

## **Extracellular biosynthesis of silver nanoparticles from *Cyanobacteria*.**

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### **Abstract**

The field of nanotechnology is being now extensively studied by researchers particularly synthesis of nanoparticles with non-toxic, environmentally benign techniques and using mainly microorganisms including bacteria, fungi and algae. These bio-synthesized nanoparticles are aimed to be used as alternatives to the medical agents; however, their activity depends on their size, morphology, distribution and the source of bio synthesis. In this study silver nanoparticles were synthesized from three types of *Cyanobacteria* naming *Nostoc sp.*, *Scytonema sp.* and *Phormidium sp.* Higher antibacterial activity was obtained with *Scytonema* Nanoparticles (NP) extracts against MRSA followed by *Phormidium sp.* compared to other algal NP extracts against *Staphylococcus aureus*, *E. coli* and *Pseudomonas aeruginosa*. Chemical analysis of the studied algal NP extracts revealed higher peaks at 3278  $\text{cm}^{-1}$  and 3262  $\text{cm}^{-1}$  for *Phormidium* and *Nostoc* algal species, indicating the presence of NP particles to which the antibacterial activity is being attributed.

**Keywords:** *Scytonema*, *Phormidium*, *Nostoc*, *Cyanobacteria*, Antibacterial, Nanoparticles.

### **Abbreviations:**

Silver Nanoparticles (AgNPs); Silver Nitrate ( $\text{AgNO}_3$ ).

Accepted on June 25, 2018

### **Introduction**

Nanoparticle synthesis particularly from plants and algae nowadays is becoming the booming of most recent scientific research. However, the efficacy of these nanoparticles resides in the methods of synthesis where the biological techniques were found to be more effective than the chemical and the physical ones; it is also and most importantly linked to the type of algae or plants used.

Biological methods for synthesizing nanoparticles have shown better results compared to chemical and physical techniques due to time consumption, slow kinetics [1,2] low cost [3] and most importantly it is more eco-friendly compared to the chemical and physical techniques [4].

Algae used in the biosynthesis of nanoparticles have recently attracted the attention of many scientific researches [2] due to their rapid growth, to their biomass formation and to their different biological activities by nature. Extensive work has been conducted on different types of algae mainly *Spirulina platensis* [5-7] for the bio-synthesis of nanoparticles. So far, Mubarak et al., Mahdieh et al. and Pal [5-7] have used *Cyanobacteria* for the synthesis of silver and gold nanoparticles; they reported good antibacterial activity against some human pathogenic bacteria. Thus taking into consideration the importance of algae, two types of *Cyanobacteria* were investigated in this study namely *Nostoc*

*sp.* and *Scytonema sp.* together with *Phormidium sp.* for the biosynthesis of Silver Nanoparticles (AgNPs). The algal AgNPs were determined using the FTIR spectrophotometer and the transmission electron microscope, and then tested *in vitro* against some gram positive and gram negative bacteria.

### **Materials and Methods**

#### ***Algal samples growth and collection***

*Nostoc*, *Scytonema* and *Phormidium sp.* were obtained from King Saud University, Botany Department, Riyadh, and KSA. They were grown at the botany department female section in a BG 11 medium (pH 7) [8].

#### ***Preparation of AgNP extracts***

All algal samples were washed with sterile distilled water and kept to dry at room temperature. Dried samples were then ground into powder to be used for the synthesis of AgNPs when mixed to 1 mM silver nitrate solution prepared in dark reagent bottle (1 mg/2 ml) [9]. Extracts were kept in a shaking incubator at 140 rpm and 30°C until a color change occurred from pale yellow to dark brown indicating the synthesis of AgNPs. A small amount of  $\text{AgNO}_3$  solution without algal samples was kept as a negative control.

### Bacterial strains tested

The bacterial strains tested in the present work were all obtained from the Microbiology department, King Saud University and were as follows *Staphylococcus aureus* ATCC 25923, Methicillin Resistant *Staphylococcus aureus* (MRSA) ATCC3345 as gram positive bacteria whereas the gram negative bacteria used were namely *E. coli* ATCC 25966 and *Pseudomonas aeruginosa* ATCC 27853.

All bacterial strains were grown on nutrient agar plates and then fresh bacterial suspensions of 0.5 MacFarland turbidity were prepared in 5 ml sterile nutrient broth test tubes.

