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### **Research Article**

# BIOEFFICACY OF CLEOME VISCOSA L. AND SINAPIS ALBA L. SEED EXTRACTS AGAINST HELICOVERPA ARMIGERA (HUBNER) (LEPIDOPTERA: NOCTUIDAE)

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### ABSTRACT

Solvent extracts of *Cleome viscosa* L. (Cleomaceae) and *Sinapis alba* L. (Brassicaceae) seeds were incorporated in semi-synthetic diet of *Helicoverpa armigera* (Hubner) at 0.5, 1.0, 1.5 and 2.0% concentrations to evaluate their antifeedant, larvicidal, pupicidal and adult malformation activities against *H. armigera*. Methanol extract of *S. alba* recorded very high antifeedant activity (71.42%) and pupicidal activity (54%) at 2% concentration. At 2% concentration the hexane extracts of *S. alba* (33.93%) and *C. viscosa* (32.86%) produced more deformities at adult stage of the insect. The results clearly indicated that the methanol extract of *S. alba* seeds have potential antifeedant activity against *H. armigera* and can be used to prevent crop damage by *H. armigera* in the field.

Keywords: Cleome viscosa, Sinapis alba, Podborer, Antifeedant, Pupicidal, Adult malformation.

# **INTRODUCTION**

Helicoverpa armigera is an agriculturally important insect. It is a polyphagous pest causing heavy yield loss in agricultural, ornamental and horticultural crops (Talekar et al., 2006). Worldwide, synthetic pesticides are commonly used for H. armigera control. But application of these synthetic pesticides has resulted in many effects unwanted such as environmental pollution, pesticide resistance, non-target effect and human health hazards (Raguraman and Singh, 1999). H. armigera has developed resistance to almost all classes of insecticides including pyrethroids (Babu and Subrahmanyam, 2010; Dhingra et al., 1988).

Since the past few years, people are aware of organic pesticides and are intended towards using botanical extracts to offer alternatives to synthetic insecticides (Ladhari *et al.*, 2013). Botanical pesticides have multiple modes of actions such as antifeedant activity, growth inhibiting, repellent and oviposition deterrent

activities (Metcalf et al., 1992). Plant-derived phytochemicals and have been extracts investigated on insect pests for the past 30 years alternatives conventional to develop to insecticides (Ca'ssia Seffrin et al., 2010). Solvent extracts of many plants have been reported as antifeedants, larvicides and growth regulators against H. armigera (Paulraj and Ignacimuthu, 2009; Baskar and Ignacimuthu, 2012; Pavunraj et al., 2013; Sivaraman et al., 2014). Aqueous extracts of neem seed and leaf were found to extend the larval developmental period and reduced adult emergence, longevity, fecundity and fertility in H. armigera (Wondafrash et al., 2012).

*Sinapis alba*, the White mustard, is an annual plant of the family Brassicaceae and is distributed worldwide. It is grown for its seeds, mustard, as fodder crop or as a green manure. The seeds of *S. alba* have many medicinal properties including anti-colon cancer property (Yuan *et al.*, 2011). The genus has a sub

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cosmopolitan distribution throughout the tropical and warm temperate regions of the world. Species of *Cleome* are commonly known as spider flowers, spider plants, spider weeds, or bee plants. The plant is used for treating various human ailments (Aparadh *et al*, 2012).

In the present study solvent extracts of *Cleome viscosa* and *Sinapis alba* seeds were screened against third instar larvae of *H. armigera* for antifeedant, larvicidal and growth inhibiting properties.

### MATERIALS AND METHODS

#### **Plant seeds**

Dry seeds (500gm) of *C. viscosa* and *S. alba* were purchased from the market at Parrys in Chennai, Tamil Nadu, India. The seeds were identified by taxonomist at Entomology Research Institute and voucher specimens (*C. viscosa*: ERI-BP-GS-008; *S. alba*: ERI-BP-GS-009) were deposited at the herbarium of the Institute.

