study, the microbes of human origin are affecting the marine environment and antibiotic resistant determinants are being transferred to other bacteria and human being. Immunosuppressive person and old age people are easily affect the salmonellasis diseases. In our study, provides new information on the concentration of heavy metals in the fish from Rameswaram and Tuticorin Harbor Area. Heavy metal concentrations in coastal waters as well as fish tissues have been found variable. The public health implication of the research seems to show no possibility of acute toxicity of heavy metals (Cu, Cd, Cr,) of edible fishes consumed. Nonetheless, continuous monitoring of biodata in these areas should continue while government should enforce existing pollution control laws, so that the metal concentrations do not get to critical levels but the fish adopted to that metal concentration then it will accumulate more metals in body that will threaten human health.

ACKNOWLEDGMENT

The authors wish to acknowledge the head of the institutions for the laboratory facilities provided.

CONFLICTS OF INTEREST

The authors declare that there are no conflicts of interest associated with this article.

REFERENCES

- Abdel-Monem, M.H. and Dowidar, A.A., 1990. Recoveries of *Salmonella fromsoil* in Eastern region of Saudi Arabia Kingdon, *J. Egypt. Public Health Assoc.*, 65: 61-75.
- Abila, R.O. and E.G. Jansen, 2003. From Local to Global markets: The fish exporting and fishing meal industries of Lake Victoria-Structure, Strategies and Socio-economic Impacts in Kenya, IUCN Eastern Africa programme. Socio-economics of Lake Victoria fisheries: Report No.2, The World Conservation Union, Nairobi, Acha, P.N; B. Szyfres, "Zoonoses and communicable diseases common to man and animals," Bacterioses and mycoses. 3rd ed. Scientific and Technical Publication, vol.1, No. 580, Pan American HealthOrganization, Regional Office of the WHO, Washington, USA, p. 384.

- Al-Bahry, S.N., Mahmoud, I.Y., Elshafie, A.E., Al-Harthy, A., Al-Ghafri, S., Al-Amri, I., and Alkindi, A.Y., 2009. Bacterial flora and antibiotic resistance from eggs of green turtles *Chelonia mydas:* an indication of polluted effluents. *Mar. Pollut. Bull.*, 58: 720–725.
- Al-Saleh, I. and Shinwari, N., 2002. Preliminary report on the levels of elements in four fish species from the Arabian Gulf of Saudi Arabia. *Chemosphere*, 48: 749-755.
- Altındag, A. and Yig, S., 2005. Assessment of heavy metal concentrations in the food web of lake Beysehir, Turkey. *Chemosphere*, 60: 552-556.
- Al-Yousuf, M.H., El-Shahawi, Al-Ghais, S.M., 2000a. Trace metals in liver, skin and muscle of *Lethrinus lentjan* fish species in relation to body length and sex. Sci. Total Environ., 256(2-3):87-94.
- Al-Yousuf, M.H., El-Shahawi, M.S. and Al-Ghais, S.M., 2000b. Trace metals in liver, skin and in Fish and Sediments from Lake Boeuf, Southeastern Louisiana. *Microchemical J.*, 62: 299-307.
- Aucoin, J; R. Blanchard, C. Billiot, C. Partridge, D. Schultz, and Mandhare, K., 1999. Trace Metals; K. Lemarchand, A. Brisabois, P. Lebaron, 2000. "Diversity of *Salmonella* strains isolated from the acquatic environment as determined by serotyping and amplification of ribosomal DNA spacer regions. *Appl. Environ. Microbiol.*, 66(4): 1544-1552.
- Bhaftopadhyay, P., 2000. Fish catching and handling. In: Robinson R.K. (ed.): Encyclopedia of Food Microbiology, Academic Press, London, Vol. 2, p. 1547.
- Black, R.E., Brown, K.H. Becker, S., 2003. Effects of diarrhoea associated with specific enteropathogens on the growth of children in rural Bangladesh. *Pediatrics*, 73: 799-805.
- Canli, M. and Atli, G.Z., 1987. The relationships between heavy metal (Cd, Cr, Cu, Fe, Pb, Zn) levels and the size of six Mediterranean fish species. *Environ. Pollut.*, 121: 129-136.
- Chao, W., Ding, R. and Chen, R. 1987. Survival of pathogenic bacteria in environmental

- microcosms, *Chinese J. Microbial Immun.*, 20: 339-348.
- Davies, R.H and Wray, C., 1996. Seasonal variations in the isolation of *Salmonella typhimurium, Salmonella enteritidis, Bacillus aureus* and *Clostridium perfringens* from environmental samples, *J. Vet. Med. Ser.m* 43: 119-127.
- Dural, M., Ziya Lugal Goksu, M. and Ozak, A.A., 2007. Investigation of heavy metal levels in economically important fish species captured from the Tuzla lagoon, *Food Chem.*, 102: 415–421.
- Dutch, H. and Altwegg, M., 1995. Evaluation of Five New Plating Media for Isolation of *Salmonella* species. *J. Clin. Microb.*, 33: 802-304.
- El-Shahawi, M.S., 1996. Spectroscopic and electrochemical studies of chromium III complexes with some naturally occurring ligands containing sulphur. *Spectrochim. Acta*, 52: 139-148.
- Esaki, H., Morioka, A. Ishihara, K., 2004. Antimicrobial susceptibility of Salmonella isolated from cattle, swine, and poultry (2001-2002): Report from the Japanese Veterinary Antimicrobial Resistance Monitoring Program. *Antimicrob. Chemother.*, 53: 266-270.
- Evans, D.W., Doo Do, D.K. and Hanson, P., 1993. Trace element concentration in fish livers: Implication of variations with fish size in pollution monitoring. *Mar. Pollut. Bull.*, 26: 329-354.
- Gomez, T.M., Motrjemi, Y. Miyagawa, S. Kaferstein, F.K. and Stohr, K., 1997. Foodborne *Salmonellosis*. *World Health Stat. O.*, 50: 81-89.
- Gooze, L.M.D., 1998. Bacterial Infections associated with HIV, HIV Insight Knowledge Base Chapter. Stanford Univ. Sch. of Medicine.
- Haltha, A. and Lakshmanaperumalsamy. M., 1997. Prevalence of Salmonella in fish and crustaceans from markets in Coimbatore, South India, *Food Microbiol.*, 14: 111-116.
- Hoar, W.S. and Randall, D.J., 1984. Fish Physiology. Vol. X. Gills. Part A. Anatomy, Gas Transfer, and Acid-Base Regulation. Academic Press, Orlando, FL.

