Quantitative determination of resveratrol in *Polygonum cuspidatum* and its anti-proliferative effect on melanoma A375 cells.

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Abstract

Objective To establish HPLC method for determination of resveratrol content in Polygonum cuspidatum, and to study its inhibitory effect on human melanoma A375 cells. Methods HPLC with an Agilent column (model Angilent ZORBAX Eclips Plus C18, 4.6 × 250 mm, 5 µm) is used; mobile phase: methanol-water (75:25); flow rate: 1.0 mL/min; UV detection wavelength: 300 nm; injection volume: 5 µL; and column temperature: 25 °C. Anti-melanoma activity of resveratrol is analyzed by observing changes in cell morphology under inverted microscope and by MTT assay. Results Resveratrol shows a good linearity within a 1.248-12.48 µg range, and its recovery is 98. 58%, with a RSD of 1.62% (n = 6). MTT assay results show that the viability of A375 cells decreases with increasing concentration of resveratrol; and A375 cell inhibition rate of resveratrol exhibits rather evident dose-response relationship within the experimental concentration range. At high magnification, cells in the control group have clear nuclear membranes; a large number of mitochondria are visible within cytoplasm; double membranes and ridge structure are all clear. A small number of rough endoplasmic reticula. free ribosomes and lysosomes are seen, and cell structure is intact. Cells treated with resveratrol change rather greatly, with swollen organelles, increased number of intracellular granules, and even cell membrane disintegration, liquefaction degeneration cytoplasm, substantial disappearance of mitochondria and endoplasmic reticulum structure, release of cellular contents and occurrence of apoptosis. Cell morphology is not intact, and number of adherent cells is reduced. Conclusion HPLC method is accurate, simple and reproducible, which is suitable for quantitative analysis of resveratrol in Polygonum cuspidatum. Resveratrol can effectively inhibit the proliferation of human melanoma A375 cells.

Keywords: Polygonum cuspidatum; resveratrol; melanoma A375 cell

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Introduction

Polygonum cuspidatum is the dried root and rhizome of *Polygonum cuspidatum* Sieb. et Zucc. in the genus *Polygonum* of the family Polygonaceae. As a perennial shrubby herb, it is mainly distributed in China's various regions south of the Yangtze River, as well as Shaanxi, Hubei and Sichuan. Polygonum cuspidatum is used for joint arthralgia, amenorrhea, water and fire burns, bruises, carbuncles, sores, cough, excessive phlegm, etc. [1]

Polygonum cuspidatum mainly contains anthraquinones & anthraquinone glycosides, stilbenes, flavonoids, phenolic compounds, etc [2-4], of which monomer

components like polydatin and physcion have been widely used in medicine industry. Polygonum cuspidatum has become an important source of raw material for these monomer components. Modern pharmacological studies have shown that Polygonum cuspidatum has antibacterial, anti-inflammatory, anti-viral [5-6], vascular smooth muscle dilating, microcirculation improving, antiplatelet aggregation, hypolipidemic, anti-tumor, antioxidation, hepatoprotective and choleretic effects [7-9]. Particularly in recent years, American Institute of Natural Medicine has reported that resveratrol in Polygonum cuspidatum has an anti-HIV effect, which has further promoted the R & D of Polygonum cuspidatum. In order to further research and develop Polygonum cuspidatum, this paper studies the method for quantitative determination of resveratrol, a signature component of Polygonum cuspidatum, and explores its anti-melanoma activity, in order to provide evidence for its clinical application.

Materials

Instruments and reagents

Angilent 1100 HPLC system; Angilent ZORBAX Eclips Plus C18 column. Reagents used for preparation of standard and test solutions were all of analytical grade, and those used for liquid chromatography were HPLC grade reagents; water used for liquid chromatography was ultrapure water. DMSO and MTT were purchased from Sigma, USA; FBS was purchased from Gibco; and RPMI1640 from Invitrogen.

Drugs and cells

Polygonum cuspidatum medicinal herb was purchased from the medicine market in Anguo, which was identified as Polygonum cuspidatum. Resveratrol reference substance was self-prepared, whose purity was determined to be 99.5% by area normalization. Melanoma

A375 cells were purchased from Shanghai ELISA Kit Assay Center.

