Mycobacterial secretory SEVA TB ES-31 antigen, a chymotrypsin-like serine protease with lipase activity and drug target potential.

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Abstract

Mycobacterium tuberculosis is known to secrete number of proteins which play an important role in pathogenicity and diagnosis. A diagnostically important secreted antigen, Excretory Secretory-31 (SEVA TB ES-31) protein with serine protease activity was isolated from *Mycobacterium tuberculosis* H₃₇Ra culture filtrate. Serine proteases like Chymotrypsin have been reported to possess lipase activity as the catalytic triad is similar in serine protease and some lipase enzymes. In this study, ES-31 showed presence of 44.5U/mg pr of lipase activity by titrimetric assay. The lipase activity was inhibited by lipase inhibitor, Orlistat by 100%, as well as by serine metalloprotease inhibitors, Phenyl methyl sulphonyl fluoride and Ethylene Diamine Tetraacetic acid by 100%. The serine protease activity of ES-31 was inhibited by Phenyl methyl sulphonyl fluoride, Ethylene Diamine Tetraacetic acid, and orlistat by 78.3%, 64.3%, 89.4% respectively. Inhibition of serine protease activity of ES-31 by lipase inhibitor of lipase activity of ES-31 by serine protease inhibitor suggests that the catalytic site of ES-31 is sensitive to both types of the inhibitors and ES-31 may be a chymotrypsin-like protein with drug target potential.

Keywords: Mycobacterium tuberculosis H₃₇Ra, Mycobacterial secreted serine protease, Lipase, Orlistat.

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Introduction

Mycobacterium tuberculosis (M.tb.) is an intracellular pathogen, living in the macrophages of host and persists there for a long time. In general, exported proteases in bacteria are associated with bacterial pathogenic virulence, however little information is available on *M*. tb secreted proteins [1]. In our laboratory, SEVA TB Excretory Secretory-31 (SEVA TB ES-31), a 31kDa secreted antigen was isolated from culture filtrate of M.tb. H₃₇Ra previously characterized as a serine metallo protease. The proteolytic activity of ES-31 was inhibited by different inhibitors [2,3]. Some lipases share a common Ser-His-Asp/Glu catalytic triad with serine proteases like trypsin, chymotrypsin and subtilisin in their active sites [4,5]. Some of the serine proteases like chymotrypsin and subtilisin show lipase activity however not all lipases tested show serine protease activity except commercially available porcine pancreatic lipase (PPL) [5,6].

The present study was carried out to find out the presence of lipase activity in ES-31, as serine protease activity is already determined. Hydrolysis of short chain (water soluble) esters serves the estimation of esterase activity and that of long chain fatty acids provide the measure of lipase activity [7]. Therefore olive oil, a substrate of lipase was emulsified in water for the determination of lipase activity by titration. In this study, inhibition of lipase and serine protease activities of ES-31 was assessed using acid-base titration and Azocasein assay respectively.

Materials and Methods

Isolation of tubercular ES-31 serine protease

ES-31 antigen was isolated from *M.tb.* H_{37} Ra ES antigens by affinity chromatography using anti ES-31 antibody coupled Sepharose-4B column (Pharmacia Biotechnology AB, Uppsala, Sweden) [8]. Briefly, Cyanogen bromideactivated Sepharose 4B beads were coupled with purified anti ES-31 antibody. DSS antigen was passed through column and ES-31 antigen was eluted by glycine HCl buffer (0.01 mol/l, pH2.5) and neutralized with Tris-HCl buffer (0.01M, pH8.6).

Titrimetric Assay

Lipase activity of SEVATB ES-31 antigen was studied at pH 8.5 by titrimetric method using olive oil as a substrate at pH 8.5 at 37^{0} C [9]. In brief, an emulsion of substrate containing 2% olive oil with 8mg/ml gum Arabic in 26.7mM Tris-Cl buffer was prepared by sonication. 6.5 ml of substrate emulsion and 1ml of enzyme was incu-

bated for 30 min at 37[°]C. Few drops of phenolphthalein indicator solution were added to incubation mixture and titrated till the appearance of light pink color with 5mM NaOH solution. The lipase activity was determined in terms of the amount of NaOH required for the titration. The amount of NaOH required for color change was noted for three different concentrations of ES-31. When Triacylglycerol lipase hydrolyses olive oil, fatty acids are liberated in the substrate solution (pH 8.5) and pH of solution decreases. Phenolphthalein turns pink from colorless on the addition of NaOH at pH 8.3. To analyze inhibition of lipase activity if any by serine metallo protease inhibitors like Phenyl methyl sulphonyl fluoride (PMSF) (1mM) and Ethylene Diamine Tetraacetic acid (EDTA) (1mM) was checked by titrimetric assay. To find inhibitory effect, ES-31 antigen was preincubated at 37°C for 1 hour with PMSF (1mM), EDTA (1mM) and Orlistat (1mM)(Commercial Name- Lipocut 60, Manufacturer-Lupin India).

