The effects of hesperidin on idiopathic pulmonary fibrosis evaluated by histopathological-biochemical and micro-computed tomography examinations in a bleomycin-rat model.

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Abstract

Idiopathic Pulmonary Fibrosis (IPF) is a chronic, progressive parenchymal lung disease. The pathology is characterized by recurrent injury to microscopic alveolar epithelial cells. These injuries activate inflammatory cells, resulting in the proliferation of fibroblasts and alveolar tissue damage. Interstitial inflammation, advanced oxidative stress, and abnormal antioxidant activity were demonstrated to be the main causes of IPF. Hesperidin (HP) is a bioflavonoid with anti-inflammatory, antioxidant, anticarcinogenic, and analgesic actions. HP may be able to prevent pulmonary fibrosis, and may ultimately lead to healthy lung function. We hypothesized that HP could prevent Bleomycin (BLC)-induced pulmonary fibrosis due to its biochemical, antioxidant, and anti-inflammatory properties and may ultimately lead to healthy lung function. Based on these findings, we hypothesized that HP could prevent BLC-induced pulmonary fibrosis due to its biochemical, antioxidant, and anti-inflammatory properties. The animals were divided into 4 groups with 14 rats per group. The experimental treatments were as follows: Control, BLC, HP, and BLC+HP. Six of the 14 lungs in each group were sent for micro-CT analysis. The remaining 8 lung specimens were harvested for histopathological and biochemical analyses. BLC-treated rats showed marked histopathological changes in the lungs. In these rats, thickening of interalveolar septa due to macrophage and lymphocyte infiltration, as well as fibroblast proliferation, were observed. Histopathological changes were less severe in the BLC+HP group compared with the BLC group. HP treatment led to a decrease in lipid peroxidation and an increase in antioxidant status compared with the BLC group. Also micro-CT showed a significant positive correlation with histopathological and biochemical results. To the best of our knowledge, this is the first study to evaluate the beneficial effects of HP against pulmonary fibrosis using histopathological, biochemical, and micro-CT analyses and HP successfully minimized the severity of BLC-induced lung injury, which was used as a model for IPF.

Keywords: Idiopathic pulmonary fibrosis, Micro-computed tomography, Hesperidin.

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Introduction

Idiopathic Pulmonary Fibrosis (IPF) is a chronic, diffuse, progressive parenchymal lung disease and is the most common form of interstitial pneumonia [1]. The incidence of IPF is 6.8-8.8 cases per 100,000 [2]. This progressive lung disease has a poor prognosis with a median survival of only 2-3 years. The pathology is characterized by recurrent injury to microscopic alveolar epithelial cells. These injuries activate inflammatory cells, which release harmful amounts of Reactive Oxygen Species (ROS), resulting in the proliferation of fibroblasts and alveolar tissue damage [3]. Myeloperoxidase is an enzyme that is released from activated neutrophils and interacts with Hydrogen Peroxide (H₂O₂), which then forms highly toxic hydroxyl radicals [4]. Repetitive injury, failed repair, fibrosis, and deposition of extreme Extracellular Matrix (ECM) result in pathologic tissue scarring that negatively affects parenchymal architecture. In addition, interstitial inflammation, advanced oxidative stress, and abnormal antioxidant activity were demonstrated to be the main causes of IPF [5,6]. Therefore, as an adjuvant therapy, supplementing patients with anti-inflammatory and antioxidant agents may help prevent and treat IPF.
Animal models of lung fibrosis have been useful for understanding the fibrotic process. Bleomycin (BLC)-induced pulmonary fibrosis is the most widely used animal model of human IPF. Intratracheal application of BLC causes the proliferation of fibroblasts, an inflammatory response in alveolar epithelial cells, and the deposition of collagen [7,8]. In this study, antioxidant agents were tested in a BLC-induced lung fibrosis animal model, and were found to be beneficial in preventing fibrosis [9-11].

Flavonoids are a class of plant pigments, and Hesperidin (HP) is a bioflavonoid with anti-inflammatory, antioxidant, anti-carcinogenic, and analgesic actions [12,13]. In addition, HP was found to be protective against the toxic effects of cisplatinum in rat eyes, and was reported to activate antioxidant function with its strong free oxygen radical scavenging activity [14] thus, HP may be able to prevent pulmonary fibrosis caused by interstitial inflammation and advanced oxidative stress due to its anti-inflammatory and anti-oxidative properties, and may ultimately lead to healthy lung function.

