

Biological potential and GC-MS analysis of phytochemicals of *Farsetia hamiltonii* (Royle)

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Abstract

Objective: The study was designed to evaluate the biological and phytochemical potential of *Farsetia hamiltonii* (Royle), a desert medicinal plant belongs to family Brassicaceae and is traditionally used by residents of Cholistan to treat diabetic, different infectious diseases and gastrointestinal problems.

Method: Different biological/enzyme inhibition activities of crude extract of *Farsetia hamiltonii* were performed first time in this study. The dichloromethane and methanol extracts of aerial part of *Farsetia hamiltonii* were studied for anti-alpha-glucosidase, anti-chymotrypsin, anti-urease, anti-butrylcholinesterase and anti-bacterial activities. The plant was explored first time regarding phytochemical constituents by using the GC-MS technique.

Results: The results of crude extracts of *Farsetia hamiltonii* were significant and provide justification for the traditional therapeutic potential in treating the diabetes, infectious diseases and gastrointestinal problems. Nine (9) compounds were identified by GC-MS technique.

Keywords: Anti-bacterial activity, Anti-chymotrypsin, Anti-alpha-glucosidase, *Farsetia hamiltonii* royle.

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Introduction

Cholistan desert is situated in south west of Punjab (Pakistan) and its area is about 26000 Km² having highly saline soil [1]. It is unique wild land having endemic flora containing 28 families and 138 species out of which about 64 species have been identified as medicinal plants [2]. The plants are traditionally used by inhabitants to treat different types of ailments as well as for animals by forming decoction, pellets and powder of herbs [3].

Farsetia hamiltonii (Royle) is a desert plant and is endemic to the desert area of Pakistan and India. It is an annual shrubby, often woody at base, 10-40 cm long, vary variable in size, densely appressed white hairs, braches erect or sub spreading, leaves oblong linear, thick woody roots, fruit a little longer, narrowed at either end, are distinctive. Flowers are white or pink in color, petals slightly longer than sepals and flowering season is March to September [4]. The medicinal use of plant is to treat pain, inflammation and swelling in joints, diabetes, gastrointestinal and infectious diseases. This plant is used with "Ghee" as a cooling medicine and tonic. Its boiled juice is applied to treat camel wound in Cholistan [3]. The local name is "Fareed Booti" [5]. This plant is selected for study keeping in view the medicinal importance of genus *Farsetia*. The ethyl alcohol extract of aerial parts of *F. aegyptia* was in vitro investigated for cytotoxicity and exhibited cytotoxicity against A549 and HepG2 carcinomas. The phenolic rich fraction was further fractionized and isolated a new flavonoid kampferol-8-C-O-β-diglucoopyranoside which showed the

high cytotoxicity against Hela and MCF-7 cell lines [6]. The alcoholic extract of *F. aegyptia* is evaluated for anti-bacterial and anti-fungal activities which show maximum inhibition against *Klebsiella pneumonia* and no activity against *Candida albicans* [7]. According to literature survey, no phytochemical and biological activities on *F. hamiltonii* have been reported and this species is found unexplored.

Methods

Enzymes, chemicals and equipments

Alpha glucosidase (EC 3.2.1.20), chymotrypsin (EC 3.4.21.1), urease (EC 3.5.1.5), Butyryl cholinesterase (EC 3.1.1.8), Acarbose, Chymostatin, Thiourea, Eserine, chloroform, acetone, diethyl ether, dichloromethane and methanol were purchased from Sigma-Aldrich and Merck companies. Ciprofloxacin, KH₂PO₄, DTNB [5,5-dithiobis (2-nitrobenzoic acid)], phosphate buffer, N-succinyl phenylalanine-P-nitroanilide and 4-nitrophenyl acetate were of analytical grade.

The gram-positive bacteria (*Bacillus subtilis*, *Staphylococcus aureus*) and gram-negative (*Escherichia coli*, *Pseudomonas aeruginosa*, *Salmonella typhi*) were used in this study.

The instruments synergy HT BioTek 96 microplate reader, Rotary evaporator (Buchi), Vacuum pump, Spectrophotometer (Shimadzo, Japan), EZ-Fit Enzyme Kinetics software (Perrella Scientific Inc. Amherst, USA) and Gas chromatography-mass spectrometry (GC Agilent system, USA) were used.

Plant collection and extraction

The fresh plants of *F. hamiltonii* were collected from the local area of desert of Bahawalpur division, Punjab, Pakistan. The plant specimen was identified by research officer/taxonomist, Mr. Hafiz Muhammad Waris from Cholistan Institute of Desert Studies (CIDS), The Islamia University of Bahawalpur, Pakistan. The plant specimen is deposited in the herbarium of CIDS for future reference and allocated voucher number 3470/CIDS/IUB.

