Histomorphological analysis of peri-implant bone tissue of an implant in use for 10 years: A case report.

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Abstract

In this case report, it was described the histomorphology through optical microscopy of the implant-bone interface in an implant in function during 10 years, which had to be removed due to problems associated with the connection of the abutment. A 73-year-old male patient came asking for prosthesis mobility. Due the impossibility to replace the abutment it was decided to remove the implant with a 5 mm internal diameter trephine and analyze this sample histomorphologically. The histological analysis showed a mature trabecular tissue between the implant threads with a regular cell distribution and lamellar organization of calcified matrix, observing that bone tissue follows the shape of the implant’s surface with which it was in contact. Also, was observed low osteoclast activity, collagen type I, without signs of inflammation or resorption. In conclusion, implants have been shown in this case report, as well as by studies in humans to have long duration and outstanding biocompatibility that permits the formation of mature and regular peri-implant bone tissue in certain conditions.

Keywords: osseointegration, peri-implant tissue, bone-implant interface; histomorophology.

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Introduction

The use of dental implants to support prostheses is a common practice today [1], because of its high success rate, which varies between 90% and 95% in patients with no or few risk factors [2]. However, the success and long-term duration of this type of treatment is directly related to the formation of a bone–implant contact [3], which is also referred as osseointegration. The osseointegration process was originally defined as a direct structural and functional connection between the bone and the implant [4], which has been histologically demonstrated [5]. This phenomenon is a complex process with a multitemporal and multiscale nature [6].

Bone has the ability to regenerate being indistinguishable from the underlying bone tissue [7]. Histological studies of properly osseointegrated implants describe newly formed bone tissue in close contact with the implant and with a regular organization [5]. Other studies in animals show similar histological results [8] and have also made it possible to detail the process of osseointegration of implants in successive periods of time [9], a matter that is difficult to achieve in humans for bioethical reasons.

Despite a high success rate [10], implant failure may occur in some cases. Early failure may be due to factors such as inadequate surgical techniques [11], quality of and/or insufficient bone, patient's unhealthy habits, patient's systemic disease, and contamination during surgery [12,13], among others. Late implant failure, defined as a pathological process involving an osseointegrated implant, is less understood but is classified as overload or infection [11].

The aim of this study was to describe the histomorphology through optical microscopy of the tissue around a dental implant functioning for 10 years, which had to be removed due to problems associated with the connection of the implant with the abutment.

Case report

Patient and procedure

A 73-year-old male patient without systemic diseases or negative health habits came asking about prosthesis mobility. At clinical examination, damage was identified in the inner thread of the posterior–superior endosteal implant. A review of his medical record indicated that the implant (Ankylos, Dentsply, New York, NY, USA) of 3.5 mm in diameter and 11 mm in length, had been in place for 10 years and 3 months. At the time of implantation, the bone level was optimal, so no ridge augmentation techniques were used.
Histomorphological analysis of peri-implant bone tissue

Figure 1. Mature trabecular tissue between implant threads of the implant. White arrows: lamellar apposition lines. MT: medullar spaces (Van Gieson Stain).

Figure 2. Lamellae have an organization of parallel apposition to the implant surface. Black arrows: osteocytic gaps with low metabolic activity. White arrows: lamellar apposition lines. MT: medullar spaces (Hematoxylin-Eosin stain).

Figure 3. All regions analyzed (A to D) showed a bright red appearance characteristic for the presence of collagen type I. Presence of type III collagen was not observed in bone tissue, which is characteristic of an early bone regeneration step. In addition, the white arrows indicate the organization of the collagen fibers in the form of overlapping layers, typical from lamellar bone in an advanced stage and properly organized (Picosirius Red stain, polarized light).

No pain, swelling, or implant mobility were observed. However, due the impossibility to replace the abutment it was decided to remove the implant with a 5 mm internal diameter trephine. The patient agreed with the suggested treatment and authorized through an informed consent the implant site extraction and histological examination of the sample. After removal of the implant, the resulting defect was filled with a biomaterial (DynaGraft-D putty, Keystone Dental, Burlington, MA, USA) for achieve bone tissue regeneration in order to schedule surgical placement of a new implant at a later date.