### Antibacterial assay

*In vitro* antibacterial activity was performed using agar well diffusion technique [10], where Mueller Hinton plates were inoculated first with the freshly prepared bacterial suspension separately using sterile cotton swabs; then the plates were perforated using sterile cork borer of 6 mm diameter into four equidistant wells to be filled with 70  $\mu$ l of each of the algal Ag NPs extracts correspondingly and the negative control AgNO<sub>3</sub> solution. Plates were incubated at 37°C for 24 h. The antibacterial activity was determined by measuring the inhibition zone around each well respectively and average of 3 repeated experiments were obtained. Data were compared to positive control standard antibiotic discs and to the negative control AgNO<sub>3</sub> solution.

### FTIR analysis

Chemical analysis using the FTIR spectrophotometer was performed on a Perkin-Elmer FTIR spectrophotometer (USA) in the diffuse reflectance mode at a resolution of 4 cm<sup>-1</sup> in KBr pellets to reveal the presence of AgNPs biosynthesis using the algal samples in study. Silver nanoparticles are observed at 3000-4000 cm<sup>-1</sup> absorption.

### Transmission Electron microscope TEM

5  $\mu$ l of *Phormidium* AgNPs extract were placed on a carbon-coated copper grid. Grids were dried under infrared lamp and inspected in TEM microscope (Joelle, Japan), operated at an accelerating voltage of 100 kV. All particles showed a circular shape and an average size of AgNPs (5-10 nm).

### Results and Discussion

The *Cyanobacteria* filtrates when added to 1 mM AgNO<sub>3</sub> changed in color from pale yellow to dark brown indicating the biosynthesis of silver nanoparticles which were further revealed by the FTIR analysis which indicated very strong absorption peaks of silver nanoparticles at 3278 and 3262 cm<sup>-1</sup> from *Phormidium*, *Scytonema* and *Nostoc* respectively. These bands were assigned to the stretching vibrations of primary and secondary amines respectively. The corresponding bending vibrations were seen at 1628.63 and 1634 cm<sup>-1</sup>. The band observed at 1317.48 cm<sup>-1</sup> can be assigned to C-N stretching vibration of amines. In Agreement with our findings, Mubarak

et al. [5] confirmed the presence of a protein shell which is responsible for the nanoparticles biosynthesis and considered as the stabilizing agent surrounding the AgNPs and to which this effective antibacterial activity could be correlated (Figures 1-3).

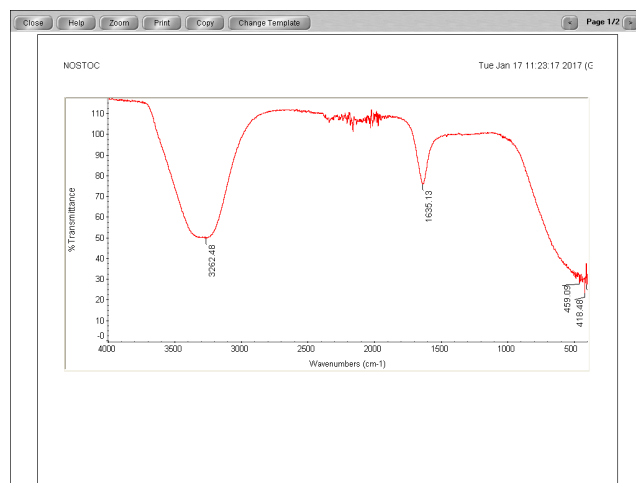


Figure 1. FTIR spectrum for *Nostoc* indicating the presence of AgNPs at 3262 cm<sup>-1</sup>.

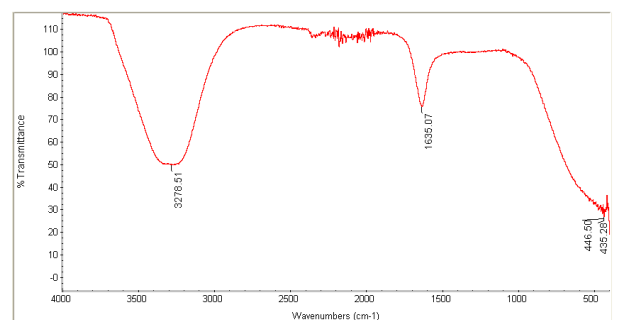


Figure 2. *Phormidium* FTIR spectrum revealing the presence of AgNPs at 3278 cm<sup>-1</sup>.

