LANDLOCKED FALL CHINOOK SALMON EGG SIZE IS POSITIVELY RELATED TO HATCHING TIME

Wyatt Krebs, Eric Krebs, Nathan Huysman and Michael E. Barnes*

South Dakota Department of Game, Fish and Parks, McNenny State Fish Hatchery, 19619 Trout Loop, Spearfish, South Dakota, 57783 USA

ABSTRACT

Within fish species, there can be considerable variation in the sizes of eggs produced by individual females. Specifically within the family Salmonidae, the relationship between egg size and hatching time is uncertain. This study examined eggs obtained during the spawning of landlocked fall Chinook salmon from Lake Oahe, South Dakota, USA. Diameters were obtained from the eggs of 24 randomly-selected females spawned in October, 2017, and the number of days to hatch was recorded after incubation at 11°C. Mean egg diameter was 7.22 mm and ranged from 6.36 to 8.36 mm. Over 90% of the eggs survived, with mean hatching occurring from 46 to 49 days of incubation. Significant correlations were observed between egg diameter and time-to-initial-hatch \((r=0.581; p=0.003)\), as well as between egg diameter and mean hatching time \((r=-0.450; p=0.027)\). There was no correlation between egg diameter and survival to hatch \((r=-0.363; p=0.081)\). These results are the first reported relationship between egg size and time-to-hatch in landlocked fall Chinook salmon.

Keywords: Chinook salmon; *Oncorhynchus tshawytscha*; Egg size; Hatch time; Lake Oahe

INTRODUCTION

Compared to other freshwater fishes, trout and salmon have relatively large eggs with long developmental periods (Stickney, 1994). While temperature is the primary determinant of when eggs hatch (Blaxter, 1969; Alderdice and Velsen, 1978; Réalis-Doyelle et al., 2016), other factors may also be involved. For example, low dissolved oxygen levels can influence hatching time (Blaxter, 1969), possibly by activating hatching enzymes (Czerkies et al., 2001). Egg size has also been implicated as having an effect on hatching times (Pauly and Pullin, 1988; Gilliooly et al., 2002). For example, Thorpe et al. (1984) noted that larger Atlantic salmon (*Salmo salar*) eggs took longer to hatch than smaller eggs. Selj et al. (2018) also reported that smaller steelhead trout (*Oncorhynchus mykiss*) eggs hatched sooner than larger eggs. However, no relationship between egg size and hatching time was observed for coho salmon (*O. kisutch*) or chum salmon (*O. keta*) by Beacham et al. (1985) and Beacham and Murray (1985). The lack of effect of egg size on hatching time has also been reported for Chinook salmon (*O. tshawytscha*), Arctic char (*Salvelinus alpinus*), and Atlantic salmon (Kinnison et al., 1998; Einum and Fleming, 2000; Leblanc et al., 2016).

Landlocked fall Chinook salmon from Lake Oahe, South Dakota, USA have relatively large eggs with long developmental periods (Stickney, 1994). While temperature is the primary determinant of when eggs hatch (Blaxter, 1969; Alderdice and Velsen, 1978; Réalis-Doyelle et al., 2016), other factors may also be involved. For example, low dissolved oxygen levels can influence hatching time (Blaxter, 1969), possibly by activating hatching enzymes (Czerkies et al., 2001). Egg size has also been implicated as having an effect on hatching times (Pauly and Pullin, 1988; Gilliooly et al., 2002). For example, Thorpe et al. (1984) noted that larger Atlantic salmon (*Salmo salar*) eggs took longer to hatch than smaller eggs. Selj et al. (2018) also reported that smaller steelhead trout (*Oncorhynchus mykiss*) eggs hatched sooner than larger eggs. However, no relationship between egg size and hatching time was observed for coho salmon (*O. kisutch*) or chum salmon (*O. keta*) by Beacham et al. (1985) and Beacham and Murray (1985). The lack of effect of egg size on hatching time has also been reported for Chinook salmon (*O. tshawytscha*), Arctic char (*Salvelinus alpinus*), and Atlantic salmon (Kinnison et al., 1998; Einum and Fleming, 2000; Leblanc et al., 2016).