Methods

Chromatographic conditions

Agilent column (model Angilent ZORBAX Eclips Plus C18, 4.6×250 mm, 5 µm); mobile phase: methanol-water (75:25 by volume); flow rate: 1.0 mL/min; UV detection wavelength: 300 nm; injection volume: 5 µL; and column temperature: 25 °C. Under such chromatographic conditions, resveratrol was eluted, and basically achieved baseline separation; retention time was 13.104 min.

Linearity range

1 mL, 2 mL, 4 mL, 6 mL, 8 mL and 10 mL of 12.48 μ g/mL resveratrol reference solutions were precisely drawn, respectively, placed into 10 mL volumetric flasks, diluted to the mark with methanol, and injected at 10 μ L separately to measure the peak area of resveratrol under the above chromatographic conditions. Standard curve was plotted with mass concentration (X) versus peak area (Y), regression equation was Y = -18567.73X + 15622.52, r = 0.9997, and linear range: 1.248-12.48 μ g/mL.

Stability test

Each 10 μ L of the same test solution was precisely drawn, and injected at 0, 2, 4, 6, 8, 12 and 24 h, respectively, for determination according to the above chromatographic conditions, and RSD was found to be 0.85%. The results showed that the test solution was stable within 24 h.



Figure 1. HPLC chromatogram of resveratrol.



Figure 2. HPLC chromatogram of Polygonum cuspidatum sample

Precision test

Each 10 μ L of the same test solution was precisely drawn, and injected repeatedly 5 times for determination according to the above chromatographic conditions. RSD was found to be 1.2% (n = 5).

Recovery test

Six aliquots of 0.5 g of Polygonum cuspidatum coarse powders of known contents were accurately weighed, added separately with 1 ml of 3.18 µg/mL resveratrol reference solution, prepared as per the above preparation method of test solution, and determined according to the above chromatographic conditions. The results showed that the average recovery of resveratrol was 98. 58%, with a RSD of 1.62% (n = 6).

Reproducibility test

Five aliquots of samples of the same batch were accurately weighed, and prepared as per the preparation method of test solution. Then, 10 μ L of the above solution was precisely drawn, and injected for determination under the same chromatographic conditions as described above. The results showed that RSD was 1.5% (n = 5).

Sample determination

0.5 g of five different batches of Polygonum cuspidatum coarse powders were accurately weighed, and ultrasonically extracted for 1 h with 20 mL of 80% aqueous methanol, then diluted to the mark, and filtered. 1 mL of subsequent filtrates were placed into 10 mL volumetric flasks, diluted to the mark with mobile phase, and ultra-filtered. Afterwards, 10 μ L of subsequent filtrates were injected for determination of resveratrol content according to the above method and chromatographic conditions. The results showed rather big difference in resveratrol content of five different batches of samples, with an average of 0.109%. See Table 1 below.

Cell culturing

A375 cells were cultured routinely in a 37° C, 50 mL/L CO₂ incubator with RPMI 1640 medium containing 100 mL/L FBS, 100 ku/L penicillin and 100 mg/L treptomycin. Medium was replaced and cells were passaged every 2-3 days.

A375 cell viability assay by MTT

 2×10^6 /ml A375 cells were seeded in 96-well flatbottomed plates, divided into five groups, and added with 20, 40 and 80 µmol/L resveratrol for treatment. Six replicate wells were set up for each group. Cells were cultured routinely, and assayed at 48 h and 72 h, respectively. Each well was added with 20 µl of MTT (5 mg/ml), and incubated for 4 h. Before assay, supernatant was discarded, and the crystal was dissolved in 100 µl of DMSO, shaken on a micro oscillator for 5 min, and measured for absorbance at 570 nm on a microplate reader, followed by calculation of cell growth inhibition rate.

Calculation formula for cell inhibition rate: inhibition rate (%) = (OD value of control group - OD value of experimental group) / OD value of control group \times 100%.

Experimental results showed that after culturing for 48 h and 72 h, A375 cell growth was all inhibited to varying degrees, exhibiting evident concentration- and time-dependence. Compared with the control group, different concentrations of resveratrol all had significantly different inhibition rates on A375 cell growth.

