

Characterization of food poisoning bacteria: Advances in diagnostic approaches.

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Introduction

Food poisoning, caused by bacterial contamination of food, is a significant public health concern worldwide. Rapid and accurate identification and characterization of food poisoning bacteria are essential for effective outbreak investigation, surveillance, and targeted control measures. Over the years, significant advancements have been made in diagnostic approaches, enabling the precise detection, identification, and characterization of food poisoning bacteria. In this article, we will explore the recent advances in diagnostic techniques that aid in the characterization of food poisoning bacteria. Culture-based methods have long been the gold standard for isolating and identifying food poisoning bacteria. These methods involve the enrichment and selective isolation of target organisms, followed by phenotypic characterization based on colony morphology, biochemical tests, and antimicrobial susceptibility patterns [1, 2].

Although culture-based methods are still widely used, they have limitations in terms of time required for analysis, low sensitivity, and the inability to identify non-culturable or fastidious bacteria. PCR-based techniques have revolutionized the field of microbial diagnostics. PCR enables the rapid and specific amplification of target DNA sequences, allowing the detection and identification of food poisoning bacteria with high sensitivity. Real-time PCR, also known as quantitative PCR (qPCR), provides additional advantages by allowing the detection and quantification of bacterial DNA in real-time, providing rapid results. PCR-based methods can target specific genes or regions, such as 16S rRNA or virulence genes, enabling the identification and characterization of food poisoning bacteria at the molecular level. NGS technologies have transformed the field of microbial genomics and have significant applications in characterizing food poisoning bacteria [3, 4].

Whole-Genome Sequencing (WGS), a powerful NGS approach, allows the sequencing and analysis of the entire bacterial genome. WGS provides comprehensive information on bacterial species, genetic relatedness, antimicrobial resistance genes, and virulence factors. The high-resolution data obtained from WGS enables precise identification and subtyping of food poisoning bacteria, facilitating outbreak investigations and source tracking. Additionally, metagenomic sequencing allows the detection and characterization of

the entire microbial community present in a food sample, providing insights into the overall microbial ecology and potential food safety risks. Mass spectrometry (MS)-based approaches, such as matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-TOF MS), have emerged as rapid and reliable tools for bacterial identification [5, 6].

Microfluidic devices and lab-on-a-chip systems offer miniaturized platforms for food poisoning bacterial analysis. These devices integrate various analytical functions, such as sample preparation, amplification, and detection, into a single chip. Microfluidics enables precise control of fluids and reagents at the microscale, reducing the required sample and reagent volumes, and shortening analysis time. Lab-on-a-chip systems provide portable, rapid, and user-friendly diagnostic tools for the on-site detection and characterization of food poisoning bacteria. Advances in diagnostic approaches have significantly enhanced the characterization of food poisoning bacteria, enabling rapid and accurate identification, subtyping, and characterization of pathogens. Culture-based methods, PCR and real-time PCR, NGS, mass spectrometry, immunoassays, biosensors, microfluidics, and lab-on-a-chip systems offer diverse tools for food safety professionals and researchers [7, 8].

These technologies facilitate early detection of food poisoning bacteria, source tracking of outbreaks, monitoring of antimicrobial resistance, and identification of emerging pathogens. Continued research and development in diagnostic approaches will further improve our ability to detect and characterize food poisoning bacteria, enhancing food safety and public health measures. Effective characterization of food poisoning bacteria requires global collaboration and standardization of diagnostic approaches. International organizations and regulatory bodies play a vital role in coordinating efforts, establishing quality control measures, and developing reference databases. Collaboration among researchers, public health agencies, and the food industry helps ensure the harmonization and validation of diagnostic methods, facilitating data sharing and comparability. Standardization ensures consistency in results, enables robust surveillance systems, and supports effective control measures against food poisoning bacteria [9, 10].

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References

1. Sender R, Fuchs S, Milo R. Revised estimates for the number of human and bacteria cells in the body. *PLoS Biol.* 2016;14(8):1002533.
2. Chen M, Yu Q, Sun H. Novel strategies for the prevention and treatment of biofilm related infections. *Int J Mol Sci.* 2013;14(9):18488-501.
3. Paharik AE, Horswill AR. The staphylococcal biofilm: Adhesins, regulation, and host response. *Virulence mechanisms of bacterial pathogens.* 2016:529-66.
4. Zheng Y, He L, Asiamah TK, et al. Colonization of medical devices by staphylococci. *Environ. Microbiol.* 2018;20(9):3141-53.
5. Chang CY. Surface sensing for biofilm formation in *Pseudomonas aeruginosa*. *Front Microbiol.* 2018;8:2671
6. Guihen E, Hogan AM, Glennon JD. High-speed microchip electrophoresis method for the separation of (R, S)-naproxen. *Chirality.* 2009 Feb;21(2):292-8.
7. Dolník V, Liu S, Jovanovich S. Capillary electrophoresis on microchip. *Electroph Internat J.* 2000;21(1):41-54.
8. Pfeiffer AJ, Mukherjee T, Huan S. Design and optimization of compact microscale electrophoretic separation systems. *Industri Engin Chem Res.* 2004;43(14):3539-53.
9. Hradski J, Chorváthová MD, Bodor R, et al. Quantitative aspects of microchip isotachopheresis for high precision determination of main components in pharmaceuticals. *Analyti Bioanaly Chem.* 2016;408:8669-79.
10. Al-Othman ZA, Ali I. Nano capillary electrophoresis in microchips: A need of the present century. *J Liqu Chromatogra Related Techno.* 2011;34(14):1295-325.