Characterization of SNPs in Oxidised Low Density Lipoprotein Receptor 1 (*OLR1*) gene in Mehsana and Banni breeds of buffalo.

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Abstract

Single Nucleotide polymorphisms (SNP) are the most common type of DNA sequence polymorphisms and of mutation in vertebrates. Keeping in view the involvement of Oxidised Low Density Lipioprotein Receptor 1 (*OLR1*) gene in lipid metabolism and the higher milk fat percentages in *Bubalus bubalis*, the present study was carried out to explore the SNPs in *OLR 1* intron I in Mehsani and Banni breeds of buffalo by gene cloning and sequencing. Five SNPs at nucleotide positions viz. 327, 541, 957 and 1128 were detected and all the SNPs followed breed lineage. The SNP at positions 327 and 541 were found in Mehsana breed alone while as rest of the explored SNPs restricted to Banni breed of *Bubalus bubalis*. The frequency of each SNP was found to be 0.025.

Keywords: Single Nucleotide Polymorphisms, buffalo, Oxidised Low density Lipoprotein Receptor 1.

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Introduction

Oxidised Low Density lipoprotein Receptor 1 (OLR1) is the major protein that binds, internalizes, and degrades oxidized low-density lipoprotein. The role of OLR1 in lipid metabolism and the results of previous whole genome scan studies prompted the investigation of OLR1 as a candidate gene affecting milk composition traits [1]. In another study conducted in Dutch Holstein-Friesian cattle population in OLR 1 gene to estimate genotype effects on milk production traits revealed that $OLR_{g.8232C>A}$ had a significant effect (P < 0.05) on milk-fat percentage [2].

The oxidized form of the low-density lipoprotein (oxLDL) is involved in endothelial cell injury, dysfunction, and activation, all of which are implicated in the development of atherosclerosis [3]. It has been shown that oxLDL and its lipid constituents have numerous damaging effects on secretory activities of the endothelium, including induction of apoptosis [4]. The major protein that binds, internalizes, and degrades oxLDL, oxidized LDL receptor 1 (*OLR1*), was initially identified in bovine aortic endothelial cells [5]. In addition to binding oxLDL, *OLR1* removes aged and apoptotic cells from blood circulation (Oka et al., 1998). The bovine *OLR1* gene encodes 270 AA and has 72% identity to the human protein [6]. It has

been previously demonstrated that the human *OLR1* contains 6 exons, of which the first 3 corresponded to the Nterminal cytoplasmic, transmembrane, and connecting neck domains, and the last 3 encoded the lectin domain [7]. The genomic sequence of bovine *OLR1*, recently released by Baylor College of Medicine, contains 5 exons {GenBank accession no. NW_215807}.

Based on the aforementioned studies indicating the role of *OLR1* in lipid metabolism and its correlation with milk fat percentage, the study of SNP exploration in this gene in *Bubalus bubalis* was initiated considering the higher milk fat percentages in buffaloes.

Materials and Methods

Sampling and DNA extraction

Blood samples, ten each from two breeds of buffalo, Mehsani (from Dudhsagar Dairy, Mehsana, Gujarat) and Banni (from field animals of Kachchh district, Gujarat) were collected at random from non related animals. Genomic DNA was extracted by Phenol:Chloroform method. The concentration and the purity of the DNA obtained were assessed by spectrophotometery and electrophoresis in 0.8% agarose gel respectively.

PCR amplification

The genomic DNA from each individual was amplified by PCR. A pair of primers designed using the reference sequence (Accession no. NC_007303.3) of OLR1 of Bos taurus (Forward:AGAAAACTACAATGCCTGGGT & Reverse: CACCACAAGGCAGAGAGT) was used to amplify a gene segment of 1163bp. Approximately 30ng of genomic DNA was amplified in 25µl PCR reaction consisting of 2X Master mix (MBI Fermentas). Thermal cycling was performed on Veriti (Applied Biosystems) with an initial denaturing step at 95°C for 10 mins followed by 40 cycles of denaturation at 95^oC at 60 seconds, primer annealing at 56° C for 75 seconds, and primer extension at 72°C for 45 seconds. The PCR products were confirmed on a 1.5% agarose in 0.5% TBE buffer, visualized under ultraviolet light and eluted (Qiaquick gel extraction kit) for purification of the PCR product.

Ligation and transformation

The purified PCR products were ligated in pTZ57R/T vector using InsT/AcloneTM PCR product cloning kit (Cat no. K1213, MBI Fermentas). Ligation of the PCR fragment was confirmed by PCR using M13 specific primers targeting M13 sequences contained in pTZ57R vector. Ligated recombinant vector was transformed in competent *E. coli* (DH5- α) cells. Recombinant clones were selected by Blue-white screening. Recombinant plasmids were isolated and used for cycle sequencing, that were later processed for sequencing using the universal M13 primers.

Sequencing and SNP detection

Extension products obtained from cycle sequencing reaction were purified by vacuum manifold using AcroPrepTM 96 filter plates, (Cat. no. 5033, Omega 3K, Pall Corporation, USA). Cycle sequencing products were run on ABI-PRISM automated DNA sequencer and raw data was collected by 310 Data Collection Software (Version 3.1.0). Sequences obtained by sequence analyzer were curated and vector sequences were removed using Vecscreen (NCBI) programme. These filtered sequences were then subjected to local alignment with Genebank database sequences using BLASTn protocol available at NCBI.

A consensus sequence of *OLR1* Intron I of buffalo was generated and used as a reference sequence. Forty sequences were compared by multiple sequence alignment by using a multiple sequence alignment tool, Bioedit (v 7.0.7.1). SNP detection was done by multiple sequence alignment of consensus sequences of *Bubalus bubalis* in Bioedit (v 7.0.7.1) and MEGA4. SNP in a sequence was judged as a mismatch between samples.

