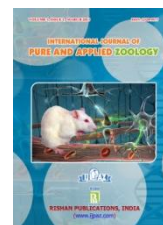




ISSN Print/Online: 2320-9577/2320-9585
 INTERNATIONAL JOURNAL OF PURE AND APPLIED ZOOLOGY
 Volume 1, Issue 1, March 2013
 Available online at: <http://www.ijpaz.com>
 RISHAN PUBLICATIONS



RESEARCH ARTICLE

OPEN ACCESS

LAMBDA CYHALOTHRIN INDUCED BIOCHEMICAL AND HISTOLOGICAL CHANGES IN THE LIVER OF *OREOCHROMIS MOSSAMBICUS* (PETERS)

K. MUTHUKUMARAVEL*, O. SATHICK AND S. RAVEENDRAN
 P. G. and Research Department of Zoology, Khadir Mohideen College,
 Adirampattinam-614 701, Tamil Nadu, India.

*Corresponding author's e-mail address: kumar_phd_2003@yahoo.co.in, Tel +91 9791387363.

Article History: Received: 26.02.2013, Accepted: 09.3. 2013

ABSTRACT

To investigate the sublethal effects of pesticide lambda cyhalothrin on biochemical and the liver histology of *Oreochromis mossambicus* under long term exposure. The toxicity tests were conducted following method of Finney (1964). Based on the acute toxicity assay the fish were exposed to 0.0025 ppm sublethal concentration. The glycogen content in liver tissue and glucose content in blood were estimated at the end of 10, 20 and 30 days. The liver tissues were removed and fixed in Bouin's fluid and then they processed for microphotographs. Glycogen content was found to be depleted in liver at all periods after treatment. In contrast to the observation of glycogen, the glucose level was elevated in the blood at all periods. In control tilapia, the liver was comprised of polygonal hepatocytes with centrally placed nucleus. Pesticide induced dilation of blood sinusoids, vacuolization and disintegration of cell boundaries were noticed in the 10 and 20 days of exposed fish. The complete damage of hepatocytes and loss of integrity of cell wall was noticed about 30 days of exposure. From the results stated above it can be concluded that the finding of the present study indicate that histological changes observed serve as "biomarkers" for assessing pesticide toxicity in aquatic environment.

Key words: Lambda cyhalothrin, toxicity, liver, histology and *O. mossambicus*.

INTRODUCTION

Pesticides have been reported to make a revolution in the agriculture production in India by controlling pests of various crops (Matsumura, 1985). Pesticides used in pest control programmes resulted in contaminating the ecosystem and entering food chains causing damaging effects on the ecosystem and non-target species (Ruvio, 1972 and Bakre *et al.*, 1990).

Lambda cyhalothrin is a synthetic pyrethroid insecticide widely used for pest

management and public health applications to control insects. Lambda cyhalothrin is categorized as a restricted use pesticide in the Extension Toxicology Network for its toxicity to fish (Mound, 1998). But its usage for the control of major pest in agriculture is being continued in developing countries like India. The pesticide exposure causes severe alterations in the tissue biochemistry and histology of fishes (Velisek *et al.*, 2009; Anita Susan, *et al.*, 2010 and Saravanan, *et al.*, 2010). In view of this in the present study the effect of sublethal concentration of lambda

cyhalothrin on the liver biochemistry and histology of a freshwater teleost, *Oreochromis mossambicus*.

MATERIALS AND METHODS

The fish, *Oreochromis mossambicus* weighing approximately 20g were collected from ponds in and around Adirampattinam. They were acclimatized for 15 days in large cement tanks (temperature 28 ± 2 °C; pH 8.0 ± 0.02 & DO 6.2 ± 0.4) previously washed with 1% potassium permanganate. The water was renewed every 24 hr. A stock solution of Lambda cyhalothrin was prepared and the toxicity tests were conducted following method of Finney (1964). Based on the acute toxicity studies (96 h), LC₅₀ value for test fish was found to be 0.025 ppm. For biochemical and histological studies, *O. mossambicus* were reared in 10% sub lethal concentration for a period of 10, 20 and 30 days. After treatment, both the control and experimental fishes were sacrificed at the end of 10, 20 and 30 days. The glycogen content was estimated in liver tissue by the method as described by Seifter

et al.(1950) and the blood glucose was estimated by O-toluidine method of Hartel *et al.*(1969). The liver tissues were removed and fixed in Bouin's fluid and then they processed adapting the usual procedure (Gurr, 1950). The sections (8µm) were stained with Haematoxylin and Eosin.