Landlocked fall Chinook salmon from Lake Oahe, South Dakota, USA have relatively large eggs with long developmental periods (Stickney, 1994). While temperature is the primary determinant of when eggs hatch (Blaxter, 1969; Alderdice and Velsen, 1978; Réalis-Doyelle et al., 2016), other factors may also be involved. For example, low dissolved oxygen levels can influence hatching time (Blaxter, 1969), possibly by activating hatching enzymes (Czerkies et al., 2001). Egg size has also been implicated as having an effect on hatching time in landlocked fall Chinook salmon. Thus, the objective of this study was to ascertain if hatching time is influenced by egg size in landlocked Chinook salmon from Lake Oahe, South Dakota.

MATERIAL AND METHODS

Spawning

Landlocked fall Chinook salmon from Lake Oahe, South Dakota were spawned on October 16, 2017 at Whitlocks Spawning Station near Gettysburg, South Dakota, USA. Milt was collected from males, pooled in a container, and kept on ice until use. Eggs from 24 females were used in this study. Each individual female was pneumatically spawned using compressed oxygen at low pressure to express the eggs into a mesh net. Lake water was added to the spawn to activate the sperm for fertilization. After 2 min eggs were rinsed with lake water and allowed to water harden in lake water for at least 1 h prior to shipment. This process was repeated for each of the spawns used in this study (24 times).

Egg Incubation

After water hardening, eggs were transported approximately 4 h to McNenny State Fish Hatchery, rural Spearfish, South Dakota, USA. Upon arrival at the hatchery, eggs were disinfected with 100 mg/L buffered free iodine (Syndel, Ferndale, Washington, USA) for 10 min. Eggs were then inventoried using water displacement (Piper et al., 1982) and placed into vertical-flow incubation trays (MariSource, Fife, Washington, USA) with discrete compartments for the spawn from each female. Each spawn was maintained discretely during spawning, fertilization, transportation, and incubation. Well water \((11^\circ C); \text{total hardness } 360 \text{mg/L CaCO}_3; \text{alkalinity as } \text{CaCO}_3, 210 \text{mg/L}; \text{pH } 7.6; \text{total dissolved solids}

390 mg/L) at 12 L/min was used throughout incubation. Daily treatments of 1,667 mg/L formalin (37% formaldehyde, 6 to 14% methanol; Paracide F, Syndel, Ferndale, Washington) for 15 min (Piper et al., 1982) were administered with a Masterflex model 7524-00 microprocessor peristaltic pump (Cole-Parmer Instrument Company, Vernon Hills, Illinois, USA). Dead eggs were removed on incubation day 30 (eyed egg stage) and the remaining viable eyed eggs were returned to their respective incubation tray.

**Egg Measurement**

Ten eyed eggs from each of the individual female spawns were then randomly selected and placed within round, wire-mesh, labeled baskets (8.9 cm diameter, 4.5 cm deep), and randomly placed into one of two semi-square 190 L tanks. Each tank contained 12 baskets, with each basket containing eyed eggs from an individual female. The diameters of three eggs from each spawn were randomly selected and measured to the nearest 0.01 mm using a digital caliper (SPC O/P Digimatic Caliper; Mitutoyo America Corporation, Aurora, Illinois, USA). Tank flows were set to 15 L/min using the same water source as the incubation stack. The egg baskets were checked daily, and the number of eggs that had hatched or died was recorded. Percent (%) egg survival to hatch was calculated by dividing the initial number of eggs (10) by the number of eggs that survived, and multiplying this amount by 100.

**Data Analysis**

Regression and correlation analysis was used to examine possible relationships between the variables. All data analysis was done using the SPSS (9.0) statistical analysis program (SPSS, Chicago, Illinois, USA), with significance predetermined at p<0.05. This experiment was carried out within the American Fisheries Society Guidelines for the Use of Fishes in Research and within the guidelines of the Aquatics Section Research Ethics Committee of the South Dakota Department of Game, Fish and Parks.

**RESULTS**

Mean egg diameter was 7.22 mm and ranged from 6.36 to 8.36 mm (Table 1). Over 90% of the eggs survived, with hatching occurring on average from 46 to 49 days of incubation.

Significant relationships were observed between egg diameter and initial hatch (p=0.003; Figure 1), as well as between egg diameter and mean hatch (p=0.027; Figure 2). The relationship between egg diameter and final hatch approached significance (p=0.068; Figure 3). There was no correlation between egg diameter and survival to hatch (r=-0.363, p=0.081).