### Extraction

The seeds of both plants were powdered coarsely powdered using electric blender. One part of seed powder (500 gm) was soaked sequentially in three parts (1.5L) of hexane, chloroform, ethyl acetate and methanol for a period of 48 h with intermittent shaking and filtered through a Buchner funnel using Whatman number 1 filter paper. The extracts were concentrated under reduced pressure using rotary evaporator (Medica instruments Mgf. Co. Sl. No: EV11. JF. 012) and stored at 4°C.

#### **Phytochemical analyses**

Phytochemical analysis of *C. viscosa* and *S. alba* seeds extracts were carried out following the method of Harbone (1998).

#### Insects

Third instar larvae of *H. armigera* were obtained from a pure stock culture maintained over ten generations on semi-synthetic diet (Vanderzant *et al.*, 1962) in the Institute laboratory at  $27\pm1^{\circ}$ C;  $11\pm0.5$  h photoperiod and 65-70% R.H.

#### **Antifeedant activity**

All the solvent extracts of the two seeds were screened at four different concentrations viz., 0.5, 1, 1.5 and 2%. Extracts were incorporated into the semi-synthetic diet as described by Nathala et al. (2006) and Jadhav et al. (2012). About 1 gm of extract-incorporated diet was weighed (AB54-S Meter Toledo Balance, Switzerland) and kept in petri-dish (13 mm  $\times$  90 mm). A single third-instar larva (3 h pre-starved) of H. armigera was placed on the treated diet taken in the petri-dish. Untreated diet was maintained as a control. After the treatment period of 24 h, the unfed diet in control and treatments were quantified and the larvae were reared continuously on fresh untreated semidiet. Twenty replications were synthetic maintained for each treatment and the control. The antifeedant activity was calculated according to the formula of Bentley et al. (1984):

Antifeedant activity =  $C-T/C \times 100$ 

Where 'C' is the consumption in control and 'T' is the consumption in treatment.

### Larvicidal bioassay

The treatments were given orally through semisynthetic diet to the larvae according to the antifeedant experiment bioassay method. After 24 h, the treated diet was removed and the larvae were continuously maintained on normal fresh semi-synthetic diet. Then larval mortality was calculated up to 96 h after the treatment. Control larvae were maintained separately on untreated diets. Ten replications were maintained for each concentration and each experiment was repeated five times.

#### Pupicidal activity and adult abnormalities

The survived larvae in the larvicidal experiments were continuously fed with untreated normal fresh diet pupation. Pupal mortality was calculated by subtracting the number of emerging adults from the total number of pupae. Abnormalities in adult moths were recorded and percentages of abnormal adults developed from each treatment were calculated.

# Statistical analysis

Data obtained from antifeedant, pupicidal and adult malformation experiments were subjected to one way Analysis of Variance (ANOVA) and mean values were separated by Tukey's multiple range tests using SPSS software package (11.5 version).

# RESULTS

Preliminary phytochemical analysis: Preliminary phytochemical analysis of seed extracts of two plants was done and the results are presented in table 1. The results clearly showed that the ethyl acetate extracts of both plant seeds contained more number of secondary metabolites. Ethyl acetate extract of S. alba showed positive results for five phytochemical groups namely alkaloids, coumarins, quinones, tannins and terpenoids. Ethyl acetate extract of C. viscosa contained four groups of secondary metabolites namely coumarins, guinones, tannins and terpenoids. The most effective treatment, i.e. the methanol extract of S. alba contained three

types of metabolites *viz.*, alkaloids, quinones and saponins.

Antifeedant activity: Table 2 shows the percentage of antifeedant activities of *C. viscosa* and *S. alba* seeds extracts against *H. armigera* third instar larvae at 0.5, 1.0, 1.5 and 2% concentrations. Methanol extracts of seeds of both plants were found to be the most effective. The significantly greater antifeedant activity (71.42%) was recorded in methanol extract of *S. alba* seeds at 2% concentration (p 0.01). The methanol extract of *C. viscosa* presented 50.91% antifeedant activity at 2% concentration. Hexane extracts of two plants presented the lowest antifeedant activity. In all the treatments, the antifeedant activity was directly proportional to the concentration.