Based on these findings, we hypothesized that HP could prevent BLC-induced pulmonary fibrosis due to its biochemical, antioxidant, and anti-inflammatory properties. Therefore, we investigated oxidative status, histopathological changes, and micro-Computed Tomography (micro-CT) in the lungs of rats.

Materials and Methods

Chemicals

BLM was obtained from Onko Medical Company (BLEOCIN-S®, 15 mg, Istanbul, Turkey) and HP was obtained from Molecula Limited (Gillingham, UK). All other chemicals were purchased from Sigma Chemical Co. (St Louis, MO, USA) and were either analytical grade or the highest grade available.

Study design

This experimental study was approved by the Animal Ethics committee (reference number, 2014/A-81). Experiments were performed based on the Animal Welfare Act and the Guide for the Care and Use of Laboratory animals (National Institutes of Health [NIH] publication No. 5377-3, 1996), Local Animal Ethics Committee. In total, 42 healthy adult male Sprague-Dawley rats (3-4 months of age and 250-300 g in weight) were obtained from the Experimental Animal Institute, Malatya, Turkey. The animals were placed in sterilized polypropylene rat cages, and housed in a room maintained at 21 ± 2°C and 60 ± 5 % humidity, with a 12-h light-dark cycle.

The animals were randomly divided into 4 groups with 14 rats per group. The experimental treatments were as follows: Control, BLC, HP, and BLC + HP. Rats in the control group received intratracheal injection (2.5 mg/kg body weight) in 0.25 mL Phosphate Buffered Saline (PBS) using corn oil as a vehicle. In the BLC group, a single intratracheal injection of BLC hydrochloride (2.5 mg/kg body weight) in 0.25 mL PBS was administered while the rats were anesthetized to achieve lung fibrosis. In the HP group, 50 mg/kg/day HP was given by gavage for 14 consecutive days. In the BLC+HP group, 50 mg/kg oral HP was given 1 day before the intratracheal BLC injection and continued for 14 days. These procedures were applied as the similar studies that presented before [11,14]. Fourteen days after the BLC injection, all rats were sacrificed by an overdose of anesthesia. Six of the 14 lungs in each group were sent for micro-CT analysis. The remaining 8 lung specimens were harvested for histopathological and biochemical analyses. The right section of each lung was placed in liquid nitrogen and stored at -70°C until tested for Thiobarbituric Acid Reactive Substances (TBARS), Catalase (CAT), Superoxide Dismutase (SOD), Glutathione Peroxidase (GPx), and reduced Glutathione (GSH). The left section of each lung was placed in formaldehyde prior to histopathological evaluation under light microscopy. Previous dose-response studies on anti-inflammatory and anti-oxidative effects in rats and lung fibrosis were used to choose the HP and BLC doses [15-17].

Micro-CT scanning and image analysis

After sacrifice, 5 mL of air was introduced into the trachea using a syringe. The trachea was then bound to prevent air leakage. The lungs were extracted and scanned using a desktop micro-CT system (SkyScan 1072, SkyScan, Kontich, Belgium) to evaluate and quantify the total lung injury area.

Each specimen was mounted on the computer-controlled turntable such that the x-ray beam was perpendicular to the horizontal axes of the rat lung. Digital section images were acquired under the following conditions: 100 mA beam current, 100 kV accelerating voltage, 0.5 mm aluminum filter, 27.37 μm pixel size at 1,000 × 1,000 resolution, and 180° rotation at the 0.3° step. About 650-700 cross-sectional images were taken from each sample using an 11 Mp camera. The full scan time duration was 55-60 min. All three-dimensional (3D) images were evaluated on a 17-inch TFT computer screen by the same operator.

Figure 1. Evaluation of micro-CT images.

