

## **Evaluation of cyclic nucleotide phosphodiesterase III inhibitors in animal models of epilepsy**

\*J Nandhakumar and #Manoj G Tyagi

#Department of Pharmacology, Christian Medical College, Vellore, India

\*Nandha College of Pharmacy, Erode, Tamil Nadu, India

**Key words:** PDE-3 inhibitors, amrinone, milrinone, mice, seizures.

*Accepted November 17 2007*

### **Abstract**

**This study was conducted to investigate the role of specific phosphodiesterase-3 (PDE-3) inhibitors like amrinone and milrinone in the generation of seizures in albino Swiss mice. Generation of seizures were carried out in the animals by subjecting them to injection of a chemical convulsant, isoniazid (INH) at the dose of 500mg/kg, s.c and by subjecting them to maximal electroshock (MES) at 60 mA for 0.2 sec. The animals were pre-treated with various dose levels of amrinone (0.5 mg/kg, 0.6 mg/kg and 0.7 mg/kg, i.p) and milrinone (50µg/kg, 100 µg/kg, 200µg/kg, 300 µg/kg, i.p) 20 mins prior to the INH or MES. The control group of animals received normal saline (5 ml/kg i.p) 20 mins prior to the injection of INH , or before subjecting the animals to MES. PDE-3 inhibitors significantly enhanced the onset of seizures induced by INH and MES. In particular, milrinone potentiated the convulsive phenomenon more significantly ( $p<0.05$  and  $p<0.001$ ) when compared with amrinone.**

### **Introduction**

Epilepsy is one of the most common affliction of human beings with a prevalence rate of approximately 1 % of the total population [1]. Seizure is a characteristic feature in epilepsy and is associated with disordered and rhythmic high frequency discharge of impulses by a group of neurons in the brain. The pathophysiological basis for epileptic disorders is both complex and intricate. The search for newer antiepileptic drugs have focussed the research on cell signaling elements like the cytoskeletal structures, transmembrane enzymes and ion channel modulators.

There is recent evidence that the cyclic nucleotide phosphodiesterases exist in several molecular forms and that these isozymes are unequally distributed in various tissue makes the phosphodiesterases particularly suitable targets for pharmacologic manipulation, for it suggests that by finding selective inhibitors of the different phosphodiesterase isozymes, one may be able to raise the concentration of cyclic nucleotides in discrete cell types [2]. Through the selective inhibition of the major phosphodiesterase isozyme of a diseased tissue, it may then be possible to alter the course of diseases characterized by an abnormal metabolism of cyclic nucleotides.

Twelve members of the family have been identified and these can be further divided into a number of subtypes and splice variants. The PDE types differ in their amino-acid sequence, substrate specificities, kinetic properties, allosteric regulators, inhibitor sensitivities and in their organ, tissue and sub cellular distribution [3,4]. PDE-3 is characterized by its high affinity for cAMP and cGMP. cAMP is postulated to be anticonvulsant while cGMP is considered to be proconvulsant [5]. PDE-3 enzyme is highly expressed in the hippocampus, striatum and other discrete sites of the brain and may affect the influx of  $Ca^{2+}$  ions [6,7].

In mammals, PDE are encoded by at least 19 different genes and PDE isoforms are expressed differently in different tissues [8]. Electroshock has been reported to increase the expression of PDE 4 isoform in rat brains suggesting ECS regulates the activity of cAMP system by modifying PDE isoform expression [9,10]. However these studies were limited to PDE 4 family in the cerebral cortex and the hippocampus.

This study was hitherto designed carefully to examine and investigate the role of cyclic phosphodiesterase III in the generation of convulsive seizures. We used pharmacological tools like amrinone and milrinone to block the

PDE-3 and evaluate the effect on maximal electroshock and chemical convulsant induced seizures in mice.

## Materials and Methods

### Animals used

Swiss Albino mice of either sex weighing between 22-25 g were utilized for this study. The animals were kept under standard laboratory conditions. A 12:12 dark: light cycle was followed during the experiments. Animals had free access to food and water *ad libitum*. The Institutional Animal Ethical Committee approved the protocol of this study.

### Drugs and Chemicals

The following drugs and chemicals were used for conducting this study. Normal saline (0.9%), Nandha College of Pharmacy, Erode. Pentylentetrazole (Sigma, USA), Isoniazid (Fourts India Ltd, Chennai, India), Amrinone (Samarth Pharma, India), Milrinone (Sanofi Synthelabo Ltd, Mumbai, India). Both amrinone and milrinone were diluted with sterile water for injection. Normal saline was administered in a volume of 5ml/kg, i.p.