Table 1. Determination of resveratrol content in Polygonum cuspidatum from different habitats

	Sample	Sample size (g)	Content (%)	Average content (%)
1		5.012	0.086	
2		4.998	0.121	
3		5.023	0.094	0.109
4		5.108	0.113	
5		5.088	0.132	

Table 2. Inhibitory effect of resveratrol on growth of A375 cells

Group	Concentration (mg/ml)	Inhibition rate (%, 24 h)	Inhibition rate (%, 48 h)
Control group	0	0	0
Experimental group	20µmol/L	42.34±5.43	66.87±3.62
Experimental group	40µmol/L	45.76±3.89	74.32±2.57
Experimental group	80µmol/L	58.97±4.53	76.86±3.51



TEM observation of cell morphology

The above concentrations of resveratrol solutions were added into cell culture flasks. After treating for 48 h, cells were collected, and washed. Then, the cells were fixed in 1% osmic acid, wash with 0.1% PBS, dehydrated routinely, impregnated, embedded in epoxy resin, and cut into ultra-thin sections for observation of cell morphology and photography under TEM.

Under electron microscope, cells in the control group had regular nuclear morphology, which were generally elliptical or oval. Nuclear membranes were clear, and a large number of mitochondria were visible within cytoplasm. Double membranes and ridge structure were all clear. After treating with resveratrol for 48 h, part of A375 cells were swollen, and bulges were formed in cell membranes. Swelling and chromatin margination were found in mitochondria, endoplasmic reticula and nuclei. A very small number of cells presented typical early apoptotic changes. In high concentration group, complete disruption of nuclear membranes, structure collapse, karyolysis and cell content overflow, i.e. oncosis-like changes, were observed.

Discussion

Melanoma was first proposed by Rene Laenneic (Jay V.2000) in 1804 [10]. There is only one type of malignant cells that constitute all malignant melanomas, which are malignant pigment-producing cells called melanocytes. Yet they have some small differences in terms of morphology, such as spindle ones and cylindrical ones.

Melanoma is a highly malignant tumor, which originates from the melanocytes in pigmentation areas of skin, mucosa, eyes and central nervous system. Its clinical symptoms include bleeding, itching, tenderness, ulcers, etc. In general, symptoms of melanoma are age-related; younger patients generally manifest itching, and increased colorific change and area of skin lesions, while elderly patients generally manifest ulcers of skin lesions, which usually indicates poor prognosis [11-12].

Skin lesions of cutaneous malignant melanoma are associated with anatomical location and growth pattern of tumors, that is, associated with histological type. Histological type varies greatly due to age, gender and race differences. Different types of melanomas have different causes and genetic backgrounds. At present, Clark's level is used for clinical histological classification of melanomas, which include four types: lentigo maligna melanoma (LMM); superficial spreading melanoma (SSM); acral lentiginous melanoma/mucosal melanoma; and nodular melanoma (NM). Among malignant melanomas suffered by Caucasians, about 70% are SSM.

When malignant melanomas spread, they usually do not produce melanin, so they are called non-pigmented melanomas, but non-pigmented and pigmented malignant melanomas have almost the same malignant degree. Malignant melanomas occur most commonly in the skin, but 10% occur in the eye. Most common types of cutaneous malignant melanomas are called superficial spreading malignant melanoma and small nodular malignant melanoma.

Pathogenesis of melanoma is generally believed to be the malignant transformation of melanocytic nevus caused by repeated friction, grab and injury; improper excavation or drug corrosion may convert benign melanocytic nevus into malignant melanoma. This study shows that different concentrations of resveratrol can inhibit the growth of A375 cells to varying degrees in a concentration- and time-dependent manner. Under electron microscope, cells in the control group have regular nuclear morphology, which are generally elliptical or oval. Nuclear membranes are clear, and a large number of mitochondria are visible within cytoplasm. Double membranes and ridge structure are all clear. After treating with resveratrol for 48 h, part

of A375 cells are swollen, and bulges are formed in cell membranes. Swelling and chromatin margination are found in mitochondria, endoplasmic reticula and nuclei. A very small number of cells present typical early apoptotic changes. In high concentration group, complete disruption of nuclear membranes, structure collapse, karyolysis and cell content overflow, i.e. oncosis-like changes, are observed. The specific mechanisms of action of resveratrol need further confirmation.

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