The micro-CT images were evaluated using the CTAn 1.13.5.1 program (SkyScan, Kontich, Belgium). Ground glass densities and peripheral patchy consolidation indicated lung injury. The right and left lower zones of the lungs were separated and cylinder shapes (each 1.5 mm × 8 mm) were chosen as the
Regions of Interest (ROIs). The CTAn 1.13.5.1 program (SkyScan, Kontich, Belgium) was used to analyze the multiplanar images in the axial, coronal, and sagittal planes for each reconstructive lung image, and to calculate the lung injury volume fraction (injury volume/total volume) (Figure 1).

**Biochemical assays**

Tissue was homogenized in a Teflon glass homogenizer in 150 mM KCl (pH 7.4) to obtain a 1:10 (w/v) dilution of the whole homogenate. The homogenates were centrifuged at 18,000 g (4°C) for 30 min to determine TBARS content, reduced Glutathione (GSH) concentration, and CAT activity, and at 25,000 g for 50 min to determine glutathione peroxidase (GSH-Px) activity. The levels of TBARS, an indicator of lipid peroxidation, were determined in 1 g homogenized lung tissue by the thiobarbituric acid reaction using previously described methods [18]. The product was evaluated spectrophotometrically at 532 nm and the results were expressed as nmol/g tissue. The reduced GSH content of the lung homogenate was measured at 412 nm using the methods of Sedlak and Lindsay [19]. GSH levels were expressed as nmol/mL. CuZn-SOD activity was measured by the inhibition of Nitroblue Tetrazolium (NBT) reduction due to O$_2^-$ generated by the xanthine/xanthine oxidase system [20]. One unit of SOD activity was defined as the amount of protein resulting in 50% inhibition of the NBT reduction rate. The product was evaluated spectrophotometrically at 560 nm. Results are expressed as IU/mg protein.

CAT activity was determined according to the method of Aebi et al. [21]. The enzymatic decomposition of H$_2$O$_2$ was followed directly by a decrease in absorbance at 240 nm. The difference in absorbance per unit time was used as a measure of CAT activity. The enzyme activities are shown in kU/mg protein. GSH-Px activity was measured using the methods of Paglia and Valentina [22]. In the presence of glutathione reductase and NADPH, the oxidized Glutathione (GSSG) is immediately converted to the reduced form with a concomitant oxidation of NADPH to NADP. The decrease in absorbance at 340 nm was measured. GSH-Px activity was expressed as IU/mg protein. Tissue protein content was determined according to the method developed by Lowry et al. [23], using Bovine Serum Albumin (BSA) as the standard.

**Histopathological examination**

After sacrifice, the lungs were removed and fixed in 10% neutral buffered formalin. The lung tissues (3 specimens/animal) were processed for routine paraffin embedding, and sectioned at a 5 μm thickness. The cross sections were stained with hematoxylin & eosin for histological examination. The scoring system described by Ashcroft et al. [24], was used to estimate the severity of pulmonary fibrosis on a numerical scale (Table 1). Using a 10X objective, an average of 10 fields per slide were assessed for pulmonary fibrosis severity. Each field was assigned a score between 0 (normal lung) and 8 (total fibrosis). The mean of the scores for all fields was taken as the fibrotic score of the section.

**Table 1. Criteria for grading lung fibrosis described by Ashcroft et al.**

<table>
<thead>
<tr>
<th>Score</th>
<th>Histological features</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>Normal lung</td>
</tr>
<tr>
<td>1</td>
<td>Minimal fibrous thickening of alveolar or bronchiolar walls</td>
</tr>
<tr>
<td>2</td>
<td>Moderate thickening of walls without obvious damage to lung architecture</td>
</tr>
<tr>
<td>3</td>
<td>Increased fibrosis with definite damage to lung structure and formation of fibrous bands or smallfibrous masses</td>
</tr>
<tr>
<td>4</td>
<td>Severe distortion of structure and large fibrous areas</td>
</tr>
<tr>
<td>5</td>
<td>Total fibrous obliteration of the field</td>
</tr>
</tbody>
</table>

Means bearing different superscripts within same column were significantly different (P<0.05).

