Role of YAP-siRNA in reverse sensitivity of human lung cancer A549/TAX cells to taxol and its possible mechanism.

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Abstract

Lung cancer is a most common malignancy in both men and women. Studies have indicated that YAP gene was closely related to chemo-resistance of cancers. However, whether YAP expression participates in the resistance of human lung cancer A549 cells to Taxol (TAX) remains unclear. The aim of this study is to investigate the role of YAP in the resistance of A549 cells to TAX. The sensitivity of cells to TAX was firstly confirmed by MTT assay. The expression levels of YAP in A549 cells and A549/TAX cells were examined by QRT-PCR and Western blot. Small interference RNAs (siRNAs) were used to knock down the gene expression of YAP in A549/TAX cells. Then the cell viability and apoptosis were assessed following treatment with taxol using MTT assay and flow cytometry. The levels of drug resistance related genes, MDR1, MRP1 and BCRP, were examined in YAP-siRNA A549/TAX cells. Furthermore, Western blot analysis was utilized to determine ERK pathway following YAP knockdown. The results demonstrated that the expression of YAP was significantly increased in A549/TAX cells. YAP gene knockdown significantly decreased the IC50 value of A549/TAX cells to TAX and increased taxol-induced apoptosis in the A549/TAX cells. The expression of MDR1, MRP1 and BCRP significantly decreased, and YAP knockdown markedly decreased the activation of ERK pathway. In conclusion, YAP knockdown could reverse the resistance of A549/TAX cells to taxol, thus it may act as a candidate for therapeutic target of lung cancer.

Keywords: YAP, Lung cancer, Chemo-resistance, Taxol, MEK/ERK pathway.

Introduction

Lung cancer is the most common cancer in the world with more than 1 million cases diagnosed every year, and remains the second leading cause of cancer death in both men and women [1]. Advanced Non-Small Cell Lung Carcinoma (NSCLC) accounts for >80% of lung cancer cases [2]. More than half of these patients have developed metastasis at the time of diagnosis and chemotherapy remains the most effective treatment option [3]. Platinum-based doublets are considered to be the standard treatments for NSCLC [4]. The clinical effectiveness of the chemotherapeutic agents used in the treatment of NSCLC is often negated by drug resistance [5,6].

Yes-Associated Protein (YAP) was first discovered and named in 1994, it is a multifunctional intra-cellular junction protein and transcriptional co-activator. YAP plays important roles in regulating signal transduction and gene transcription in normal cells. Studies have confirmed that YAP is the target molecules in Hippo pathway which is a newly discovered cell signal transduction pathway and plays a critical role in regulating the volume and size of organs [7,8]. Researches have shown that YAP was over-expressed in a variety of cancers including liver cancer, gastric cancer, ovarian cancer, prostate cancer, lung cancer, etc., and it was related to the differentiation, proliferation, metastasis, and chemotherapy sensitivity, indicating that YAP is involved in the development of tumor [9-13]. Recent studies indicated that YAP could confer the resistance of cancers to chemotherapy drugs [14], and high expression of YAP was associated with poor patient prognosis [15].

Taxol is an important anti-tumor agent that is particularly effective in the treatment of ovarian and breast cancers, although its usefulness is often hampered by the development of drug resistance. As a new anti-tubulin drug [16], taxol can enhance the binding ability of 14-3-3 protein with YAP without changes of tublins stability and YAP phosphorylation status [17]. This limits the function of YAP as a transcription factor. However, the suppression of taxol on YAP function can be reversed by YAP up-regulation [18].

Based on previous reports, we hypothesized that YAP may be aberrantly expressed in lung cancer cells with chemo-resistant ability and regulation of YAP may affect the chemo-sensitivity of lung cancer cells to chemotherapeutic agents. In this study, we firstly evaluated the YAP expression in A549 (normal lung cancer cells) and A549/TAX (lung cancer cells resistant to
taxol). Then YAP-siRNA was transfected into A549/TAX cells to decrease the expression of YAP. The cellular sensitivity to taxol was measured by MTT assay and cell apoptosis was examined by flow cytometry. The underlying molecular mechanism was revealed through QRT-PCR and Western blot.