RESULTS

Blood glucose and liver glycogen interrelationship

The fish was exposed to sublethal concentration for a period of 30 days. Glycogen content was reduced in liver of pesticide exposed fishes with a concurrent increase in blood glucose concentrations. The percentage change in the liver over the control was decreased by -12.07, -29.71 and -57.77 at 10, 20 and 30 days of exposure (Table: 1). The increased trend of blood sugar level over the control was observed at sublethal concentrations at respective exposure period. The increased percentage changes over the control were +18.92, + 47.84 and + 93.29 (Table 2).

Table 1. Glycogen content (mg/gm) in the liver of *O. mossambicus* exposed to lambda cyhalothrin.

Treatment	Exposure period (Days)		
	10	20	30
Control	28.67±0.42	27.39±0.23	28.82±0.37
Sublethal	25.21±0.16	19.25±0.31	12.17±0.26
% Change	-12.07	-29.71	-57.77

Values are mean ±SE of 6 individual observations.

Table 2. Blood glucose levels (mg/100ml) in the *O. mossambicus* exposed to lambda cyhalothrin.

Treatment	Exposure period (Days)		
	10	20	30
Control	36.42 ± 0.24	39.51 ± 0.38	38.47 ± 0.29
Sublethal	43.31 ± 0.37	58.41± 0.26	74.36± 0.41
% Change	+18.92	+ 47.84	+ 93.29

Values are mean ±SE of 6 individual observations.

Histology of fish liver

In untreated *O. mossambicus*, the liver is made up of continuous mass of hepatocytes arranged in irregular cords. The hepatic cells are polygonal in shape with distinct central nuclei. A large number of blood sinusoids were also seen around the hepatocytes (Figure1).

Pathology of liver tissue under Lambda cyhalothrin toxicity

Marked toxic effects were observed at the structural and cellular level in the liver. In 10 days exposed fish, disintegration of cell boundaries and slight dilation of blood sinusoid was observed (Figure 2). After 20 days, many hepatic cells were completely damaged. Intracellular vacuolation was also apparent (F Figure 3). After 30 days, damage to the hepatocytes was prominent. In most of the hepatic cells the integrity of cell wall was completely lost (Figure 4).

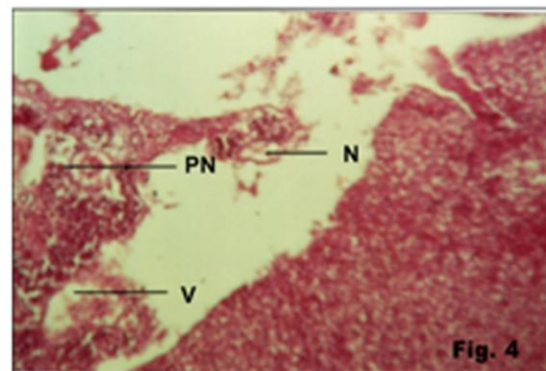
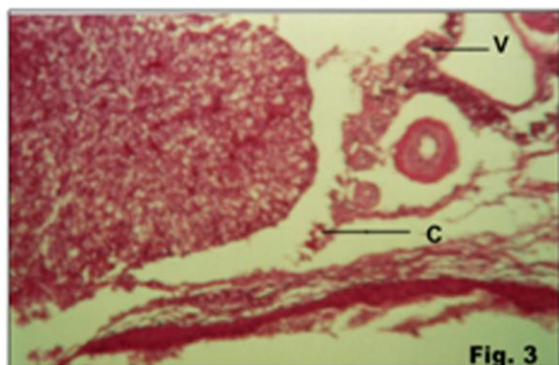
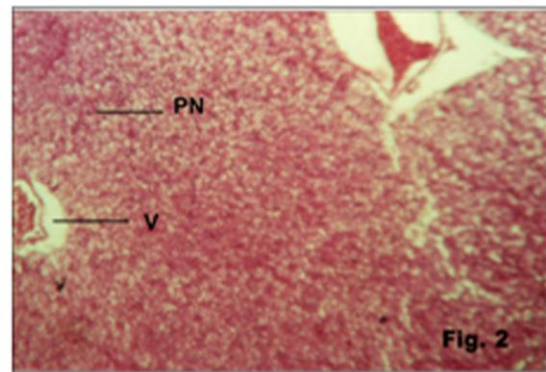
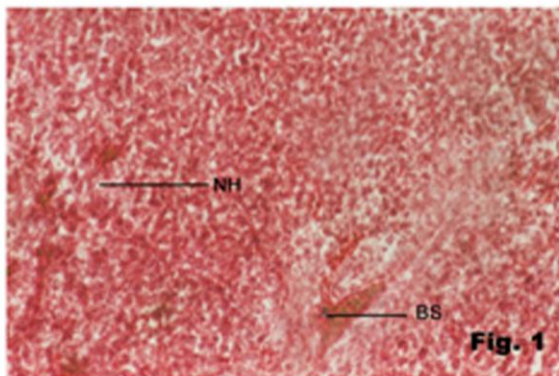