**Figure 2.** A and B. Photomicrographs showing the normal histological appearance of the lungs in the control (A) and the HP-only group (B) rats. Hematoxylin and eosin. C. Photomicrograph of the lung of a rat in the BLC-only group showing the thickening of the interalveolar septa by mononuclear cell infiltration and fibroblast proliferation. Hematoxylin and eosin. D. Photomicrograph of the lung of a rat in the BLC+HP group. Note that the histopathological changes were less severe compared with the BLC-only group (C). Hematoxylin and eosin. Abbreviations : HP; hesperidin, BLC; Bleomycin.
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Statistics

Data analysis was performed using IBM SPSS Statistics 22.0 (IBM, Armonk, NY, USA). Data are presented as the median (min-max). The Kruskal-Wallis test was used to statistically evaluate the data among the groups. The Conover multiple comparison test was used to differentiate one group from the other. A P-value<0.05 was considered statistically significant.

Results

Histopathological findings

The lung tissue sections of the control and HP-treated groups showed normal histological structure (Figures 2A and 2B). BLC-treated rats showed marked histopathological changes in the lungs. In these rats, thickening of interalveolar septa due to macrophage and lymphocyte infiltration, as well as fibroblast proliferation, were observed (Figure 2C). Neutrophils were observed in both the alveoli and interstitium. The alveolar architecture was damaged by desquamation of type I pneumocytes, proliferation of type II pneumocytes, and accumulation of foamy macrophages. Emphysematous changes, including fragmentation of alveolar septa and expansion of alveolar spaces, were also seen. Histopathological changes were less severe in the BLC+HP group compared with the BLC group (Figure 2D). The BLC group had the highest Ashcroft fibrosis score (3.24 ± 0.35). Although there was a marked reduction in fibrosis between the BLC and BLC+HP groups (Ashcroft score: 1.71 ± 1.20) the difference was not statistically significant (Table 2).

Biochemical results

The SOD, GPx, GSH, and TBARS levels are shown in Table 3. BLC treatment resulted in a significant increase in the TBARS levels and a significant decrease in the SOD, GPx, and GSH levels compared with the other groups. The TBARS levels and SOD activity were significantly different in the HP-treated groups compared with the control group. There were no significant changes in oxidant/antioxidant parameters between the control and HP groups. Additionally, BLC+HP combined led to a significant decrease in the TBARS levels, and a significant increase in SOD, GPx, and GSH levels. HP treatment led to a decrease in lipid peroxidation and an increase in antioxidant status compared with the BLC group.

Micro-CT findings

The severity of lung injury was 1.52% in the control group and 1.75% in the HP group due to traumatic damage caused while harvesting (Figures 3A and 3B). There were no significant differences in the lesion size between these groups. In the BLC group, the lung injury volume fraction was 14.4%. The BLC+HP group showed reduced fibrosis severity with a score of 4.38% (Figures 3C and 3D), which was significantly different from the BLC group (Figure 4).

Table 3. The levels of SOD, GSH and TBARS in rat lung tissue (Mean ± SD).

<table>
<thead>
<tr>
<th>Group</th>
<th>TBARS (nmol/g tissue)</th>
<th>Reduced GSH nmol/ml</th>
<th>SOD (U/mg protein)</th>
<th>GPx (U/mg protein)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>8.81 ± 0.68^a</td>
<td>51.5 ± 8.95^a</td>
<td>30.1 ± 4.74^a</td>
<td>221.5 ± 14.2^a</td>
</tr>
<tr>
<td>Hesperidin</td>
<td>5.36 ± 0.74^b</td>
<td>52.5 ± 4.01^a</td>
<td>29.0 ± 3.00^b</td>
<td>219.5 ± 22.1^a</td>
</tr>
<tr>
<td>Bleomycin</td>
<td>13.2 ± 0.84^c</td>
<td>35.7 ± 5.24^b</td>
<td>17.2 ± 1.21^c</td>
<td>141.2 ± 13.0^b</td>
</tr>
<tr>
<td>Bleomycin+hesperidin</td>
<td>10.1 ± 1.24^d</td>
<td>41.6 ± 3.83^c</td>
<td>22.4 ± 2.31^d</td>
<td>178.5 ± 21.1^c</td>
</tr>
</tbody>
</table>

Means with different superscripts within the same column were significantly different (P<0.01). SOD: Superoxide Dismutase, GSH: reduced Glutathione, TBARS: Thiobarbituric Acid Reactive Substances, SD: Standard Deviation

Figure 3. Severity of lung injury in control (A), HP (B), BLC (C) and BLC+HP (D) groups.