*Pupicidal activities*: None of the treatments caused larval mortality during the observation period, but caused the pupal mortality. Table 3 shows pupicidal activity of *C. viscosa* and

Table 1. Preliminary phytochemical analysis of C. viscosa and S. alba seed extracts.

Plant species	Extracts -	Phytochemical groups								
		А	С	F	Ph	Q	S	St	Т	Тр
C. viscosa	Hexane	-	-	-	-	-	-	-	-	+
	Chloroform	-	-	-	-	+	-	-	-	+
	Ethyl acetate	-	+	-	-	+	-	-	+	+
	Methanol	+	-	-	-	+	-	-	+	-
S. alba	Hexane	-	-	-	-	-	-	-	-	+
	Chloroform	+	-	-	-	+	-	-	-	+
	Ethyl acetate	+	+	-	-	+	-	-	+	+
	Methanol	+	-	-	-	+	+	-	-	-

A-Alkaloids; C-Coumarin; F-Flavanoids; Ph – Phenols; Q- Quinones; S-Saponins; St- Steroid; T-Tannins; Tp-Terpenoids (+ indicates presence; – indicates absence).

Table 2. Antifeedant activity of C. viscosa and S. alba seed extracts against H. armigera larvae.

Plant	Extracts	Concentration (%)					
species	Extracts	0.5	1.0	1.5	2.0		
a	Hexane	$1.78 \pm 3.10^{d}$	$8.48 \pm 4.22^{e}$	11.51±3.54 <sup>f</sup>	23.71±6.05 <sup>e</sup>		
viscosa	Chloroform	$11.64 \pm 5.06^{\circ}$	17.13±9.29 <sup>de</sup>	$21.44\pm6.48^{e}$	$26.88 \pm 6.32^{de}$		
	Ethyl acetate	$25.00 \pm 6.72^{b}$	$30.85 \pm 7.24^{bc}$	38.37±6.13 <sup>bc</sup>	$47.81 \pm 8.13^{b}$		
Ċ.	Methanol	$23.34 \pm 5.62^{b}$	$38.09 \pm 6.18^{ab}$	$43.26 \pm 6.14^{b}$	$50.91 \pm 6.42^{b}$		
S. alba	Hexane	$16.62 \pm 8.68^{bc}$	$22.02 \pm 8.66^{cd}$	$28.35 \pm 7.94^{de}$	34.71±9.26 <sup>cd</sup>		
	Chloroform	$12.76 \pm 6.64^{\circ}$	$33.28 \pm 9.47^{b}$	$36.31 \pm 8.45^{bcd}$	41.57±8.33 <sup>bc</sup>		
	Ethyl acetate	$21.92 \pm 8.79^{b}$	$27.24 \pm 9.73^{bcd}$	29.91±8.56 <sup>cde</sup>	$42.87 \pm 9.16^{bc}$		
	Methanol	$43.53 \pm 8.93^{a}$	$47.69 \pm 12.06^{a}$	55.05±12.13 <sup>a</sup>	$71.42 \pm 9.96^{a}$		

Within the column, mean  $\pm$  SD followed by the same letter do not differ significantly (Tukey's test, P 0.05), (n=20).