The aerial parts of plants were dried under shade for 15 days and ground in fine powder by the mill. The powdered plant material of aerial parts of *F. hamiltonii* (900 g) were macerated in dichloromethane (4.5 liters) for 24 hours, the process was repeated for three days and then same in methanol for three days. The dichloromethane and methanol extracts are concentrated in solid residue by using the rotary evaporator under reduced pressure. The extracts were abbreviated as FHAD for dichloromethane extract of aerial parts of *F. hamiltonii* and FHAM for methanol extract of aerial parts of *F. hamiltonii*. The yields of extraction are 1.8% and 2.8% respectively.

Biological and enzymatic activities

Alpha glucosidase inhibition activity: This assay was performed according to the slightly modified method [8]. The 70 µL phosphate buffer (50 mM, pH 6.8), 10 µ of plant extract solution (0.5 mg/ml), then, added 10 µl alpha glucosidase enzyme (0.057 units). The contents were mixed and pre-read. The reaction was initiated by the addition of 10 µl of 0.5 mM per well substrate. The absorbance was measured at 400 nm. Acarbose was used as standard. The results are given in Table 1.

Chymotrypsin inhibition activity: A standard procedure was followed with slight modification [9]. 60 µl Tris-HCl buffer (50 mM, pH 7.6), 10 µl plant extract solution (0.5 mg/ml) and 15 µl purified α-chymotrypsin enzyme (0.9 units) were mixed in 96-microwell plate. The contents were pre-read. The reaction was initiated by the addition of 15 µl N-succinyl phenyl-alanine-p-nitroanilide (1.3 mM). The absorbance was measured at 410 nm. Chymostatin (0.5 mM) was used as standard (Table 1).

Urease inhibition activity: It is the modified form of Berthelot activity [10]. The phosphate buffer (10 µl, pH 7.0), 10 µl of plant extract solution (0.5 mg/ml) and 25 µl

of urease enzyme solution (0.1347 units) were used. Then, 40 µl of urea stock solution (20 mM) was added and 115 µl phenol hypochlorite reagents (prepared by mixing 45 µl phenol reagents with 70 µl of alkali reagent) were added. The absorbance was measured at 625 nm and Kojic acid was used as standard (Table 1).

Butyrylcholinesterase inhibition activity: Butyryl cholinesterase (BChE) inhibition activity was performed according to the method [11]. In this assay, 60 µl KH₂PO₄ buffer (100 mM, pH 7.7) and 10 µl plant extract solution (0.5 mg/ml) were mixed, followed by the addition of 10 µl enzyme butyryl cholinesterase. The contents were pre-read. The reaction was initiated by the addition of 10 µl of DTNB. The absorbance was measured at 405 nm and Eserine was used as standard (Table 1).

Anti-bacterial activity: The method [12] was used, two gram-positive bacteria (*Bacillus subtilis*, *Staphylococcus aureus*) and three gram-negative (*Escherichia coli*, *Pseudomonas aeruginosa*, *Salmonella typhi*) were included. The absorbance was measured at 540 nm, before and after incubation and the difference was noted as an index of bacterial growth. Ciprofloxacin (0.5 mM) was used as a standard. The percent inhibition was calculated using the formula:

$$\text{Inhibition (\%)} = 100 \times (X - Y)/X$$

Where X is absorbance in control with bacterial culture and Y is absorbance in test sample (plant extract solution). The results are given in Table 2.

Gas chromatography-mass spectrometry (GC-MS): GC-MS analysis is used for identification of chemical constituents present in the sample. It has great importance in drug detection and identification of unknown phytochemical constituents present in the medicinal plants. So, GC-MS analysis included in the study to investigate the unknown constituents of the selected medicinal plants *F. hamiltonii* [13].

GC-MS analysis was performed on GC Agilent system (B 7890) with mass spectrometer detector (MSD-5977A) employing the following condition: Column HP-5MS, size 30 m × 0.25 mm, 0.25 µ, composed of 100% dimethyl poly siloxane. The source temperature for ionization was set at 250°C. The 2 µl of dichloromethane and methanol extracts of aerial parts of *F. hamiltonii* were used in GC-MS analysis [14].

Table 1. Enzyme inhibition assay of dichloromethane and methanol extracts of *Farsetia hamiltonii* (Royle).