#### (i) Isoniazid (INH) induced seizures

Albino mice were divided into different groups each containing six animals (n=6). Seizures were induced in the animals by using chemical convulsant, Isoniazid (INH). INH is a GABA synthesis inhibitor, which was injected to induce seizures at the dose of 500 mg/kg, s.c as described earlier [11]. 15 mins prior to the injection of INH the animals were pre-treated with varying doses of amrinone (0.5 mg/kg, 0.6 mg/kg and 0.7 mg/kg, i.p) and milrinone (50µg/kg, 100 µg/kg, 200 µg/kg and 300 µg/kg, i.p). Onset of action, myoclonic jerks, clonus, tonic flexion and mortality were noted and tabulated.

#### (ii) Maximal Electroshock (MES) Method

MES were induced in the animals using a technique described earlier [12]. The animals were pre-treated with amrinone and milrinone in the same dose as mentioned in the corneal electrodes. The animals were subjected to electroshock (60mA/0.2 secs) via the corneal electrodes. After induction of seizures, tonic limb flexion, tonic extensor, clonus, stupor and recovery/mortality of the animals were observed and tabulated.

### Biometric Analysis

The data is represented as mean  $\pm$  SEM. Statistically significant difference was ascertained by 'P' value which is considered significant of  $P < 0.05$  and highly significant of  $P < 0.01$  as comparisons of different groups were done using one way ANOVA followed by Dunnett's test.

## Result

### Evaluation of onset of seizures

#### (i) INH induced seizures

Table 1 and 3 shows the data obtained from experiments conducted with INH induced seizures. In animals treated with normal saline onset of action were noticed  $2830 \pm 52.33$  sec and convulsions appeared  $3065 \pm 45.43$  sec after INH. Amrinone in a dose of 0.5 mg/kg significantly potentiated the onset of action, jerky movements and convulsions ( $p < 0.05$ ) where as the rate of onset of action, jerky movements and convulsions time was reduced significantly in the doses like 0.6 mg/kg and 0.7 mg/kg of amrinone ( $p < 0.001$ ).

Simultaneously the rate of onset of action, jerky movements and convulsion time was reduced at the great extent even in the low doses like (200 µg/mg and 300 µg/mg) of milrinone ( $p < 0.001$ ) considerable mortality (67%) was observed while using amrinone (0.6 mg/kg and 0.7 mg/kg) and milrinone (100 µg/kg, 200 µg/kg and 300 µg/kg).

#### (ii) Maximal Electroshock Test

Table 2 and 4 illustrates the action of various dose levels of amrinone and milrinone against MES induced seizures. In which 0.6 mg/kg and 0.7mg/kg of amrinone produced a gradual reduction in tonic limb flexion significantly ( $p < 0.05$ ) when compared with normal saline. Significant ( $p < 0.001$ ) was observed in stupor phase of convulsion at the dose of 0.6 mg/kg and 0.7 mg/kg of amrinone. Likewise milrinone treated animals showed a significant ( $p < 0.001$ ) reduction in tonic limb tonic extensor and stupor flexion, phases of convulsion in the 200µg/kg and 300 µg/kg dose levels.

Milrinone in the doses like 200 µg/kg and 300µg/kg treated animals produced the significantly reduced the clonus phases of convulsion at the level of  $p < 0.005$  and  $p < 0.001$  respectively. Mortality (67%) was observed in both doses like 200µg/kg and 300µg/kg of milrinone.

**Table 1: Action of various dose levels of amrinone on chemoshock seizures in mice (n=6)**

| Treatment<br>(mg/kg, i.p)       | CHEMO-CONVULSANT<br>ISONIAZID (INH)                             |                   |                   |
|---------------------------------|---|-------------------|-------------------|
|                                 | Onset Time Of Various Phases Of Convulsions (in sec.)<br>Action | Jerky Movements   | Convulsions       |
| Normal Saline<br>(5 ml/kg, i.p) | 2830 ± 52.33  | 3000 ± 46.47      | 3065 ± 45.43      |
| Amrinone<br>(0.5 mg/kg, i.p)    | 2190 ± 45.82*   | 2293.33 ± 44.24*  | 2320 ± 43.42*     |
| Amrinone<br>(0.6 mg/kg, i.p)    | 1938 ± 37.62**  | 2025 ± 39.30**    | 2063.33 ± 34.79** |
| Amrinone<br>(0.7 mg/kg, i.p)    | 1601.67 ± 17.78**   | 1681.67 ± 20.56** | 1730 ± 15.27**    |