- Huang, W.B., 2003. Heavy metal concentrations in the common benthic fishes caught from the coastal waters of Eastern Taiwan. *J. Food and Drug Analysis*, 11 (4): 324-330.
- Hunter, C. and A. Mac Donald, 1991. Seasonal market in Khartoum state. J. Bacteriology Res., 1: changes in the sanitary bacterial quality of water 085-088. Draining a small upland catchments in the Yorkshire.
- Hyland, R., Byrne, J., Selinger, B. and Graham, T., 2003. Spatial and temporal distribution of faecal indicator bacteria within the oldman river basin of South Alberta, Canada. *Water Qual. Res. J. Canada*, 38: 15-32.
- Kayambo, S. and Sven, E.J., 2006. Lake Victoria. Experience and lessons learnt. A case Study for Preliminary Risk Assessment Report, pp. 431-446.
- Kendrick, M.H., May, M.T., Plishka, M.J. and Robinson, K.D. 1992. Metals in Biological Systems. Ellis Horwood Ltd., England.
- Kraemer, L.D., Campbell, P.G.C. and Hare, L., 2006. Seasonal variations in hepatic Cd and Cu concentrations and in the sub-cellular distribution of these metals in juvenile yellow perch (*Perca flavescens*). *Environ. Pollut.*, 142: 313-325.
- Krishnamurthi, Asha Jyothi and Nair, V.R., 1999. Concentration of metals in fishes from thane and Bassein creeks from Bombay, India. *Indian J. Mar. Sci.*, 28: 39-44.
- Kümmerer, K., 2009. Antibiotics in the aquatic environment. A review. Part II. *Chemosphere*, 75: 435-441.
- Levy, S.B., 1992. The Antibiotics Paradox: How Miracle Drugs are Destroying the Miracle. Plenum, New York.
- Levy, S.B., 1994. Balancing the drug resistance equation. *Trends Microbiol.*, 2, 41-342.
- Lipp, E.K. and Rose, J.B., 1997. The role of sea food in food borne diseases in the United States of America. *Rev. Sci. Tech. OIE*, 16: 620-640.
- Liston, J., 1980. Microbiology in fishery science. In Connell, JJ. (ed). Advances in

- Fish Science and Technology; Jubilee Conference of Torry Research Oxford: Fishing News Books Ltd., Farnham, UK.
- Michael, G; Simoneti, R. de Costa, M. and Cardoso, M., 2003. Comparison of Different Selective Enrichment Steps to Isolate Salmonella Sp. from Feces of Finishing Swine. *Brazil J. Microbiol.*, 34: 138-142.
- Onyango, D., Wandili, S. Kakai, R. and Waindi, E.N., 2008. Isolation of Salmonella and Shigella from fish harvested along winam Gulf of lake Victoria, Kenya, *J. infect. Dis. In Developing Countries*, 2: 106-111.
- Pang, J. Bhutta, Z.A., Finlay, B.B., Altwegg, M., 1995. Typhoid, fever and other Salmonellosis: a continuity challenge. *Trends Microbial.*, 3: 253-255.
- Prabakaramurthy, P.V.S. and Satyanarayana, D., 1999. A comparative study of atomic absorption spectrometry and anodic stripping voltametry for the determination of trace metals Zn, Cd, Pb and Cu in the coastal waters of Visakhapatanam, east coast of India. *Indian J. Mar. Sci.*, 28, 365-369.
- Romeo, M., Siau, Y., Sidoumou, Z. and Gnassia-Barelli, M., 1999. Heavy metal distribution in different fish species from the Mauritania coast, *The Sci.Total Environ.*, 232: 169-175.

- Tauxe, R.V., 1997. Emerging food borne diseases: an evolving public health challenge. The fish exporting and fishmeal industries of Lake Victoria structure, strategies and socio-economic impacts in Kenya "Annual Report, Emerge. *Infect. Dis.*, 3: 425-434.
- Usero, J., Izquierdo, C., Morillo, J. and Gracia, I., 2003. Heavy metals in fish (*Solea vulgaris*, *Anguilla anguilla* and *Liza aurata*) from salt marshes on the southern Atlantic coast of Spain. *Environ. Int.*, 29: 949-956.
- Tian, Y.Q., Gong, P.D., Radke, J. and Scarborough, J., 2002. DNA probe by colony hybridization using non-Spatial and temporal modeling of microbial isotopic and isotopic labeling. APMIS, 100: 623-628.
- Yılmaz, A.B. 2003. Levels of heavy metals (Fe, Cu, Ni, Cr, Pb and Zn) in tissue of *Mugil cephalus* and *Trachurus mediterraneus* from Iskenderun Bay, Turkey. *Environ. Res.*, 92: 277-281.
- Zlotkin, A., Eldar, A., Ghifino, C. and Bercovier, H., 1989. Identification of *Lactococcus garvieae* by PCR. *J. Clin. Microbiol.*, 36: 983-985.