Enzyme Assay	DCM* Extract		Methanol Extract		Standard	
	Inhibition (%)	IC50 (µmoles)	Inhibition (%)	IC50 (µmoles)	Inhibition (%)	IC50 (µmoles)
α-glucosidase inhibition assay	35.51 ± .83	Inactive	97.54 ± .39	25.32 ± .12	92.23 ± 0.14	Acarbose 38.25 ± 0.12 Acarbose
Chymotrypsin inhibition assay	69.64 ± .01	333.12 ± .01	76.93 ± .01	238.57 0.01	93.50 ± 0.91	Chymostatin 8.24 ± 0.11 Chymostatin
Urease inhibition assay	56.77 ± .27	381.50 ± .52	63.31 ± .46	351.60 ± 0.54	82.11 ± 0.14	Thiourea 22.04 ± 0.12 Thiourea
Butyrylcholin-esterase inhibition assay	21.96 ± .22	Inactive	21.96 ± .22	Inactive	91.29 ± 1.17	Eserine 0.04 ± 0.001 Eserine

Note: All values are mean ± SEM of triplicate. *DCM: Dichloromethane

The GC-MS results revealed the presence of nine (09) compounds in two different extracts of aerial part of *F. hamiltonii*. The name, molecular formula, molecular weight and structure of the compounds are given in Tables 3 and 4 with their pharmacological effects. The five (5) compounds are identified in dichloromethane extract whereas four (04) compounds are identified in methanol extract of aerial part of *F. hamiltonii*.

Statistical analysis

The percentage enzyme inhibition was calculated by the following formula:

Inhibition (%) = $100 \times (\text{Absorbance of test sample} / \text{Absorbance of control}) \times 100$.

IC₅₀ values (concentration at which there is 50% in enzyme catalyzed reaction) compounds were calculated using EZ-Fit Enzyme Kinetics Software (Perrella Scientific Inc. Amherst, USA). For the determination of IC₅₀ values, test solutions were assayed at various dilutions i.e., 0.5, 0.25, 0.125, 0.0625

mg/ml. All the results (Tables 1 and 2) are mean \pm SEM (standard error of mean) of triplicate values (n=3).

Results

Farsetia hamiltonii (Royle) is endemic desert plant used by hakims and residents of Cholistan desert by forming decoction or in powder form to treat diabetes, infections, pain, inflammation and gastrointestinal diseases [3]. To confirm the traditional anti-diabetic use of *F. hamiltonii*, alpha glucosidase inhibition activity was evaluated. The methanol extract of aerial parts of *F. hamiltonii* possess (%) inhibition 97.54 ± 2.39 with IC₅₀ = $25.32 \pm 0.12 \mu\text{M}$, was more potent in crude form than standard Acarbose having 92.23 ± 0.14 with IC₅₀ = $38.25 \pm 0.12 \mu\text{M}$, which indicates that this FHAM crude extract may have some anti-diabetic compounds. The alpha glucosidase enzyme catalysis the hydrolysis of disaccharides into glucose, inhibition of this enzyme can suppress the post prandial hyperglycemia and this inhibition will be useful invention for management of diabetes type II [15]. The presence of this activity only in

Table 2. Results of anti-bacterial activity with MIC₅₀ of crude extracts of aerial parts of *F. hamiltonii*.

Extract Code	Type	<i>Bacillus subtilis</i> (G+ve)	<i>Staphylococcus aureus</i> (G+ve)	<i>Pseudomonas aeruginosa</i> (G-ve)	<i>Salmonella typhi</i> (G-ve)	<i>Escherichia coli</i> (G-ve)
DCM Extract (FHAD)	Inhibition* (%)	65.17 \pm 4.28	60.17 \pm 4.28	74.02 \pm 1.39	74.10 \pm 1.33	68.64 \pm 1.36
	MIC ₅₀	13.93 \pm 0.84	13.21 \pm 1.58	9.69 \pm 1.90	9.44 \pm 1.09	12.69 \pm 0.37
Methanol Extract (FHAM)	Inhibition (%)	54.70 \pm 0.75	56.33 \pm 1.50	54.63 \pm 0.05	58.13 \pm 1.46	51.40 \pm 1.15
	MIC ₅₀	13.40 \pm 0.34	12.70 \pm 0.33	13.48 \pm 0.12	12.37 \pm 0.12	17.87 \pm 0.41
Ciprofloxacin (standard)	Inhibition (%)	91.23 \pm 1.07	91.23 \pm 1.07	90.88 \pm 0.16	92.65 \pm 1.10	91.45 \pm 2.19
	MIC ₅₀	7.52 \pm 0.67	7.03 \pm 0.53	7.58 \pm 0.19	7.23 \pm 0.71	8.21 \pm 0.11

*Concentration of extract was 100 $\mu\text{g}/\text{well}$

Table 3. Phytochemical constituents identified by GC-MS in dichloromethane extract of aerial part of *Farsetia hamiltonii* Royle.