Plant	Extra ata	Concentration (%)					
species	Extracts	0.5	1.0	1.5	2.0		
a	Hexane	14.0±4.9 <sup>b</sup>	$18.0 \pm 4.0^{b}$	$24.0\pm 4.9^{b}$	32.0±4.0 <sup>b</sup>		
viscosa	Chloroform	$4.0 \pm 4.49^{\circ}$	$6.0 \pm 4.49^{cd}$	$16.0 \pm 4.49^{bc}$	$22.0 \pm 4.0^{\circ}$		
	Ethyl acetate	$0^{c}$	$6.0 \pm 4.9^{cd}$	$20.0 \pm 8.94^{bc}$	$26.0 \pm 4.9^{bc}$		
C.	Methanol	$0^{c}$	$6.0 \pm 4.9^{cd}$	$12.0 \pm 4.0^{\circ}$	$22.0 \pm 4.0^{\circ}$		
	Hexane	$6.0 \pm 4.9^{\circ}$	$10.0\pm0^{\circ}$	$18.0{\pm}4.0^{\rm bc}$	$24.0\pm 4.9^{bc}$		
alba	Chloroform	$0^{c}$	$0^{d}$	$10.0\pm0^{\circ}$	$22.0 \pm 4.0^{\circ}$		
S. a	Ethyl acetate	$0^{c}$	$0^d$	$0^{d}$	$0^d$		
	Methanol	$22.0{\pm}7.48^{a}$	$32.0 \pm 7.48^{a}$	$38.0 \pm 4.0^{a}$	$54.0{\pm}4.9^{a}$		

Table 3. Pupicidal activity of seed extracts of C. viscosa and S. alba against H. armigera (Mean±SD; n=5).

Means followed by same letter(s) in a column do not differ significantly by Tukey's test at P 0.05.

**Table 4.** Percent deformed adult moths of *H. armigera* produced by *C. viscosa* and *S. alba* seed extract treatments (Mean±SD; n=5).

Plant	Extracts	Concentration (%)					
species	Extracts	0.5	1.0	1.5	2.0		
C. viscosa	Hexane	$5.0\pm6.12^{a}$	12.22±0.56 <sup>a</sup>	13.21±0.87 <sup>b</sup>	$32.86 \pm 8.57^{a}$		
	Chloroform	$0^{\mathrm{a}}$	$0^{c}$	$0^{d}$	$0^{c}$		
	Ethyl acetate	$0^{\mathrm{a}}$	$0^{c}$	$0^{d}$	$0^{c}$		
	Methanol	$0^{\mathrm{a}}$	$0^{c}$	$0^d$	$0^{c}$		
S. alba	Hexane	$6.67 \pm 5.44^{a}$	$13.33 \pm 4.44^{a}$	$19.44 \pm 5.76^{a}$	$33.93 \pm 4.37^{a}$		
	Chloroform	$4.0{\pm}4.9^{a}$	$8.0{\pm}4.0^{b}$	$11.11 \pm 0^{b}$	$20.71 \pm 6.83^{b}$		
	Ethyl acetate	$0^{\mathrm{a}}$	$0^{c}$	$6.0 \pm 4.9^{\circ}$	$16.0 \pm 4.9^{b}$		
	Methanol	$0^{\mathrm{a}}$	$0^{c}$	$0^d$	$0^{c}$		

Means followed by same letter(s) in a column do not differ significantly by Tukey's test at P 0.05.

*S. alba* seed extracts. Among the different treatments, methanol extract of *S. alba* seeds recorded the highest pupal mortality (54%) at 2% concentration. In *S. alba*, the highest pupicidal activity (32%) was recorded in hexane extract. Ethyl acetate extract of *S. alba* did not show any pupicidal activity.

Growth inhibiting properties: Growth inhibiting property of seed extracts of two plants was expressed through adult deformities. The deformed adult moths were recognized by their relatively poor body size, highly curled wings and undergrown wings. High number of deformed adult moths was produced by hexane extracts of the two plants. At 2% concentration hexane extracts of S. alba and C. viscosa seed extracts produced 33.93 and 32.86% deformed adults respectively (Table 4). Chloroform, ethyl acetate and methanol extracts of C. viscosa seeds did not record any malformed adults and methanol extract of S. alba seed did not produce any deformed adults.