Figure 1. Control liver of *O. mossambicus*: NH-Normal hepatocytes and BS-blood sinus.

Figure 2. Exposed liver of *O. mossambicus* in 10% SLC of lambda cyhalothrin(10days): PN- pycnotic nuclei and V- vacuolization.

Figure 3. Exposed liver of *O. mossambicus* in 10% SLC of lambda cyhalothrin(20days): C- cytolysis and V- vacuolization.

Figure 4. Exposed liver of *O. mossambicus* in 10% SLC of lambda cyhalothrin(30days): PN- pycnotic nuclei,V- vacuolization and N-necrosis.

DISCUSSION

The hepatic cells have many vital functions such as secretion of bile, detoxification, synthesis of several components of blood plasma, storage of glycogen and release of glucose in the blood. Glycogen content was found to be depleted in liver tissue at all periods after treatment with a sublethal dose of lambda cyhalothrin. The results suggest that glycogen being a ready source of energy, reduction in glycogen is probably due to its more rapid breakdown (glycogenolysis) which releases glucose into the circulatory system to meet the increased energy requirements in a stressed condition. Sastry and Subhadra(1982), Larsson and Haux (1982) and Haux and Larsson (1984) have also made similar observations in cadmium treated *Heteropneustes fossilis* and *Salmo gairdneri*. The observations of Palanivelu *et al.*(2004) on glycogen level in cartap hydrochloride treated *O. mossambicus* was also comparable to those of the present study. In this study the blood sugar gradually increasing with increase in exposure period. Elevation of blood glucose level has been reported in *Labeo rohita* exposed to heavy metals (Neha Bhatkar *et al.*,2004). A similar trend has also been observed in mercury treated fish, *L. rohita* (Rajasubramaniam, 2006).

Morphological and histological alterations related to pesticide toxicity in the liver of fish have been showing that the substances cause severe damage to the liver cells (Ahamed and Srivastava, 1985; Dutta *et al.*,1993 and Ortiz *et al.*, 2003). Liver is an important organ of detoxification and biotransformation process and due to these reasons the hepatic cells are damaged severely. Several works have reported degenerative changes in hepatic tissue subjected to pollution by various pesticides and insecticides (Kumar and Pant, 1984; Gill *et al.*, 1990; Pandey *et al.*,1993; Tilak *et al.*, 2005; and Sakr and Jamal Al Lail, 2005). Dubale and Shah (1979) reported that *Channa punctatus* under malathion toxicity showed the degenerative changes in liver. Radhaiah and Jayantha Rao (1992) reported moderate cytoplasmic degeneration in hepatocytes, formation of vacuoles, rupture of blood vessels and

picnotic nuclei in the liver of *Tilapia mossambica* exposed to fenvalerate. The liver of pesticide treated fish showed dilation of blood sinusoids, vacuolization, disintegration of cell boundaries and necrosis. The present results are in agreement with those observed by many authors who studied the effects of different pollutants of fish liver (Mohamed,2001; Ptashynski *et al.*, 2002 and Fanta *et al.*, 2003). According to Rodrigues *et al.* (2001) the liver is an organ that frequently undergoes changes when exposed to insecticide at sublethal doses. The changes may be attributed to direct toxic effects of pollutants on hepatic cells, since the liver is the site of detoxification of all types of toxic substances (Soufy *et al.*, 2007).