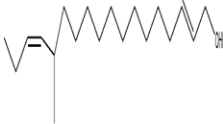
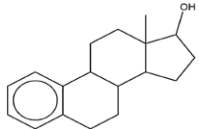


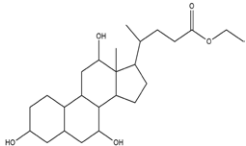

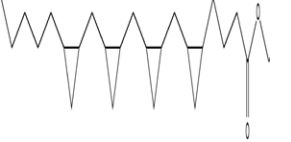
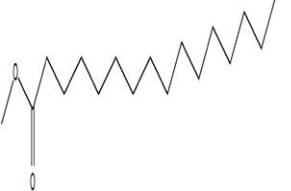
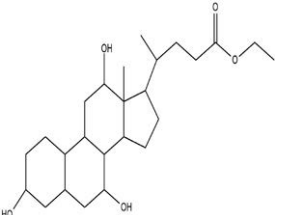
S. No.	Phytochemical compound	RT (min)	Mol. formula	Mol. weight	Structure	Pharmacological effect
1	12-methyl-E,E-2,13-octadecadien-1-ol	12.55	C ₁₉ H ₃₆ O	280		Anti-microbial and anti-fungal agent, Already identified in plant <i>Gossypium saeed</i> and <i>T. hemprichii</i> [17].
2	Estra-1,3,5(10)-trien-17 β -ol	13.42	C ₁₈ H ₂₄ O	256		Anti-proliferative effect for treatment of primary breast cancer [18].
3	Oleic acid	14.36	C ₁₈ H ₃₄ O ₂	282		Deficiency cause the acne and skin problems, anti-oxidant, reduce blood pressure, prevents ulcerative colitis [19], Already identified in plant <i>Euphorbia hirta</i> Linn [24].
4	13-Heptadecyn-1-ol	14.53	C ₁₇ H ₃₂ O	252		Anti-inflammatory and anti-fungal activity, Already identified in plant <i>Sinapis arvensis</i> [20] and <i>Euphorbia hirta</i> Linn [24].
5	Ethyl iso-allocholate	17.13	C ₂₆ H ₄₄ O ₅	436		Anti-bacterial agent, Inhibitor of dihydropteroate synthetase enzyme, Already identified in from medicinal rice Karungkavuni [21].

Table 4. Phytochemical constituents identified by GC-MS in methanol extract of aerial part of *Farsetia hamiltonii* Royle.

S. No	Phytochemical compound	RT (min)	Mol. formula	Mol wt.	Structure	Pharmacological effect
1	Octadecan, 6-methyl	6.17	C ₁₉ H ₄₀	268		Not found from literature.
2	Cyclo propane botanic acid, 2-[[2-[[2-[(2-pentyl cyclo propyl)methyl] cyclo propyl] methyl] cyclo propyl] methyl]-methyl ester	13.17	C ₂₅ H ₄₂ O ₂	374		Already identified in <i>Wrightia tinctoria</i> seed [22].
3	Hexadecanoic acid, methyl ester	14.29	C ₁₇ H ₃₄ O ₂	270		Anti-oxidant, Hypocholesterolemic effect, Nematicide, Pesticide, anti-androgenic, Hemolytic, 5-alpha reductase inhibitor [23], Already identified in <i>Fluggea leucopyrus</i> and <i>Gossypium</i> seeds [17].
4	Ethyl iso-allocholate	14.37	C ₂₆ H ₄₄ O ₅	436		Anti-microbial activity, Already identified in from medicinal rice Karungkavuni [21].

methanol extract indicates that some polar biomolecules are responsible for anti-diabetic effect. The results are shown in Table 1.

Traditional practitioners use the plant *F. hamiltonii* to treat gastrointestinal problems and other illnesses. The enzyme inhibition studies for different enzymes (chymotrypsin, urease and butyryl cholinesterase) for both dichloromethane and methanol extracts of *F. hamiltonii* were performed in triplicate and presented with SEM in Table 1. The dichloromethane extract possess significant inhibition of chymotrypsin (69.64 ± 0.01% with IC50 value=333.12 ± 0.01 µM) and urease (56.77 ± 0.27% with IC50 value=381.50 ± 0.52 µM) activities, whereas methanol extracts exhibited chymotrypsin inhibition (76.93 ± 0.01% with IC50 value=238.57 ± 0.01 µM) and urease inhibition (63.31 ± 0.46% with IC50 value=351.60 ± 0.54 µM) activities.