Values are mean ± SEM, represents onset time of various phases of convulsion in seconds. Treatments were given 20 mins prior to chemical convulsant injection of INH (500 mg/kg, s.c.). The data were analysed by one-way ANOVA followed by Dunnett's test. \*  $p < 0.05$  and \*\*  $p < 0.001$ , compared to the normal saline treated group.

**Table 2: Action of various dose levels of amrinone on maximal electroshock induced convulsions in mice (n=6)**

| Treatment<br>(mg/kg, i.p)       | Onset Time Of Various Phases Of Convulsions (in sec.) |                |              |               |                |
|---------------------------------|---|----------------|--------------|---------------|----------------|
|                                 | Tonic Limb Flexion                                    | Tonic Extensor | Clonus       | Stupor        | Recovery/Death |
| Normal Saline<br>(5 ml/kg, i.p) | 5.67 ± 0.33   | 23.33 ± 0.67   | 36.83 ± 1.38 | 66.67 ± 1.23  | 196.25 ± 5.55  |
| Amrinone<br>(0.5 mg/kg, i.p)    | 4.33 ± 0.21   | 19.83 ± 0.31   | 37 ± 0.73    | 59.83 ± 0.6*  | 226 ± 0.82     |
| Amrinone<br>(0.6 mg/kg, i.p)    | 3.5 ± 0.22 *  | 15.5 ± 0.43 *  | 33.33 ± 0.42 | 53.83 ± 0.6** | 207 ± 3.72     |
| Amrinone<br>(0.7 mg/kg, i.p)    | 3.17 ± 0.17 *   | 9.83 ± 0.31**  | 28 ± 0.51*   | 43 ± 0.86**   | 207.33 ± 0.42  |

Values are mean ± SEM, represents onset time of various phases of convulsion in seconds. Treatments were given 20 mins prior to maximal electroshock (60 mA, 0.2 sec). The data were analysed by one-way ANOVA followed by Dunnett's test. \*  $p < 0.05$  and \*\*  $p < 0.001$ , compared to the normal saline treated group.

**Table 3: Action of various dose levels of milrinone on chemoshock seizures in mice (n=6)**

| Treatment<br>(mg/kg, i.p)       | CHEMO-CONVULSANT<br>ISONIAZID (INH)                             |                 |                |
|---------------------------------|---|-----------------|----------------|
|                                 | Onset Time Of Various Phases Of Convulsions (in sec.)<br>Action | Jerky Movements | Convulsions    |
| Normal Saline<br>(5 ml/kg, i.p) | 2830 ± 52.33  | 3000 ± 46.47    | 3065 ± 45.43   |
| Milrinone<br>(50 µg/kg, i.p)    | 2520 ± 34.65  | 2860 ± 25.31    | 3060 ± 21.92   |
| Milrinone<br>(100 µg/kg, i.p)   | 2220 ± 34.65*   | 24833 ± 26.04*  | 2610 ± 25.71*  |
| Milrinone<br>(200 µg/kg, i.p)   | 1870 ± 36.04**  | 2100 ± 30.98**  | 2270 ± 36.04** |
| Milrinone<br>(300 µg/kg, i.p)   | 1670 ± 21.91**  | 1890 ± 25.71**  | 2010 ± 33.76** |

Values are mean ± SEM, represents onset time of various phases of convulsion in seconds. Treatments were given 20 mins prior to chemical convulsant injection of INH (500 mg/kg, s.c.). The data

were analysed by one-way ANOVA followed by Dunnett's test. \*  $p < 0.05$  and \*\*  $p < 0.001$ , compared to the normal saline treated group.