# DISCUSSION

Plant seed extracts were found to be possessing feeding deterrent molecules, toxicants and growth inhibitors against *H. armigera* (Sonkamble *et al.*, 2000; Lingathurai *et al.*, 2010; Baskar and Ignacimuthu, 2012). In the present investigation maximum antifeedant activity (71.42%) was observed in methanol extract *S. alba.* 

Previously several investigators (Yasui *et al.*, 1998; Koul *et al.*, 2000; Rongsriyam *et al.*, 2006; Kamaraj *et al.*, 2008) have reported that methanol extracts of plants had higher antifeedant, insecticidal and growth inhibiting activities. Arivoli and Tennyson (2012) showed that methanol extract of *Zanthoxylum limonella* leaves showed antifeedant activity of 44.11% in 72 h at 1000 ppm concentration against *S. litura* larvae. Somboon and Pimsamarn (2006) studied the contact toxicity of methanol extracts of three *Cleome* species against *Sitophilus oryzae*. They found that *C. viscosa* was the most toxic plant, which recorded the least median lethal

concentration  $(LC_{50})$ . They also reported ovipositon deterrent and ovicidal activities of C. viscosa against S. oryzae. Koul et al. (2000) showed that the methanol extract of Melia dubia had antifeedant activity and inhibited the growth of Spodoptera litura and Helicoverpa armigera larvae. Deborah et al. (2001) observed that the methanol extract of Trichilia americana exhibited antifeedant activity of 50% in a choice leaf disc bioassay with 0.18  $\mu g$  cm<sup>-2</sup> on Spodoptera litura. Methanol extracts Ocimum canum and Rhinacanthus nasutus leaves reported mortality of 100% at 1000 ppm against the larvae of S. litura and Culex quinquefasciatus (Kamaraj, et al., 2008). Contrary to the above findings Islam et al. (2014) reported that chloroform extract of C. viscosa caused higher mortality in Tribolium castaneum and Culex mosquito larvae.

Many plants contain phytochemicals like phenols, alkaloids, flavanoids, terpenes, quinone, coumarin etc., which play a defensive role against insect pests. These phytochemicals possess broad range of biological activities such as antifeedant, oviposition deterrent, insecticidal, ovicidal and growth regulatory activities on agricultural pests (Jeyasankar et al., 2012). In the present study the preliminary phytochemical analysis of the effective treatment, the methanol extract of S. alba seeds, revealed the presence of alkaloids, quinone and saponins. Alkaloids present in Tylophora asthmatica plant inhibited the feeding of S. litura (Verma et al., 1986). González-coloma et al. (2004) and Kathuria and Kaushik (2005) reported that alkaloids were effective feeding deterrents against lepidopteran pests.

The present study clearly showed that the seed extracts of C. viscosa and S. alba were not lethal the larval stage, but at the pupal stage. The treatments also caused abnormal growth as evidenced from the development of deformed adults. Methanol extract of S. alba seeds was the most lethal to the pupae and hexane extract produced higher number of abnormal adults. These findings suggested that the active principles in the plant extracts interrupted the metamorphosis process and led to the pupicidal activity and abnormalities in H. armigera. Similarly, Jeyasankar et al. (2012) have reported that ethyl acetate extract of Solanum pseudocapsicum seeds produced pupal mortality and adult deformities in H. armigera. Methanol

extract of *S. alba* seeds contained major groups of phytochemicals such as quinones, alkaloids, saponins and hexane extract contained terpenoids.

# CONCLUSIONS

The present study showed that *C. viscosa* and *S. alba* seed extracts possess antifeedant and chronic toxic properties against *H. armigera*. These extracts can be used for controlling *H. armigera* in ecofriendly pest management programmes.

# **CONFLICTS OF INTEREST**

The authors declare that there are no conflicts of interest associated with this article.