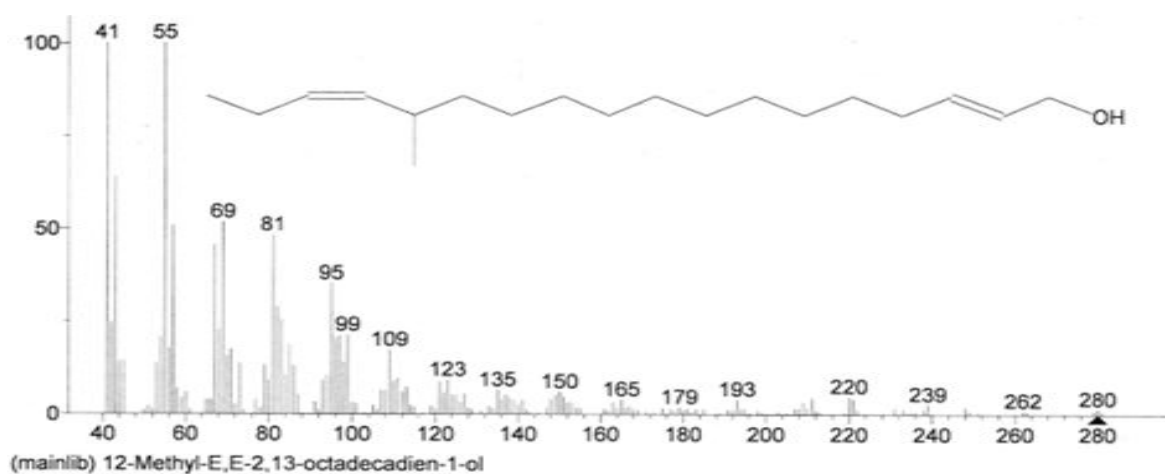
Research is going on enzyme urease regarding its correlation with several diseases including in animals and humans like urinary tract diseases and gastro duodenal diseases. An important role played in the pathogenesis of a disease by the bacterial urease including bacterial species like *Staphylococcus saprophiticus*, *Ureaplasma urealiticum*, *Yesinia enterocolitica* and *Proteus mirabilis* [16]. Both

extracts (FHAD and FHAM) of the plant not exhibited the anti-butyryl cholinesterase activity.

The anti-bacterial activity of both dichloromethane and methanol extracts was evaluated concentration (20 µg/ml) against two gram-positive bacteria (*Bacillus subtilis*, *Staphylococcus aureus*) and three gram-negative (*Pseudomonas aeruginosa*, *Salmonella typhi*, *Escherichia coli*). The percentage inhibition and MIC50 values (Table 2) shows that both dichloromethane and methanol extracts (FHAD and FHAM) has significant bacterial growth percentage inhibition and MIC50 values are very well comparable with standard ciprofloxacin. The bacterial growth inhibition ranging from 54-74% which is indicating the potential of crude extracts of *Farsetia hamiltonii* regarding presence of anti-bacterial biomolecules.

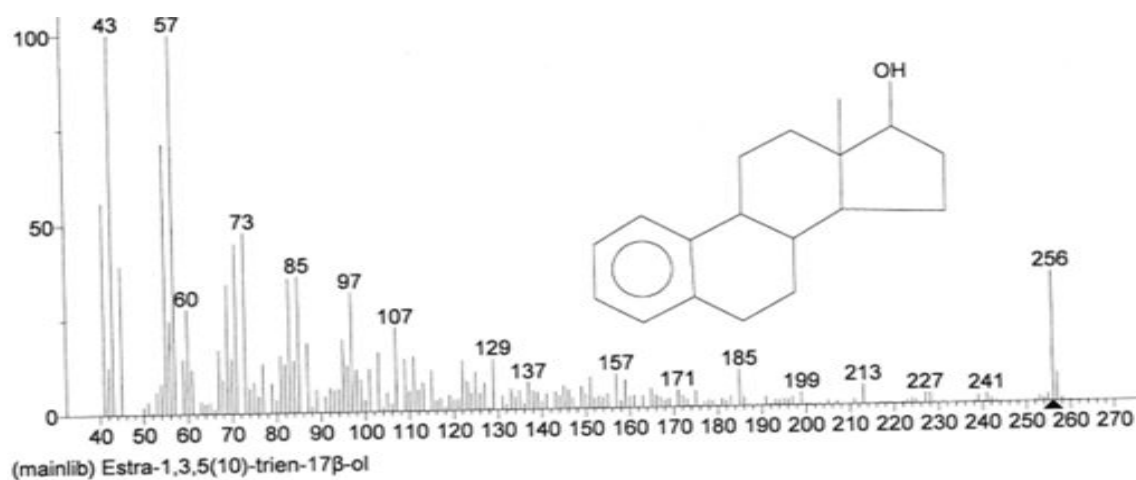
To identify the chemical constituents in the two crude extracts of *F. hamiltonii*, the GC-MS technique was used in the study. The five (5) compounds were identified in the dichloromethane extract (FHAD), whereas four (4) compounds were identified in methanol extract (FHAM) of aerial part of *F. hamiltonii* (Tables 3 and 4) [17-24].