**Table 4: Action of various dose levels of milrinone on maximal electroshock induced convulsions in mice (n=6)**

| Treatment<br>(mg/kg, i.p)       | Onset Time Of Various Phases Of Convulsions (in sec.) |                |               |                |                 |
|---------------------------------|---|----------------|---------------|----------------|-----------------|
|                                 | Tonic Limb Flexion                                    | Tonic Extensor | Clonus        | Stupor         | Recovery/ Death |
| Normal Saline<br>(5 ml/kg, i.p) | 5.67 ± 0.33   | 23.33 ± 0.67   | 36.83 ± 1.38  | 66.67 ± 1.23   | 196.25 ± 5.55   |
| Milrinone<br>(50 µg/kg, i.p)    | 4.8 ± 0.33  | 18.3 ± 0.41    | 36.8 ± 0.41   | 57.5 ± 0.78*   | 218.3 ± 0.86    |
| Milrinone<br>(100 µg/kg, i.p)   | 3.3 ± 0.20*   | 15.2 ± 0.49*   | 32.7 ± 0.82   | 53.5 ± 0.65**  | 280 ± 3.14      |
| Milrinone<br>(200 µg/kg, i.p)   | 2.7 ± 0.20 **   | 8.7 ± 0.3 3**  | 25.8 ± 0.33 * | 47.7 ± 0.65**  | 235.0 ± 1.71    |
| Milrinone<br>(300 µg/kg, i.p)   | 2.2 ± 0.17 **   | 7.3 ± 0.33**   | 20 ± 0.57 **  | 39.8 ± 0.69 ** | 237.5 ± 1.43    |

Values are mean ± SEM, represents onset time of various phases of convulsion in seconds. Treatments were given 20 mins prior to maximal electroshock (60 mA, 0.2 sec). The data were analysed by one-way ANOVA followed by Dunnett's test. \*  $p < 0.05$  and \*\*  $p < 0.001$ , compared to the normal saline treated group.

## Discussion

The therapeutic use of theophylline/ aminophylline is associated with the incidence of intractable seizures and mortality [13,14]. The mechanisms involved in these seizures are not well understood and the treatment of the life threatening condition is unsatisfactory. The results of this study suggest that PDE-3 inhibitors potentiate the electroshock and chemical induced seizures. The bipyridine derivative of selective PDE-3 inhibitors such as amrinone and milrinone is a new class of positive inotropic drugs chemically and pharmacologically distinct from digitalis and catecholamines [15,16]. The mechanism of the positive inotropic effect of PDE inhibitors is similar to that of  $\beta$ -adrenergic agents [17]. Milrinone has been the most studied and used extensively as PDE-3 inhibitor and it is currently used in the acute treatment of heart failure to diminish long term risk [18]. This study demonstrates the importance of the PDE-3 inhibitors such as amrinone and milrinone in the generation of seizure activity with the accumulation of cellular levels of cAMP and cGMP by inhibiting its metabolism. cAMP accumulation is considered to be anticonvulsant and cGMP is considered to be proconvulsant.

The data obtained from this study show that pre-treatment with PDE-3 inhibitors potentiates the onset of action and various phases of convulsions against INH and maximal electroshock induced convulsions as depicted in Table 1 to 4. Our study results also clearly suggest that rate of onset of convulsive time was significantly ( $p < 0.05$  and  $p < 0.001$ ) reduced with increasing the dose levels of both amrinone and milrinone against INH and MES induced seizures.

Earlier studies suggest that the elevated levels of cGMP was found in cortical structure in some experimental mo-

dels of epilepsy [19,20], and the neuronal excitability was regulated by cGMP and  $Ca^{2+}$ /calmodulin dependent protein kinase and its phosphorylation process [21]. Apart from these findings, PDE-3 inhibitors possess transmembrane influx of  $Ca^{2+}$ . This influx of  $Ca^{2+}$  is responsible for the phosphorylation process of intracellular proteins, such as ion channels, receptors, enzymes and transcription factors which exhibit significant neuronal excitability and epileptic seizures [22].

On the other hand, phosphorylation of variety of substrates regulates the myriad of physiological process, such as immune responses, cardiac and smooth muscle contraction, visual response, glycogenolysis, platelet aggregation, ion channel conductance, apoptosis and growth control [23]. The present study results also early correspond with the generation of seizure activity due to the breakdown of hydrolysis of cGMP which promotes protein kinase phosphorylation process.

Thus, in conclusion the study shows a definite relationship between the specific PDE-3 inhibitors and increase the cellular level of cGMP and  $Ca^{2+}$  ions with the generation of seizures. The release of free radicals have been implicated in many drug and chemical induced toxicities [24]. It is possible that increased production of reactive oxygen species could result in oxidant/ anti-oxidant imbalance and thus, precipitate neurotoxicity. Therefore it appears that non nucleotide mechanism although not well defined could also be contributing significantly to the seizure activity of phosphodiesterase 3 inhibitors.