Both the implant and the surrounding bone tissue were put in a bottle with 10% buffered formalin and stored at 4°C for 3-4 weeks.

Bone-implant separation and histological analysis
The sample was washed with PBS 1X 0.01M and then placed in a beaker with 300 mL of 10% ethylenediaminetetraacetic acid (EDTA) with 2% paraformaldehyde (pH 6.8) to partially decalcify and decrease the hardness of the bone tissue, generating a separation of the implant and bone tissue. Both solutions were changed every 6 hours for 15 days to obtain adequate decalcification.
Table I. Histological studies in humans. Surrounding tissue to implants properly osseointegrated with follow-up periods of 0.5 to 22 year

<table>
<thead>
<tr>
<th>Authors</th>
<th>Number of implants</th>
<th>Functional loading time (years)</th>
<th>Cause of removal of implants</th>
<th>Main histological findings</th>
</tr>
</thead>
<tbody>
<tr>
<td>Hansson et al., 1983</td>
<td>Not reported</td>
<td>0.5 to 7</td>
<td>Psychiatric reasons</td>
<td>Compact and well-organized bone. Haversian canals near the implant. Blood vessels frequently observed. No evidence of connective tissue, fibroblasts, or macrophages.</td>
</tr>
<tr>
<td>Prousseafs et al., 2000</td>
<td>2</td>
<td>7</td>
<td>Excessive vestibular inclination of implants made it difficult to retain the overdenture. Correctly integrated implants; bone tissue in contact with the surrounding tissue; Haversian canals near the implant.</td>
<td></td>
</tr>
<tr>
<td>Uehara et al., 2004</td>
<td>2</td>
<td>1.5</td>
<td>Abutment connection fracture</td>
<td>Dense and mineralized bone closely related to the implant. Lamellar bone, uniformly distributed and trabeculae functionally oriented.</td>
</tr>
<tr>
<td>Trisi et al., 2005</td>
<td>2</td>
<td>10</td>
<td>Postmortem</td>
<td>Intimate bone–implant contact. Lamellar bone was observed in different directions. No epithelial or connective tissue migration was observed.</td>
</tr>
<tr>
<td>Coelho et al., 2009</td>
<td>30</td>
<td>8 to 13 years</td>
<td>Prosthodontics</td>
<td>Mature/compact lamellar bone. Few and small core areas. Osteons and Haversian canals near the implant; no epithelial proliferation, bacteria, or inflammatory cells.</td>
</tr>
<tr>
<td>Iezzi et al., 2012</td>
<td>1</td>
<td>22</td>
<td>Mandibular resection</td>
<td>Mature/compact lamellar bone. Few and small core areas. Osteons and Haversian canals near the implant; no epithelial proliferation, bacteria, or inflammatory cells.</td>
</tr>
<tr>
<td></td>
<td>1</td>
<td>18</td>
<td>Prosthodontics</td>
<td>Lamellar bone, uniformly distributed and trabeculae functionally oriented. Lamellar bone was observed in different directions. No epithelial or connective tissue migration was observed.</td>
</tr>
<tr>
<td></td>
<td>1</td>
<td>5</td>
<td>Misalignment</td>
<td>Lamellar bone, uniformly distributed and trabeculae functionally oriented. Lamellar bone was observed in different directions. No epithelial or connective tissue migration was observed.</td>
</tr>
<tr>
<td></td>
<td>1</td>
<td>10</td>
<td>Fracture</td>
<td>Lamellar bone, uniformly distributed and trabeculae functionally oriented. Lamellar bone was observed in different directions. No epithelial or connective tissue migration was observed.</td>
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<tr>
<td></td>
<td>1</td>
<td>14</td>
<td>Fracture</td>
<td>Lamellar bone, uniformly distributed and trabeculae functionally oriented. Lamellar bone was observed in different directions. No epithelial or connective tissue migration was observed.</td>
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<tr>
<td></td>
<td>1</td>
<td>5</td>
<td>Psychological</td>
<td>Lamellar bone, uniformly distributed and trabeculae functionally oriented. Lamellar bone was observed in different directions. No epithelial or connective tissue migration was observed.</td>
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<td></td>
<td>1</td>
<td>6</td>
<td>Prosthodontics</td>
<td>Lamellar bone, uniformly distributed and trabeculae functionally oriented. Lamellar bone was observed in different directions. No epithelial or connective tissue migration was observed.</td>
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</table>