The names of nine (9) identified compounds are: (1) 12-methyl-E,E-2,13-octadecadien-1-ol, (2) estra-1,3,5(10)-



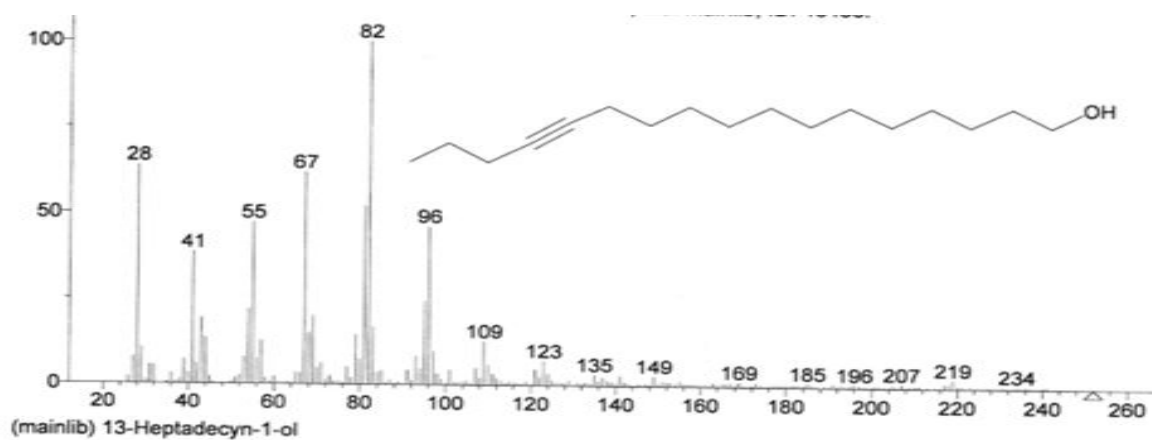
Name: 12-Methyl-E,E-2,13-octadecadien-1-ol
 Formula: $C_{19}H_{36}O$
 MW: 280 Exact Mass: 280.276615 NIST#: 130904 ID#: 17778 DB: mainlib
 Contributor: J. Klune, Insect Chem. Ecol. Lab., USDA, Beltsville, MD 20705

Figure 1. Mass spectra and structure of 12-methyl-E,E-2,13-octadecadien-1-ol.



Name: Estra-1,3,5(10)-trien-17β-ol
 Formula: $C_{18}H_{24}O$
 MW: 256 Exact Mass: 256.182714 CAS#: 2529-64-8 NIST#: 254855 ID#: 7345 DB: mainlib

Figure 2. Mass spectra and structure of estra-1,3,5(10)-trien-17β-ol.



Name: 13-Heptadecyn-1-ol
 Formula: $C_{17}H_{32}O$
 MW: 252 Exact Mass: 252.245316 CAS#: 56554-77-9 NIST#: 36374 ID#: 46436 DB: mainlib

Figure 3. Mass spectra and structure of oleic acid.

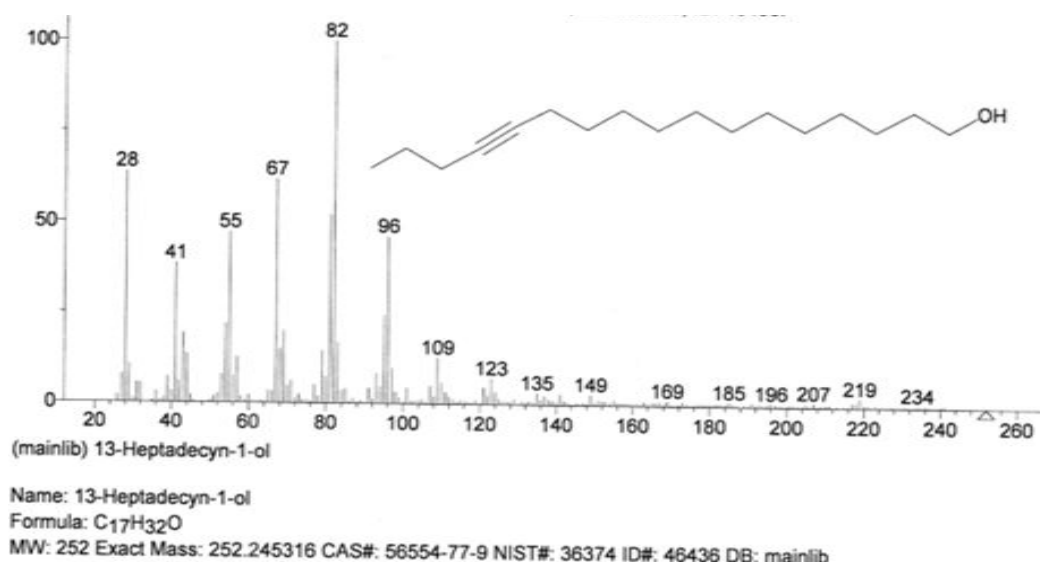


Figure 4. Mass spectra and structure of 13-heptadecyn-1-ol.