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# REFERENCES

- Aparadh, V.T., Mahamuni, R.J., Karadge, B.A., 2012.Taxonomy and physiological studies in spider flower (Cleomes pecies): Acritical Review. *Plant Sci. Feed.* 2(3): 25-46.
- Arivoli, S and Samuel Tennyson., 2012.
  Antifeedant activity of plant extracts against *Spodoptera litura* (Fab.) (Lepidoptera: Noctuidae). *American-Eurasian J. Agric. Environ. Sci.*, 12(6): 764-768.
- Babu, S.R. and Subrahmanyam, B., 2010. Biopotency of serine proteinase inhibitors from *Acacia senegal* seeds on digestive proteinases, larval growth and development of *Helicoverpa armigera* (Hübner). *Pestic. Biochem. Physiol.*, 98: 349-358.
- Baskar, K and Ignacimuthu, S., 2012. Antifeedant, larvicidal and growth inhibitory effects of ononitol monohydrate isolated from *Cassia tora* L. against *Helicoverpa armigera* (Hub.) and *Spodoptera litura* (Fab.) (Lepidoptera: Noctuidae). *Chemosphere*, 88: 384-388.
- Bentley, M.D., Leonard, D.E., Stoddard, W.F., Zalkow, L.H., 1984. Pyrrolizidine alkaloids as larval feeding deterrents for spruce budworm, *Choristoneura fumiferana* (Lepidoptera: Tortricidae). *Annal. Entomol. Soc. Am.*, 77: 393-397.

- Cassia Seffrin, R., Shikano, I., Akhtar, Y., Murray Isman, B., 2010. Effects of crude seed extracts of *Annona atemoya* and *Annonas quamosa* L. against the cabbage looper, *Trichoplus iani* in the laboratory and greenhouse. *Crop Protection*, 29: 20-24.
- Deborah, A., Wheeler Isman, M.B., 2001. Antifeedant and toxic activity of *Trichilia americana* extract against the larvae of *Spodoptera litura*. Entomologia *Experimentaliset Applicata*, 98: 9-16.
- Dhingra, S., Phokela, A., Mehrotra, K.N., 1988. Cypermethrin resistance in the populations of *Heliothis armigera* Hübner. *Natl. Acad. Sci. Lett.*, 11: 123-125.
- González-coloma, A., Reina, M., Medinaveitia, A., Guadan, A., Santana, O., martínez-díaz, R., Ruiz-mesia, L., Alva, A., Grández, M., Díaz, R., Gavin, J.A., Fuente, G.D., 2004. Structural diversity and defensive properties of norditerpenoid alkaloids. J. Chem. Ecol., 30:1393-1408.
- Harborne, J.B., 1998. Phytochemical Methods: A Guide to Modern Techniques of Plant Analysis. Chapman and Hall, London, pp.295.
- Islam, M.M., Islam, M.Z., Shaekh, M.P.E., Das, P., Chowdhury, H.K., Shahik, S.M., Muzahid, N.H., Khan, M.A., Ekram, A.E., 2014. Screening of *Cleome viscosa* (L.) for Dose Mortality, Insect Repellency, Cytotoxicity and Larvicidal Activities in the Laboratory Condition. *Int. J. Sci. Eng. Res.*, 5: 2201-2212.
- Jadhav, D.R., Mallikarjuna, N., Rathore, A. and Pokle, D., 2012. Effect of Some Flavonoids Survival and Development of on (Hübner) Helicoverpa armigera and Spodoptera litura (Fab) (Lepidoptera: Noctuidae). Asian J. Agr. Sci., 4(4): 298-307.
- Jeyasankar, A., Premalatha, S., Elumalai, K., 2012. Biological activities of Solanum pseudocapsicum (Solanaceae) against cotton bollworm, Helicoverpa armigera Hubner and armyworm, Spodoptera litura Fabricius (Lepidotera: Noctuidae). Asian Pac. J. Trop. Biomed., 2(12): 981-986.
- Kamaraj, C., Abdul Rahuman, A., Bagavan, A., 2008. Antifeedant and larvicidal effects of

plant extracts against *Spodoptera litura* (F.), *Aedes aegypti* L. and *Culex quinquefasciatus* Say. *Parasitol. Res.*, 103: 325-331.