CONCLUSION

The finding of the present study indicates that the biochemical and histological changes observed serve as biomarkers for assessing pesticide toxicity in aquatic environments.

ACKNOWLEDGEMENT

The authors are thankful to the Secretary and Principal, Khadir Mohideen College, Adirampattinam for providing necessary laboratory facilities.

REFERENCES

- Ahmad,G. and Srivastava, G.J. 1985. Histopathologic alterations in the liver and skin of a fresh water teleost, *Heteropneustes fossilis* (Block) exposed chronically to a sublethal concentration of methylene blue. *Pakistan M. Zool.*, 17: 229-246.
- Anita Susan, T. Sobha, K., Veeraiah, K. and Tilak, K.S. 2010. Studies on biochemical changes in the tissues of *Labeo rohita* and *Cirrhinus mrigala* exposed to fenvalerate technical grade. *J. Toxiol. Environ. Health Sci.*, 2(5): 53-62.

- Bakre, P.R., Mishra, B. and P. Bhatnagar, B. 1990. Residues of organochlorine insecticides in fish from Malala water reservoir, Jaipur, India. *Bull. Environ. Contam. toxicol.*, 45: 394-398.
- Dubale, M.S., Shah, P. 1979. Histopathological lesions induced by malathion in the liver of *Channa punctatus*. *Ind. J. Exp. Biol.*, 17(7): 693-697.
- Fanta, E., Rios, F., Romao, S., Vianna, A. and Freiburger, S. 2003. Histopathology of the fish *Corydoras paleatus* contaminated with sublethal levels of organophosphorus in water and food. *Ecotoxicol. Environ. Safety*, 54(2):119-130.
- Finney, D.J. 1964. Probit analysis. Cambridge University Press. Cambridge, London. P: 333.
- Gill, T.S., Pandey, J.S. and Tewari, H. 1990. Hepatopathotoxicity of three pesticides in a freshwater fish, *Puntius conchorius* Ham. *J. Environ. Sci. Hlth.*, 25A:653-663.
- Gurr. E. 1950. Methods of analytical histology and histochemistry. Leonard Hill Hd. London. Pp:45-49.
- Hartel, A.R., Helger, R. and Lang, H. 1969. A method of determination of glucose. *J. Clin. Chem. Biochem.*, 7: 183.
- Haux, C. and Larsson, A. 1984. Long term sublethal physiological effects on rainbow trout, *Salmo gairdneri* during exposure to cadmium and after subsequent recovery. *Aqu. Toxic*, 5: 129-142.
- Kumar, S. and Pant, S.C. 1984. Organal damage caused by aldicarb to a fresh water teleost, *Barbus conchoniuis* Hamilton. *Bull. Environ. Contam. Toxicol.*, 33: 50-55.
- Larsson, A. and Haux, C. 1982. Altered carbohydrate metabolism in fish exposed to sublethal levels of cadmium. *J. Environ. Biol.*, 3: 71-81.
- Matsumura, F. 1985. Toxicology of insecticides. Plenum press, New York.
- Mohamed, F.A. 2001. Impacts of environmental pollution in the southern region of Lake Manzalah, Egypt, on the histological structures of the liver and intestine of *Oreochromis niloticus* and *Tilapia zillii*. *J. Egypt. Acad. Soc. Environ. Develop.*, 2: 25-42.
- Mound, S.J., Hamer, M.J. Warinton, J.S and Kedwards T.J. 1998. Aquatic toxicology of the pyrethroid insecticide lambda cyhalothrin; consideration for higher-tier aquatic risk assessment. *Pestic. Sci.*, 54: 408-417.
- Neha Bhatkar, Vankhede, G.N. and Dhande, P.R. 2004. Heavy metal induced biochemical alterations in the fresh water fish *Labeo rohita*. *J. Ecotoxicol. Environ. Monit.*, 14(1): 1-7.
- Ortiz, J.B., Gonzalez de Canales, M.L. and Sarasquete, C. 2003. Histopathological changes induced by London (HCH) in various organs of fishes. *Sci. Mar.*, 67(1):53-61.
- Palanivelu, V., Vijayavel, K. Ezhilarasai Balasubramanian S. and Balasubramanian, M.P. 2004. Combined effects of cartap hydrochloride and urea on biochemical constituents of fresh water fish *Oreochromis mossambicus*. *J. Ecotoxicol. Environ. Monit.*, 14(1): 23-29.
- Pandey, A.K., George, K.C. and Peer Mohamed, M. 1996. Histopathological alterations induced in the liver of an estuarine mullet, *Liza*