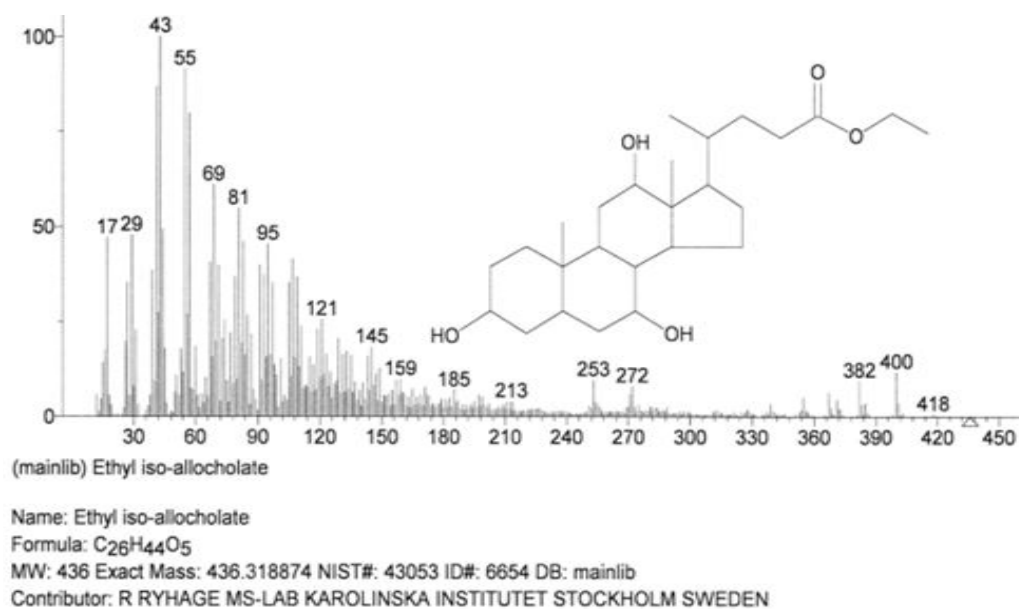


Figure 5. Mass spectra and structure of ethyl iso-allochololate.

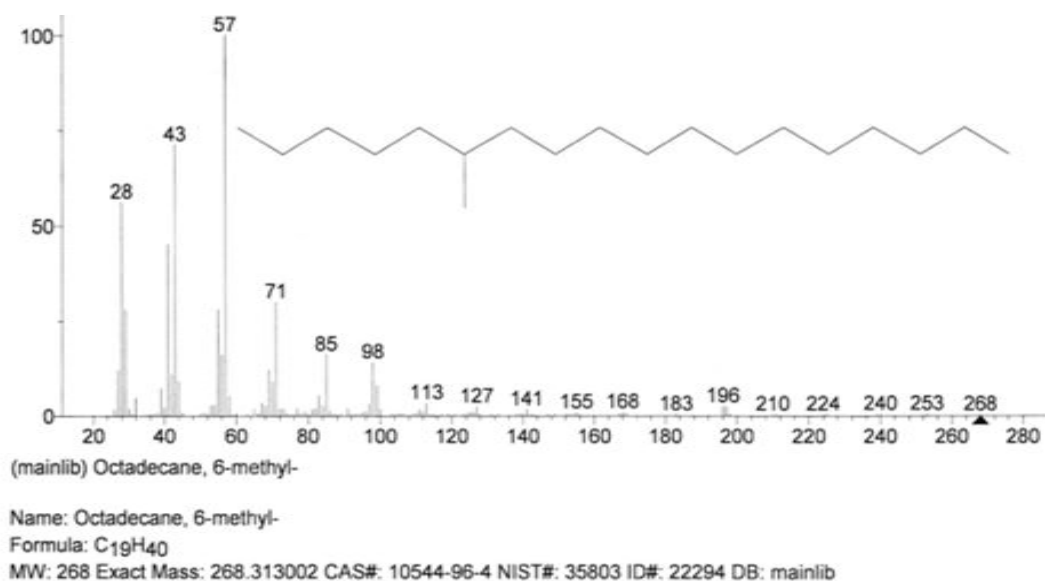


Figure 6. Mass spectra and structure of octadecan, 6-methyl.