**Materials and Methods**

**Cells and cell culture**
The human non-small cell lung cancer (NSCLC) A549 cell line and the taxol-resistant A549/TAX cell line were purchased from American Type Culture Collection (Manassas, VA, USA) and were cultured in RPMI-1640 medium (Gibco, Gaithersburg, MD, USA) supplemented with 10% Fetal Bovine Serum (FBS) (Cor., New York, NY, USA), 100 mg/ml streptomycin and 100 U/ml penicillin, and cells were incubated in a humidified atmosphere with 5% CO$_2$ at 37°C. Cells were passaged every 2-3 d.

**Cell transfection**
For cell transfection, Lipofectamine2000 transfection reagent (Invitrogen) was used. Negative control-siRNA or YAP-siRNA (Santa cruz, CA, USA) was transiently transfected into A549/TAX cells using 20 μl transfection reagent according to the manufacturer’s instructions. Cells were incubated for 48 h before the following tests.

**MTT cell viability assay**
To determine the drug resistance of taxol to A549 cells, half-maximal inhibitory concentration (IC$_{50}$) values were calculated. Cells were seeded into a 96-well plate and cultured in RPMI-1640 media for 24 h, then the cells were treated with various concentrations of taxol (Sigma, Santa Cruz, CA, USA) for 48 h. Subsequently, MTT solution (5 mg/ml) (Eli Lilly, Co. Ltd., Indianapolis, IN, USA) was added to culture medium and incubated at 37°C for 4 h. The Optical Absorbance (OD) at 450 nm was measured using a SynergyTM 2 Multi-function Microplate Reader (Bio-Tek Instruments, Winooski, VT, USA). Each experiment was independently repeated three times.

**Flow cytometry assay**
Annexin V/propidium iodide double staining method was performed to examine cell apoptosis. 48 h after cell transfection, cells were treated with 5 μl TAX for 24 h. Then cells were labeled with annexin V-FITC and Propidium Iodide (PI) for cell apoptotic rate detection following the manufacturer’s instructions, and flow cytometry (Becton Dickinson, New Jersey, USA) was performed. Each experiment was independently repeated three times.

**Western blot analysis**
Total cellular protein was extracted using RIPA buffer containing 1 mM Phenylmethanesulfonyl Fluoride (PMSF), and equal protein samples were standardized using spectrophotometer. Samples were resolved by using SDS-PAGE analysis and transferred to Polyvinylidene Fluoride (PVDF) membranes (Millipore). The membranes were blocked at room temperature for 1.5 h in PBS solution containing 0.1% Tween-20 and 5% fat-free milk, and incubated overnight with primary antibody (1:1000) at 4°C followed by secondary antibody (1:5000) at room temperature for 2 h. The TS380-H automatic washing machine (Taixing TaiSheng Medical Equipment Factory, China) was used to analyse the protein bands.

**QRT-PCR analysis**
After certain treatment, qRT-PCR was performed for gene mRNA expression level detection. GAPDH acted as an internal control. Total RNA was obtained from cells using TRIZOL reagent (Takara, Japan) in accordance with the manufacturer’s instructions. Then, RNA was transcribed into cDNA using RT-PCR detection kit (Invitrogen, Carlsbad, CA, USA) according to manufacturer’s instructions. Real-time PCR was performed to analyse the synthesized cDNA and the conditions were 95°C for 10 min, followed by 40 cycles of 95°C for 10 s, 60°C for 60 s. All the primers were performed by Sangon Biotech (Shanghai, China). The primer sequences of real-time PCR are as shown in Table 1.

**Statistical analysis**
All quantitative data were displayed as the mean ± Standard Deviation (SD). Student’s t-test was used to identify significant differences in multiple comparisons. For all statistical analyses, P<0.05 was considered statistically significant.