## Acknowledgement

The authors are thankful to the CMCH Pharmacy, Vellore for the arrangement of Amrinone (Samarth Pharma, India) and Milrinone (Sanofi Synthelabo Ltd, Mumbai, India). The authors also thankful to Thiru. V. Shanmugan, Chairman and Mr. S. Nandakumar Pradeep, Secretary, Nandha College of Paramedical Sciences, Erode 638052, Tamilnadu, India for providing infrastructural facilities to carry out this project and our colleagues who helped directly and indirectly for this study.

## References

- Delgado-Escueta AV, Treiman DM, Walsh GO. The treatable epilepsies. *N. Engl J Med* 1983; 308: 1508-1514.
- Jeon YH, Heo YS, Kim CM, Hyun YL, Lee TG, Ro S, Cho JM. Phosphodiesterase : overview of protein structures, potential therapeutic applications and recent progress in drug development. *Cell Mol Life Sci* 2005; 62: 1198-1220.
- Smith CJ, Krall J, Manganiello VC, Movsesian MA. Cytosolic and sarcoplasmic reticulum-associated low Km, cGMP-inhibited cAMP phosphodiesterase in mammalian myocardium. *Biochem Biophys Res Commun* 2005; 190: 521-561.
- Shakur Y, Holst LS, Landstorm TR, Mowsesian M, Degermen E, Manganiello V. Regulation and function of the cyclic nucleotide phosphodiesterase (PDE3) gene family. *Prog. Nucleic. Acid Res Mol Biol* 2001; 66: 241-277.
- Ray A, Gulati K, Anand S, Vijayan VK. Pharmacological studies on mechanisms of aminophylline-induced seizures in rats. *Ind J Exp Biol* 2005; 43: 849-853.
- Cho CH, Cho DH, Seo MR, Juhn YS. Differential changes in the expression of cyclic nucleotide phosphodiesterase isoforms in rat brains by chronic treatment with electroconvulsive shock. *Exp Mol Med* 2000; 32: 110-114
- Liu H, Maurice DH. Expression of cyclic GMP-inhibited phosphodiesterases 3A and 3B (PDE3A and PDE3B) in rat tissues: differential subcellular localization and regulated expression by cyclic AMP. *Br J Pharmacol* 1998; 125: 5101-1510
- Beavo JA. Cyclic nucleotide phosphodiesterases: functional implications of multiple isoforms. *Physiol Rev* 1995; 75: 725-748
- Suda S, Nibuya M, Ishiguro T, Suda H. Transcriptional and translational regulation of phosphodiesterase type IV isozymes in rat brain by electroconvulsive seizure and antidepressant drug treatment. *J Neurochem* 1998; 71: 1554-1563
- Takahashi M, Terwilliger R, Lane C, Mezes PS, Conti M, Duman RS. Chronic antidepressant administration increases the expression of cAMP- specific phosphodiesterase 4A and 4B isoforms. *J Neurosci* 1999; 19: 610-618
- Costa E, Guidotti A, Mao CL. Evidence for involvement of GABA in the action of benzodiazepines; studies on rat cerebellum. In: Costa E, Greengard P (eds) *Mechanisms of action of benzodiazepines. Advances in Biochemical Psychopharmacology, Vol 14.* Raven Press, New York 1975. pp. 113-151
- Tyagi MG, Jose VM. Anticonvulsant effect of cytoskeletal depolymerizers in combination with potassium channel opener and adenylate cyclase activator; a causative link with nerve growth factor? *Ind J Exp Biol* 2001; 39: 425-430,
- Barnes PJ. Theophylline: in Asthma: Basic mechanism and clinical management, edited by Barnes PJ, Rodger IW and Thompson N C, (Academic Press, San Diego, USA) 1998, 689
- Barnes PJ, Pauwels RA. Theophylline in the management of asthma: Time for reappraisal. *Eur Respir J* 1995; 7: 579
- Peter Honerjager. Pharmacology of bipyridine phosphodiesterase 3 inhibitors. *American Heart Journal* 1991; 121: 1939-1944
- Masaaki Ito, Toshio Tanaka, Masahiro Saitoh, Hiroshi Masuoka, Takeshi N, Hiroyoshi H. *Biochemical pharmacology* 1988; 37: 2041- 2044
- Cruickshank JM. Phosphodiesterase III inhibitors: long term risks and short-term benefits. *Cardiovasc Drugs Ther.* 1993; 7: 655-660
- Wetzel B, Huel N. New cardiotonic agents- a promising approach for treatment of heart failure. *TIPS* 1998; 9: 166-170
- Vullimoz Y, Verosky M, Alpert M, Triner L. Effect of enflurane on cerebellar cGMP and on motor activity in the mouse. *Br.J.Anesthesia* 1983; 55: 79-84
- Riazi K, Roshanpour M, Rafei-Tabatabaei N, Homayoun H, Ebrahimi F, Dehpour AR. The proconvulsant effect of sildenafil in mice: role of nitric oxide-cGMP pathway. *Br.J. Pharmacol* 2006; 147: 935-943
- Walaas S.I., Greengard P. Protein phosphorylation and neuronal function. *Pharmacol. Rev* 1991; 3: 299-349
- Butler LS, Silva AJ, Abeliovich A, Watanabe Y, Tonogawa S, Mc Narama JO. Limbic epilepsy in Transgenic mice carrying a Ca<sup>2+</sup>/Calmodulin- dependent kinase II alpha-subunit mutation. *Proc Natl Acad Sci USA.* 1995; 15: 6852- 6855
- Francis SH, Turko IV, Corbin JD. Cyclic nucleotide phosphodiesterases: relating structure and function. *Prog. Nucleic Acid Res. Mol Biol* 2001; 65: 1-52
- Lebel CP and Bondy S C. Oxygen radicals: Common mediators of neurotoxicity. *Neurotoxicol Teratol* 1991; 13: 314