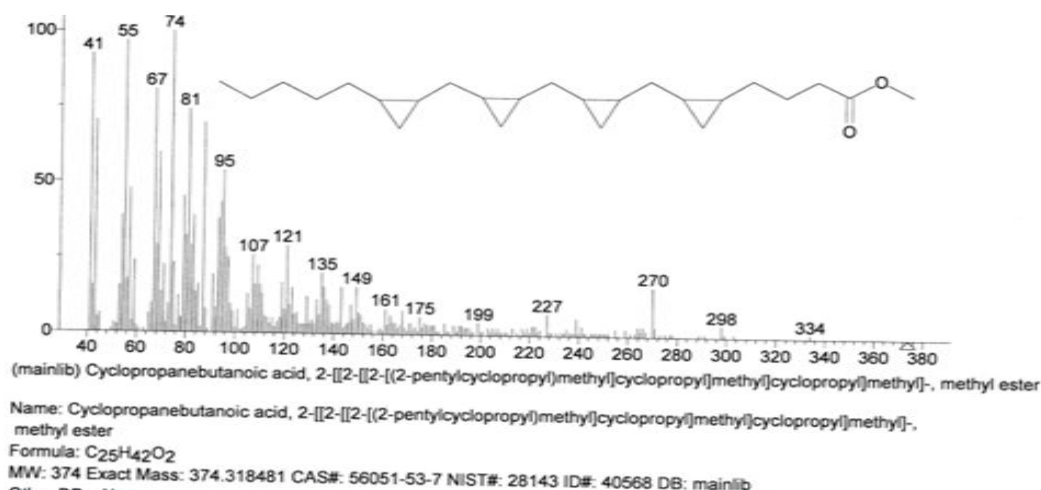


Figure 7. Mass spectra and structure of cyclo propane botanic acid, 2-[[[2-[[[2-(2-pentyl cyclo propyl)methyl] cyclo propyl]methyl]cyclo propyl]methyl]-, methyl ester:

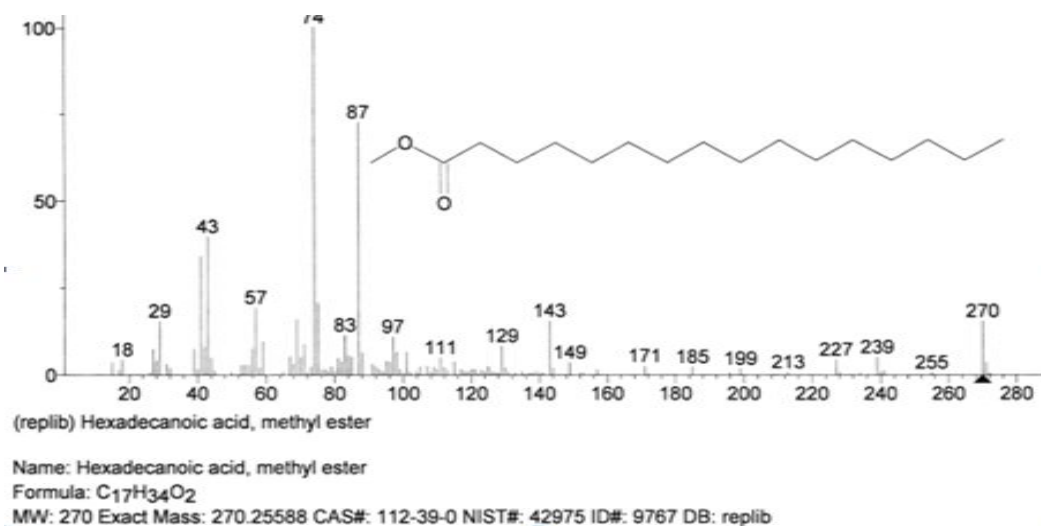


Figure 8. Mass spectra and structure of hexadecanoic acid, methyl ester:

trien-17 β -ol, (3)oleic acid, (4) 13-heptadecyn-1-ol, (5) ethyl iso-allocholate, (6) octadecan, 6-methyl-(7) cyclo propane botanic acid, 2-[[[2-[[[2-(2-pentyl cyclo propyl) methyl] cyclo propyl] methyl]cyclo propyl]methyl]-, methyl ester (8) hexadecanoic acid, methyl ester and (9) ethyl iso-allocholate were identified by performing the analysis of two extracts (FHAD, FHAM) of aerial part of *F. hamiltonii* by GC-MS technique, whereas the ethyl iso-allocholate (Compounds 5 and 9) was identified in both extracts. The Mass spectra of identified compounds are given in Figures 1 to 8. The pharmacological effects or uses of these nine identified compounds are reported in the literature and having anti-microbial, anti-fungal, anti-proliferative, anti-oxidant and anti-inflammatory potentials.

Conclusion

The results of the study validated the folkloric uses of plant *Farsetia hamiltonii* regarding its potential of anti-alphaglucoasidase, anti-chymotrypsin, anti-urease and anti-bacterial activities. Best of our knowledge, this is the first study on biological activities of crude extracts of *Farsetia hamiltonii* provides rational hypothesis for medicinal use of aerial parts of *F. hamiltonii* to treat diabetes, infectious diseases and

gastrointestinal problems. The methanol extract of aerial part of *Farsetia hamiltonii* (Royle) exhibited very strong alpha glucosidase inhibition proving its hypoglycemic use. Further, the potential of nine (09) identified phytochemical constituents from the aerial part of *Farsetia hamiltonii* (Royle) is very importantly reported in literature having anti-microbial, anti-fungal, anti-proliferative, anti-oxidant, anti-inflammatory activities.

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Conflict of Interest

No conflict of interest associated with this study.

Author's Contribution

It is declare that this work was done by the authors named in this article and all liabilities pertaining to claims relating to the content of this article will be borne by them.

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