<table>
<thead>
<tr>
<th>Variable</th>
<th>Mean</th>
<th>SE</th>
<th>Minimum</th>
<th>Maximum</th>
</tr>
</thead>
<tbody>
<tr>
<td>Egg diameter (mm)</td>
<td>7.22</td>
<td>0.13</td>
<td>6.36</td>
<td>8.36</td>
</tr>
<tr>
<td>Survival to hatch (%)</td>
<td>91.3</td>
<td>2.4</td>
<td>60</td>
<td>100</td>
</tr>
<tr>
<td>Initial hatch (day)</td>
<td>46.6</td>
<td>0.3</td>
<td>43.0</td>
<td>48.0</td>
</tr>
<tr>
<td>Mean hatch (day)</td>
<td>47.8</td>
<td>0.2</td>
<td>46.1</td>
<td>49.8</td>
</tr>
<tr>
<td>Final hatch (day)</td>
<td>49.0</td>
<td>0.2</td>
<td>47.0</td>
<td>50.0</td>
</tr>
</tbody>
</table>

Figure 1: The relationship between egg diameter (mm) and initial hatching time (d) for landlocked fall Chinook salmon from Lake Oahe, South Dakota.

Figure 2: The relationship between egg diameter (mm) and mean hatching time (d) for landlocked fall Chinook salmon from Lake Oahe, South Dakota.

Figure 3: The relationship between egg diameter (mm) and final hatching time (d) for landlocked fall Chinook salmon from Lake Oahe, South Dakota.
DISCUSSION

The positive relationship between egg size and hatching time observed in this study is supported by the results reported by Thorpe et al. (1984) and Self et al. (2018) in Atlantic salmon and steelhead trout, respectively. However, Einum and Fleming (2000) reported no such relationship in a different population of Atlantic salmon. Kinnison et al. (1998) stated that egg size was not related to hatching time in a New Zealand population of Chinook salmon. Likewise, other studies have also found no relationship between egg size and hatching time in salmonids (Beacham and Murray, 1985; Beacham et al., 1985; Leblanc et al., 2016). What are some possible reasons for the discrepancies among the studies?

Egg size varies between populations of fish species, between fish within a population, and also within the spawn of individual females (Beacham and Murray, 1987; 1993; Einum, 2003; Leblanc, 2016; Self et al., 2018). This variation may possibly explain the different results among the studies examining egg size in relation to hatch timing. Indeed, in comparison to ocean-run Chinook salmon in their native range, the landlocked, completely freshwater Lake Oahe salmon population produces smaller females that exhibit dramatically different reproductive characteristics (Barnes et al., 2000; Young et al., 2016). The differing results may also be due to the different egg incubation temperatures used in each study (Réalis-Doyelle et al., 2016; Fuhrman et al., 2018). Lastly, different techniques have been used to determine egg size. For example, egg size was defined as egg mass by Beacham et al. (1985), Kinnison et al. (1998), and Self et al. (2018), whereas egg size was defined as egg diameter by Pauly and Pullin (1988) and this study.

In this study, the size of the landlocked fall Chinook salmon eggs was comparable with eggs from Chinook salmon in their native range (Rounsefell, 1957; Beacham and Murray, 1988; Beacham and Murray, 1993). This is somewhat unusual. Lake Oahe salmon eggs are typically much smaller than those from their native range counterparts (Barnes et al., 2000; Young et al., 2016), likely because Lake Oahe Chinook salmon females are also smaller in weight (Barnes et al., 2000).

The water temperature used in this study was well within the optimal incubation temperature range of 5.5 to 16°C (Piper et al., 1982). In addition, the hatching time of 46 to 49 days (506 to 539 daily temperature units) is similar to the 510 to 529 daily temperature units needed for eggs from native range Chinook salmon incubated at 12.0°C (Beacham and Murray, 1988).

CONCLUSION

This is the first documentation of a positive relationship between egg size and hatching time in landlocked fall Chinook salmon from Lake Oahe, South Dakota, USA. This relationship is far from universal among salmonids, and may be unique to the Lake Oahe Chinook salmon population.

ACKNOWLEDGEMENT

We thank Jill Voorhees for her assistance with this project.
REFERENCES