Results and Discussion

Five SNPs (Fig. 1 and Fig. 2) were detected within the 1163 bp fragment of *OLR1* intron I gene in four breeds of *Bubalus bubalis* (Table 1). The frequency and breed composition of each of the six SNPs detected in the present study are summarized in the Table 2. The presence of all the SNP was affected by breed. The presence of SNP at nucleotide positions 327 and 541 were associated with the Mehsani breed where as SNPs at positions 813 and 957 found their association with Banni breed. SNP at all positions viz. 327, 541, 813, 846, 957 and 1128 were present in 2.5% of the population studied.

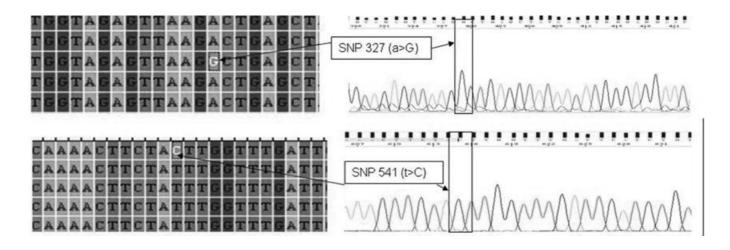


Figure A: Multiple Sequence Alignment and Sequencing Analysis files showing SNP at positions 327 and 541 in cloned gene fragment of OLR1 intron I in Bubalus bubalis.

Characterization of SNPs in Oxidised Low Density Lipoprotein Receptor

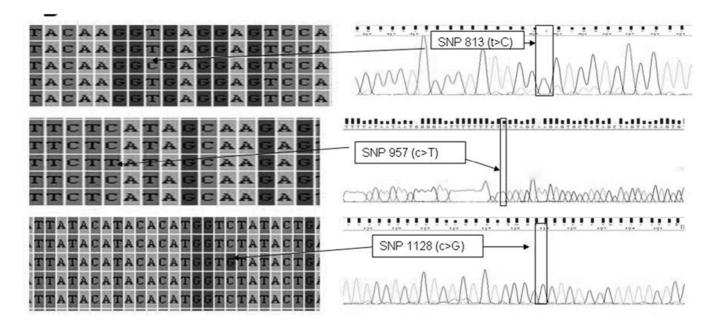


Figure B: Multiple Sequence Alignment and Sequencing Analysis files showing SNP at positions 813, 957 and 1128 in cloned gene fragment of OLR1 intron I in Bubalus bubalis.

Table 1: SNPs in OLR1 Intron I of Mehsani and Banni buffalo (Reference Sequence-GQ478035)

Sample ID	Nucleotide position	Base change	Frequency	Nucleotide Sequence Accession number	SNP ID
Mb 1	327	a>G	0.025	GQ415424	155458697
Mb 2	541	t>C	0.025	GQ415420	155458699
Bn 2	813	t>C	0.025	GQ415444	155458700
Bn 3	957	c>T	0.025	GQ415440	155458703
Bn 4	1128	c>G	0.025	GQ415447	155458704

Mb: Mehsani, Bn : Banni

Table 2. Effect of breed composition¹ on Single Nucleo-tide Polymorphism occurrence

SNP position	Frequency ^b	Breed ^a	
		Mehsani	Banni
327	0.025	1	0
541	0.025	1	0
813	0.025	0	1
957	0.025	0	1
1128	0.025	0	1

^a Number of the animals with the detected SNP by breed; Mehsani and Banni.

^bPercentage of buffalo with that SNP as observed in 20 animals.

India possesses eleven recognized breeds of buffalo. Present study was conducted in two breeds found in Gujarat state. Mehsani and banni are the medium sized breeds, having a high milk fat percentage (>8 %). The most abundant type of variation in human and cattle genomes is the single nucleotide polymorphism or SNP, where a single base pair has been changed. Millions of SNPs have been found in humans, and there are over 600,000 in cattle with more being discovered every day [8]. In our earlier study we screened the same region of OLR1 in Jaffarabadi and Surti breeds of buffalo where in we explored three SNPs at nucleotide position 423 and 866 in surti buffalo and at 843 in Jaffarabadi buffalo. Various studies in bovine species have been conducted around the globe to derive an association of OLR1 gene polymorphisms with the milk fat percentage. In a study conducted in Italian brown swiss involving the genotyping for a SNP at position 8232 in OLR1 (NW_215807:g.8232C>A) revealed a frequency of 0.95 for the g.8232C allele [9]. In another study conducted in Dutch Holstein-Friesian cattle population in OLR 1 gene revealed that $OLR_{g,8232C>A}$ had a significant effect (P < 0.05) on milk-fat percentage [2]. A study conducted in Holstein dairy cattle concluded that allele C of SNP 8232 in 3' UTR in OLR1 gene had significant effects on fat yield and fat percentage [1]. Breed specific SNPs are not uncommon. In a study conducted on heat shock protein 70 in pure Brahman, pure Angus and their intercross breeds of cattle, breed specific SNPs were explored that tended to be associated with the particular breeds only [10]. The SNPs detected in the present study also followed breed lineage. All the SNPs showed breed specific occurrence which is fully in agreement with our previous study where in we explored three SNPs in Surti and Jaffarabadi breeds of Buffalo and all the three SNPs were found to be breed specific [11]. However, as these SNPs are present in intronic region, their functional significance is required to be verified. The OLR1 sequences and SNPs explored in Mehsana and banni breeds of buffalo are accessible in National Centre of Biotechnology Information (NCBI) and their Genbank accession numbers and SNP ID's have been enlisted in the Table1.

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