**Results**

**Yap is upregulated in taxol resistant lung cancer cells**
To confirm the taxol resistant ability of A549/TAX cell line, which was obtained from ATCC, MTT assay were firstly performed on A549 and A549/TAX cell lines after treated with taxol. As shown in Figure 1A, a significant inhibiting effect of taxol on cell viability was observed in A549 cell line, thus suggesting that A549 cells were more sensitive to taxol. That is, the obtained A549/TAX cell line was resistant to taxol as expect. To investigate the biological role of YAP in the taxol resistance of non-small cell lung carcinoma, we compared the expression level of YAP in A549 and A549/TAX. The results were shown in Figures 1B and 1C. Compared with A549 cells,
the YAP expression was significantly upregulated in A549/TAX cells, indicating a possible involvement of YAP in drug resistance in lung cancer, especially NSCLC.

**Decrease of YAP mRNA and protein expression after YAP-siRNA**

To further investigate the role of YAP in A549/TAX cells, YAP gene was knocked down using siRNA technology. The YAP mRNA and protein expression was examined by qRT-PCR and Western blot after siRNA transfection. As shown in Figure 2, the mRNA and protein expression levels were decreased to a relatively low level after transfection. It was considered that YAP gene expression was efficiently suppressed and siRNA transfection could be used as an effective method to inhibit the expression of YAP.

**Increased sensitivity of lung cancer cells to taxol after YAP-siRNA transfection**

MTT assay was utilized to determine the change of sensitivity to taxol after silence of YAP expression in A549/TAX cells. As shown in Figure 2C, a significant inhibiting effect of taxol on cell viability was observed in YAP-siRNA transfection group, indicating that YAP knockdown significantly increased the sensitivity of A549/TAX cells to taxol.

**Increase of apoptosis in YAP-knockdown A549/TAX cells**

The effects of YAP knockdown on cell apoptosis in the A549/TAX cells were evaluated by flow cytometric analysis. The results were exhibited in Figure 3. After treated with taxol, the apoptosis rates were 12.16% in the parental A549/TAX cells and 13.69% in the A549/TAX cells transfected with control siRNA (P>0.05), whereas the apoptotic cells were 28.51% in A549/TAX YAP-siRNA cells (P<0.05). Compared with parental A549/TAX cells, the number of apoptotic cells was markedly increased in the YAP-knockdown A549/TAX cells. The results demonstrated that A549/TAX cells with downregulation of YAP were more sensitive to taxol compared with the control group.

In order to investigate the mechanisms of sensitization induced by taxol in the A549/TAX cells, the expression levels of apoptosis-associated proteins were investigated following taxol treatment. As shown in Figure 4, YAP knockdown significantly decreased the expression level of Bcl-2 and survivin in A549/TAX cells, while the expression level of cleaved-caspase-3 was increased remarkably. In short, these results indicated that YAP is an inducer of taxol resistance and involves in regulating cell apoptosis.

**YAP knockdown affects the expression of drug resistance associated genes in A549/TAX cells**

To explore the underlying molecular mechanisms of the roles that YAP played in taxol resistance, the expression of drug resistance related genes, *MDR1*, *MRP1* and *BCRP* was examined in A549/TAX cells. As illustrated in Figure 5, both the mRNA and protein levels of MDR1, MRPI and BCRP were decreased in A549/TAX YAP-siRNA cells. These
findings suggested that YAP may induce taxol resistance through regulating drug transportation.

Figure 3. Increase of apoptosis in YAP-knockdown A549/TAX cells. (A) Cell apoptosis detected by flow cytometry; (B) the cell apoptosis rate calculated by adding early apoptosis rate and late apoptosis rate up. Con: Control; NC: Negative Control; **p<0.01. Each experiment was independently repeated three times.

Figure 4. Effect of taxol treatment on the level of apoptosis-associated proteins in A549/TAX cells after transfection. (A) The protein expression level of Bcl-2, Survivin, cleaved-caspase3 were assayed by Western blot; (B) Densitometric analysis of each protein level. Con: Control; NC: Negative Control; **p<0.01. Each experiment was independently repeated three times.

YAP knockdown inhibits the MEK/ERK pathway in A549/TAX cells

Aberrant activation of the ERK pathway has been reported in various carcinomas and it plays a critical role in carcinogenesis by regulating cell proliferation, invasion and migration [19-21]. To further elucidate the mechanism involved in the effect of YAP on taxol resistance in lung cancer, MEK/ERK pathway was explored further. By Western blot analysis, we found that activation (phosphorylation) of MEK and ERK was reduced in YAP-siRNA group (Figure 6). That is, YAP knockdown inhibits the MEK/ERK pathway in A549/TAX cells. The results indicated that MEK/ERK pathway participated in the effect of YAP on taxol resistance in lung cancer.