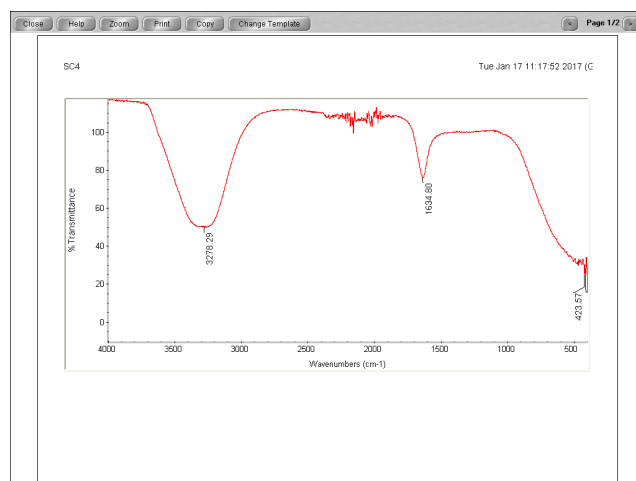


Figure 3. *Scytonema* AgNPs revealed at 3278 cm<sup>-1</sup>.

Previous studies have demonstrated that the antimicrobial nature of AgNPs to be size [11] and shape dependent [12] where smaller nanoparticles displayed better antimicrobial

activity TEM microscope was also applied in this study to measure and determine both the size and the morphology of the synthesized nanoparticles. Data showed that the size of the nanoparticles synthesized by the *Phormidium sp.* ranged between 5-10 nm, moreover all these particles were spherical in shape (Figure 4). Following the chemical composition analysis and the microscopic observation, the antibacterial activity of each of the algal nanoparticles extracts was performed against some gram negative and gram positive human pathogenic bacteria (Figure 4).

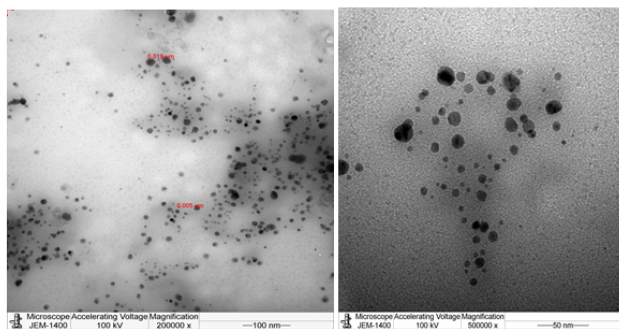


Figure 4. *Phormidium* TEM showing synthesized AgNPs of spherical shape and a size ranging between 5-10 nm.

Data obtained from this study showed greater antibacterial activity with *Scytonema sp.* silver extract followed by

Table 1. Average inhibition zone AgNPs synthesized by different algal species measured in mm compared to the silver nitrate AgNO<sub>3</sub> (negative control).

Average inhibition zone of Ag particles synthesized measured in mm				
Microorganisms	<i>Phormidium</i>	<i>Nostoc</i>	<i>Scytonema</i>	AgNO <sub>3</sub> (Negative control)
<i>Staphylococcus aureus</i> ATCC (25923)	9	10	7	9
MRSA ATCC (12498)	11.5	2	12	10
<i>Pseudomonas aeruginosa</i> ATCC (27853)	8	9	9	11.5
<i>E. coli</i> ATCC (25966)	10	9	9.5	10

### Conclusion

The extracellular biological synthesis of AgNPs from *Cyanobacteria* can be considered as one of the best techniques in the nanotechnology field since it is time and cost saving. A potent antibacterial activity was attributed to the size, shape and amount of the AgNPs synthesized and mostly to the type of the algal species; however, more intracellular investigations should be involved to determine the mechanism of action of these nanoparticles.

### Acknowledgment

This research project was supported by a grant from the “Research Center of the Center for Female Scientific and Medical Colleges”, Deanship of Scientific Research, King Saud University.

*Phormidium sp.* extract mainly against MRSA (12 mm and 11.5 mm) correspondingly, *Pseudomonas aeruginosa*, *E. coli* showed on the other hand an inhibition zone diameter ranging between 8 and 10 mm with *Phormidium sp.* and *Scytonema sp.* *Nostoc sp.* has the least antibacterial activity against MRSA (2 mm) but higher effect against *Staphylococcus aureus* (10 mm) compared to the other algal species nanoparticles extracts being studied, in agreement with [13,14] who reported that AgNPs have showed antibacterial activities against gram positive and gram negative bacteria. However, this slight difference in the antibacterial activity between the three algal samples in study (Table 1) could be explained by the aggregation, the size of the synthesized nanoparticles which may be correlated to the Surface Area and Volume (SA/V) ratio. As the size of AgNPs decreases the SA/V ratio for individual particles increases and the relative particle concentration also increases [15,16] thereby enhancing the antibacterial activity. Furthermore, the type of algal species may as well attribute to synthesizing nanoparticles with variable shapes and sizes and hence with variable potency of antibacterial activity [17]. As a conclusion, algal samples could be considered a good source of biosynthesized AgNPs applied in different therapeutic and environmental fields.

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