Figure 4. Comparison of lung injury evaluated via micro-CT between groups. HP: Hesperidin, BLC: Bleomycin.
Discussion

To the best of our knowledge, this is the first study to evaluate the beneficial effects of HP against pulmonary fibrosis using histopathological, biochemical, and micro-CT analyses. This study demonstrated that HP treatment has promising preventive effects on BLC-induced lung fibrosis. BLC causes the proliferation of fibroblasts, an inflammatory response in alveolar epithelial cells, and the deposition of collagen [7,8]. BLC-induced pulmonary fibrosis is a widely used and well-established model of IPF [25]. There are several dosing methods; however, intratracheal BLC administration is the most popular mode of administration. Lung injury occurs after the generation of oxidant species through iron-dependent mechanisms. Type I alveolar epithelial cell death occurs and is followed by inflammation during the first 7 days, which mimics acute lung injury. The inflammatory response period continues for 14 days. The inflammation then resolves, and fibrosis occurs within 3 days. After 3-4 weeks of BLC administration, the fibrotic stage begins with intense deposition of ECM [26]. Organisms maintain a balance between free radicals and antioxidants to prevent oxidative stress [27]. Imbalances in this mechanism result in exposure to free radicals with an inadequate defense mechanism, and may lead to pulmonary fibrosis [28]. TBARS, which forms due to fatty acid peroxidation with reactive oxygen radicals, reflects lipid peroxidation in the cell. Lipid oxidation can result in irreversible cell membrane damage [29]. The increased TBARS levels in BLC-treated rats demonstrate that lipid peroxidation occurs in this study. Interestingly, TBARS levels were significantly decreased in the HP+BLC group compared with the BLC group, likely due to the radical scavenging and antioxidant properties of HP. SOD, GPx, and CAT are the main defense enzymes and GSH is the main non-enzymatic defense against lipid peroxidation [10,30]. SOD forms H$_2$O$_2$ from superoxide radical anions. The H$_2$O$_2$ is then reduced to water by CAT and GPx [31]. Increased antioxidant enzyme levels were found in the HP-treated groups in this study and in several previous studies [14,32,33]. In this study, BLC-treated rats had significantly decreased SOD, GPx, and CAT activities compared with the control group. The levels of these antioxidant enzymes were significantly increased in the groups treated with HP. These findings suggest that HP promotes antioxidant activity. GSH is a major thiol tripeptide that plays an important role in the antioxidant mechanisms in the cell [34]. GSH is a cofactor for enzymes that prevent oxidative stress, transport amino acids from the basal membrane, scavenge free radicals, and regenerate vitamins C and E to their active forms [34]. Previous studies showed that decreased GSH levels by BLC is a risk factor for lung fibrosis, and that antioxidants can alter the GSH levels [7,11]. In this study, HP treatment successfully reversed GSH depletion in BLC-treated animals. In addition, similar results have been found when using HP to treat cisplatin-induced ocular toxicity and in a cerebral ischemia/reperfusion model [13,14]. This study demonstrated that BLC treatment resulted in significant histopathological alterations in the lung, including thickening of interalveolar septa through macrophage and lymphocyte infiltration, and fibroblast proliferation. HP treatment markedly attenuated these effects in BLC-treated animals. These findings are in agreement with previous studies that used antioxidant agents to treat BLC-induced lung injury [11,34]. Furthermore, this study showed that lung injury could be evaluated using micro-CT with high accuracy compared with histopathological and biochemical techniques. Although there was no significant difference between the micro-CT image analysis of the HP and control groups, there was a difference between the BLC and BLC+HP groups. While the BLC group showed significant lung injury, a considerable decrease in injury area was observed in the BLC+HP group. These results may eliminate the need for redundant biopsies in clinical trials for IPF treatment. A parallel study [35,36] showed that micro-CT was a useful method in determining lung injury, and was as accurate as histopathological techniques. However, more study is needed.

Conclusion

In conclusion, HP successfully minimized the severity of BLC-induced lung injury, which was used as a model for IPF. The positive effects of HP were evaluated through histopathological, biochemical, and micro-CT examinations. The minimizing effect of HP in pulmonary fibrosis would be a promising report for the treatment of IPF for further investigations.

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References

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