Azocasein Assay

To confirm whether the active site of ES-31 as serine protease and as lipase was the same; inhibition of serine protease activity by lipase inhibitor like Orlistat was checked by azocasein assay. In brief, 5 ml of azocasein assay incubation mixture consists of the mycobacterial ES-31 antigen (100 μ g) with 25 mg azocasein in 0.5M Sodium Bicarbonate buffer (ph 8.3). The azocasein assay mixture was incubated at 37^oC for 6 hours. Further, 1ml aliquot solution was removed and 4ml of trichloroacetic acid (5%) was added to the solution. After mixing and filtration using 0.45 μ m syringe filters, again 1ml aliquot was removed and 3ml of 500mM NaOH solution was added to the solution. Absorbance of the liberated dye at 440nm was measured using a spectrophotometer (Ultra-spec, Elico Ltd, India) [3]. To find inhibitory effect if any, 100 μ g of ES-31 was preincubated at 37⁰C for 1 hr with lipase inhibitor Orlistat (0.5mM, 1mM and 1.5mM), serine metallo protease inhibitors, phenyl methyl sulphonyl fluoride (PMSF) (1mM) and Ethylene diamine tetraacetic acid (EDTA) (1mM) and assayed for protease activity.

Results

SEVA TB ES-31, a serine protease has shown 44.5U/mg pr of lipase activity by titration. The lipase activity was found to increase in an enzyme concentration dependant manner and there exist a direct correlation between enzyme activity with enzyme concentration (Table 1).

Table 1. Assay of Lipase activity of SEVA TB ES-31 Ag.

ES-31 Ag	Volume of NaOH re- quired for Titration	Lipase Activ- ity (Units) [*]	
0.25mg	2.1ml	10.5	
0.5mg	4.6ml	23	
1mg	8.9ml	44.45	

*One Unit of lipase activity is the amount of enzyme that catalyzes the reaction of 1µmole of substrate per minute.

The lipase activity of ES-31 was inhibited by serine protease inhibitor PMSF (1mM), metalloprotease inhibitor EDTA (1mM) as well as lipase inhibitor Orlistat by 100%. Serine protease activity of ES-31 was inhibited by 1mM EDTA (64.3%) and 1mM PMSF (78.3%) as well as by 1mM Orlistat (89.4%) (Table 2).

Table 2. Effect of different inhibitors on Serine protease and Lipase activities of SEVA TB ES-31 Ag.

	Lipase ^a		Serine protease ^b	
Inhibitors (Optimum concentration)	Activity (Un- its ^c)	Inhibition (%)	Activity (Un- its ^c)	Inhibition (%)
Control ^d	44.5	0	0.39	0
$PMSF^{e}(1mM)$	0	100	0.08	78.3
EDTA ^f (1mM)	0	100	0.14	64.3
Orlistat ^g (1mM)	0	100	0.04	89.4

^{*a*} *Lipase activity was determined by Titrimetric assay.*

^b Serine protease activity was determined by Azocasein assay.

^c 1 Unit of lipase/ serine protease activity is the amount of enzyme that catalyzes the reaction of 1μ mole of substrate per minute.

^d Control contains 1mg ES-31 Ag in Lipase Assay and 100µg ES-31 Ag in Serine protease Assay and no inhibitor.^e

Serine Protease inhibitor. ^f Metallo enzyme inhibitor,

^g Lipase inhibitor

Discussion

Some lipases use the same catalytic triad as that of serine protease and also hydrolyze ester bond by the same me-48

chanism [4]. The catalytic triad (Ser-His-Asp/Glu) comprises of a Serine residue that has a role in nucleophilic attack on substrate, an acidic amino acid, Aspartate or Mycobacterial secretory SEVA TB ES-31 antigen,

sometimes Glutamate (in case of lipase) and a Histidine acts as a base [4,5,10].

Serine proteases like chymotrypsin and subtilisin have shown lipase activity [5]. ES-31, a serine protease has also shown lipase activity by titrimetric assay. Lipases hydrolyze triacylglycerol when it is aggregated in an oilwater interface [4,11]. Earlier it was studied that PMSF and EDTA inhibit Bacillus coagulance MTCC-6375 lipase [7] and ES-31 also inhibited by PMSF and EDTA [3]. To confirm whether the active site of ES-31 serine protease and ES-31 lipase is the same; its inhibition of serine protease activity by lipase inhibitor like Orlistat was checked by azocasein assay and inhibition of lipase activity by serine metallo protease inhibitors like PMSF and EDTA was checked by titration. Orlistat binds to the active site of pancreatic lipase, which leads to some conformational changes and abolishes its activity [12]. PMSF specifically binds to the serine residue of active site of serine protease and other enzymes having serine in its active site [13], EDTA binds to metal ion required by metalloproteases and inactivates them [13]. Orlistat inhibited the serine protease activity and PMSF and EDTA inhibited lipase activity suggesting that the active site of ES-31 for serine protease and lipase may be same consisting of a typical catalytic triad of Ser-His-Asp. ES-31 may be a chymotrypsin-like enzyme which needs further investigation.

From this study it was found that ES-31 is highly sensitive to PMSF, EDTA, and Orlistat. The inhibition of serine protease activity by Orlistat and inhibition of lipase activity by PMSF and EDTA suggests the presence of same catalytic triad responsible for both serine protease and lipase activity of ES-31 Antigen. Earlier studies have shown that ES-31 (Serine protease) has potential to be a drug target [3]. Present study has shown that ES-31 is highly sensitive to a commercial anti-obesity drug Orlistat for both serine protease and lipase activities and thus orlistat can also be used to inhibit the action of ES-31 Ag. Thus Serine protease and lipase inhibitors or their analogues may act as drugs and inhibit the growth of tubercle bacilli.

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