Under a stereoscopic microscope with 10X magnification, it was analyzed at the bone–implant interface at the level of the bottom of the most apical thread, where a thin metal probe was carefully inserted to separate the tissue carefully with a micro-incision metal clamp serrated 20 Ga to hold its shape. Once completely separating the bone tissue from the metal, it was immersed in EDTA to complete its decalcification.

The tissue sample was cut 3 µm longitudinally with a microtome (Microm HM 325, Thermo Scientific, Florida, FL, USA) and subsequently stained using hematoxylin–eosin techniques, Picosirius Red and Van Gieson’s stain. The samples were analyzed using an optical microscope (Olympus, Arquimed Innovation, Santiago, Chile) with magnification 4x, 10x and 50x. Histological analysis was performed on all sections obtained.

The histological analysis showed:
- A mature trabecular tissue between the implant threads with a regular cell distribution and lamellar organization of calcified matrix (Fig. 1).
- Lamellae had a parallel organization to the surface of the implant, observing that bone tissue follows the shape of the implant’s surface with which it was in contact (Fig. 1 and 2).
- Sparse tissue fibroreticular regions and no presence of chondroid tissue; trabeculae described have morphology of bone tissue.
- The trabeculae showed moderate vascularity. Vessels of different sizes were observed both parallel and perpendicular to the surface of the implant.
- Low osteoclast activity and collagen type I (Fig. 3).
- Osteocytic lacunae had a consistent morphology with low metabolic activity (Fig. 2).
- No signs of inflammation or resorption activity were observed.

In general, the peri-implant tissue in this case report was a mature, regular, and vital bone, which was remodeled actively, presumably depending on the forces applied to the implant.

Discussion

Several studies have reported histomorphology of tissue surrounding implants with early loss [14,15] removed because of infection or mobility of the implant. In gen-
eral, this is described and characterized histologically for the presence of stratified connective tissue, proliferation of epithelial tissue, and inflammatory cells, as well as a lack of osseointegration [14]. Also, there have been reports of late implant failure, because of peri-implantitis or excessive occlusal load [16,17]. In both cases, osseointegration loss is observed. The main histological features of peri-implantitis consist of the presence of bone sequestra near the implant, bacteria on the surface of the implant, and inflammatory infiltrates (macrophages, lymphocytes, and plasma cells) in the adjacent area [18].

The traditional classification of failure in endosseous implant is defined as early or late, the first being a failure to achieve osseointegration and the second the inability to maintain it. However, as observed in this study and others [5,19], implant failure can also occur as a result of factors not directly related to osseointegration as such, so this is important to consider. Therefore, histological descriptions of the implants removed for different failures from those classically described, are important evidence of bone–implant interaction.

Several studies in humans and with follow-up periods between 0.5 and 22 years have confirmed histological findings described in this study (Table I). The studies reported to date indicate that the reasons for the removal of properly osseointegrated implants are of the following types: psychiatric [5,20], prosthetic [19-21], fracture of implant or the abutment [5,20,22] and/or mandibular resection [20]. Postmortem cases have also been reported, in which osseointegrated implants are removed for histological analysis [23]. Histology described in these articles is independent of patient gender, the position of the implant and time of functional loading of the implant. In general, it describes bone tissue in intimate contact with the implant [21,23] and regular and mature organization [5,20], as here described. In addition, osteocytes and Halversian systems are described near the bone–implant interface [5,19,20]. Finally, similar to that observed in this study, there is no evidence of connective tissue formation, presence of fibroblasts, macrophages, or inflammatory cells [5,20,21].

**Conclusion**

Endosseous implants have been shown in this case report, as well as by studies in humans, to have long duration and outstanding biocompatibility that permits the formation of mature and regular peri-implant bone tissue.

**References**


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