- parsia* by mercuric chloride and DDT. *Indian J. Fish*, 43(3): 277-284.
- Ptashynski, M., Pedlar, Evan, R. Baron, C. and Klaver Kamp, J. 2002 Toxicology of dietary nickel in lake white fish (*Coregonus clupeaformis*). *Aquat. Toxicol.*, 58: 229-247.
- Radhaiah, V. and Jayantha Rao, K. 1992. Fenvalerate toxicity to the liver in a freshwater teleost, *Tilapia mossambica* (Peters). *Comp. Physiol. Ecol.*, 17(2): 48-53.
- Rajasubramaniam, V. 2006. Effect of divalent mercury on liver, muscle glycogen and blood glucose levels of freshwater teleost *Labeo rohita*. *J. Ecotoxicol. Environ. Monit.*, 16(5): 431-435.
- Rodrigues, E.D.L., Ranzani-Paiva, M.J.T., Pacheco, F.J. and Veiga, M.I.D. 2001. Histopathologic lesions in the liver of *Prochilodus lineatus* (Pisces, Prochilodontidae) exposed to a sublethal concentration of the organophosphate insecticide Dipterex 500 (Trichlorfon). *Maringa*, 23(2): 5003-505.
- Ruvio, M. 1972. Marine pollution and sea life. FAO News Ltd., London.
- Sakr, S.A. and Jamal Al Lail, S.M. 2005. Fenvalerate induced histopathological and histochemical changes in the liver of the cat fish *Clarias gariepinus*. *J. Applied Sci. Res.*, 1(3): 263-267.
- Saravanan, T.S., Rajesh, P. Sundarmoorthy, P.M. 2010. Studies on effect on chronic exposure of endosulfan to *Labeo rohita*. *J. Environ. Biol.*, 31(5): 755-758.
- Sasatry, K.V. and Subhadra, K. 1982. Effect of cadmium on some aspects of carbohydrate metabolism in a fresh water cat fish *Heteropneustes fossilis*. *Toxicol. Letter*, 14: 45-55.
- Seifter, S., Dayton, S., Novic, B., and Muntwyler, E. (1950). Estimation of glycogen with Anthrone reagents. *Arch. Biochem.* 25, 191-200(1950).
- Soufy, H., Soliman, M., El-Manakhy, E. and Gaafa, A. 2007. Some biochemical and pathological investigations on monosex tilapia following chronic exposure to carbofuran pesticides. *Global Veterinaria*, 1: 45-52.
- Tilak, K.S., Veeraiah, K. and Koteswara Rao, D. 2005. Histopathological changes observed in the gill, liver, brain and kidney of the Indian major carp *Cirrhinus mrigala* (Hamilton) exposed to chlorpyrifos. *Poll. Res.*, 24(1): 101-111.
- Velisek, J., Svobodova, Z. and Piackova, V. 2009. Effects of acute exposure to bifenthrin on some haematological, biochemical and histopathological parameters of rainbow trout (*Oncorhynchus mykiss*). *Veterinarni Medicina*, 54(3):131-137.

Cite this article as:

- K. Muthukumaravel, O. Sathick and S. Raveendran, 2013. Lambda cyhalothrin induced biochemical and histological changes in the liver of *Oreochromis mossambicus* (Peters). *Int. J. Pure Appl. Zool.*, 1(1): 80-85.
-