## Correspondence:

Manoj G Tyagi  
Department of Pharmacology  
Christian Medical College  
Vellore 632002, TN  
India

Phone: +91-416-228-4237 Fax: +91-416-2262788

e-mail: [tyagi243@yahoo.co.in](mailto:tyagi243@yahoo.co.in)

25. Butler LS, Silva AJ, Abeliovich A, Watanabe Y, Tonogawa S, Mc Narama JO. Limbic epilepsy in Transgenic mice carrying a Ca<sup>2+</sup> / Calmodulin- dependent kinase II alpha-subunit mutation. Proc. Natl. Acad. Sci USA. 1995; 15: 6852- 6855
26. Francis S.H., Turko I.V., and Corbin J.D. Cyclic nucleotide phosphodiesterases: relating structure and function. Prog. Nucleic Acid Res. Mol. Biol. 2001; 65: 1-52
27. Lebel CP and Bondy S C. Oxygen radicals: Common mediators of neurotoxicity. Neurotoxicol Teratol. 1991, 13 , 314

**Correspondence:**

Manoj G Tyagi  
Department of Pharmacology  
Christian Medical College  
Vellore 632002, TN  
India

Phone: +91-416-228-4237  
Fax: +91-416-2262788  
e-mail: tyagi243@yahoo.co.in

**Biomedical Research**

87-Greater Azad Enclave, Aligarh 202 002, India

*e-mail: biomedical44@hotmail.com*

Dr. Manoj G Tyagi  
Department of Pharmacology  
Christian Medical College Vellore  
Vellore

**INVOICE**

|    | <b>Details</b>   | <b>Amount</b> |
|----|--|---------------|
| 1. | Evaluation of cyclic nucleotide phosphodiesterase III inhibitors in animal models of epilepsy<br><br><i>Authors: J Nandhakumar and Manoj G Tyagi</i><br><br><i>Biomedical Research 2008; 19 (1):</i> |               |
| 2. | Charge for 5 printed pages (Rs.1000.00/page)   | Rs. 5000.00   |

|              |                                   |                    |
|--------------|-----------------------------------|--------------------|
| <b>Total</b> | <b>Rupees Five Thousands Only</b> | <b>Rs. 5000.00</b> |
|--------------|-----------------------------------|--------------------|

Payment may be made by Bank Draft in favour of:

**Biomedical Research, Account No. 010975840855**  
State Bank of India, Main Branch, Aligarh 202 002, India  
**Sort code: SBI NIN BB 221**

Biomedical Research  
6-B Manzar, Sir Syed Nagar  
Aligarh 202 002

September 28, 2007

e-mail: [biomedical44@hotmail.com](mailto:biomedical44@hotmail.com)  
mobile: 0091-571935825514