Figure 5. YAP knockdown affects the expression of drug resistance associated genes in A549/TAX cells. (A) The mRNA expression level of MDR1, MRP1 and BCRP; (B) the protein expression level of MDR1, MRP1 and BCRP; (C) Densitometric analysis of each protein level. Con: Control; NC: Negative Control; **p<0.01. Each experiment was independently repeated three times.

Figure 6. YAP knockdown inhibits the MEK/ERK pathway in A549/TAX cells. (A) The protein expression level of p-MEK1/2, MEK1/2, p-ERK1/2, ERK1/2 were assayed by Western blot; (B) Densitometric analysis of each protein level. Con: Control; NC: Negative Control; **p<0.01. Each experiment was independently repeated three times.

Discussion

Drug resistance is the major factor that affects the efficacy of chemotherapy in patients with lung cancer. There are many
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Factors that affect the resistance of NSCLC drug resistance, including the increases of the expression of resistant multi-drug (MDR) gene, multidrug resistance associated protein (MRP) genes and their product, as well as the decreased expression of Topoisomerase II (Topo II) and Glutathione (GSH) and the increased expression of Glutathione S Transferase (GST) system. In addition, studies have shown that the abnormal expression of related factors in cell signal transduction, the abnormal of DNA repair in tumor cells and the aberrant expression of other related genes are closely related to lung cancer drug resistance [22]. Therefore, exploring novel therapeutic targets and effective molecular markers to predict the sensitivity of patients to drugs is extremely critical for NSCLC treatment.

YAP, a critical effector of Hippo signaling pathway, is one of the major transcription activation factors and is highly active. Studies have suggested that YAP is highly expressed in both nuclear and cytoplasm of lung cancer cells and involved in the development of lung cancer [23]. Recently, a growing number of evidence indicated that the expression of YAP may associate with multi-drug resistance [24]. However, the relationship of YAP gene expression and Taxol chemoresistance in human lung cancer cells is still unclear. In the present study, the expression of YAP was significantly elevated in A549/TAX cell line both in protein and gene levels, thus indicating a possible involvement of YAP in drug resistance in lung cancer.

As an effective antitumor drug, taxol is an effective therapeutic agent to various tumor cells and it can limit the function of YAP as a transcription factor [25]. In order to precisely and definitely elucidate the role of YAP on the resistance of taxol in lung cancer, we suppressed YAP gene by siRNA in A549/TAX cells and then examined the chemo-sensitivity to taxol of A549/TAX cells. We found that YAP knockdown significantly increased the sensitivity of A549/TAX cells to taxol. Intrinsic or acquired drug resistance limits the effectiveness of chemotherapy. One of the most studied mechanisms of drug resistance in cancer cells is associated with altered anticaner drug transport, mediated by members of the ATP-Binding Cassette (ABC) superfamily of transport protein such as Multidrug Resistance Protein-1 (MRP1) and Breast Cancer Resistance Protein (BCRP) [26-28]. This resistant type has been termed Multidrug Resistance (MDR) [29]. Multi Drug Resistant Protein 1 (MDR1) has been linked to multi drug resistance and disease progression in cancers such as NSCLC [30]. Overexpression of MRP1 has been detected in a variety of tumor types, and has also been associated with drug resistance or poor patient outcome in lung cancer [31]. BCRP, which is a novel ABC transport protein, functions as an energy-dependent efflux pump. Elevated expression of BCRP in vitro causes resistance against at least two classes of anticancer drugs, anthracyclines and topoisomerase I inhibitors [32]. In the present study, we found that both the mRNA and protein levels of MDR1, MRP1 and BCRP were decreased in A549/TAX YAP-siRNA cells, thus suggesting YAP may induce taxol resistance through regulating drug transportation.


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Role of YAP-siRNA in reverse sensitivity of human lung cancer A549/TAX cells to taxol and its possible mechanism

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