- Kathuria, V and Kaushik, N., 2005. Feeding inhibition of *Helicoverpa armigera* (Hubner) by *Eucalyptus camaldulensis* and *Tylophora indica* extracts. *Insect Sci.*, 12, 249-254.
- Koul, O., Jain, M.P., Sharma, V.K., 2000. Growth inhibitory and antifeedant activity of extracts from *Melia dubia* to *Spodoptera litura* and *Helicoverpa armigera* larvae. *Indian J. Exp. Biol.*, 38(1): 63-68.
- Ladhari, A., Laarif, A., Omezzine1, F and Haouala, R., 2013. Effect of the extracts of the spiderflower, *Cleome arabica*, on feeding and survival of larvae of the cotton leafworm, *Spodoptera littoralis*. *Journal of Insect Science*, 13: 1-14.
- Lingathurai, S., War, A.R., Paulraj, M.G., Ignacimuthu, S., 2010. Antifeedant and insecticidal activities of some plants against *Helicoverpa armigera* Hubner (Lepidoptera: Noctuidae) larvae. In: Non chemical insect pest management (eds. Ignacimuthu, S. and David, B.V.). Elite Publishing House, New Delhi, pp. 58-63.
- Metcalf, R.L., Metcalf, E.R., 1992. Plant kairomones in insects ecology and control. 1<sup>st</sup>, edn. Chapman and Hall, UK.
- Nathala, E. and Dhingra, S., 2006. Biological effects of *Caesalpinia crista* seed extracts on *Helicoverpa armigera* (Lepidoptera: Noctuidae) and its predator, *Coccinella septumounctete* (Coleoptera: Coccinellidae). *J. Asia-Pac. Entomol.*, 9: 159-164.
- Paulraj, M.G. and Ignacimuthu, S., 2009. Impact of neem and karanj oil formulation and *Hyptis suaveolens* leaf extract on feeding, survival, histology and protein profile of four lepidopteran pests. In: Ecofriendly Insect Pest Management (eds. S. Ignacimuthu and B.V. David) Elite Publishing House Pvt. Ltd. pp. 200-209.
- Pavunraj, M., Baskar, K., Gabriel Paulraj, M., Ignacimuthu, S. and Janarthanan, S., 2013.
  Phagodeterrence and insecticidal activity of *Hyptis suaveolens* (Poit.) against four important lepidopteran pests. *Arch. Phytopathol. Plant Prot.*, 47: 113-121.

- Ragurman, S. Singh, R.P., 1999. Biological effects of neem (*Azadirachta indica*) seed oil on egg parasitoid, *Trichogramma chilonis. J. Econ. Entomol.*, 92: 1274-1280.
- Rongsriyam, Y., Trongtokit, Y., Komalamisra., N., Sinchaipanich, N., Apiwathnasorn, C., Mitrejet, A., 2006. Formulation of tablets from the crude extract of *Rhinacanthus* nasutus (Thai local plant) against Aedes aegypti and Culex quinquefasciatus larvae: a preliminary study. Southeast Asian J. Trop. Med. Public Health, 37(2): 265-271.
- Sivaraman, G., Gabriel Paulraj, M., Ignacimuthu,
  S., Al-Dhabi, N.A., 2014. Bioefficacy of seed extracts of *Strychnos nux-vomica* and *Semicarpus anacardium* against *Helicoverpa armigera* (Hubnar) (Lepidoptera: Noctuidae). *Int. J. Agr. Food Sci.*, 4(2): 73-77.
- Somboon,S and Pimsamarn, S., 2006. Biological Activity of *Cleome* spp. Extracts Against the Rice Weevil, *Sitophilus oryzae* L. *Agr. Sci. J.* 37: 5 (Suppl.): 232-235.
- Sonkamble, M.M., Dhanorkar, B.K., Munde, A.T and Sonkamble, A.M., 2000. Efficacy of plant indigenous extracts against Helicoverpa armigera (Hubner) and litura (Fabricius) *Spodoptera* under laboratory conditions. J. Soils Crops, 10: 236-239.

- Talekar, N.S., Open'a, R.T., Hanson, P., 2006. *Helicoverpa armigera* management: a review of AVRDC's research on host plant resistance in tomato. *Crop Protect.*, 5:461-467.
- Vanderzant, E.S., Richard, C.D and Fort, S.W., 1962. Rearing bollworm of artificial diet. *J. Eco. Entomol.*,55:140.
- Verma, G.S., Ramakrishnan, V., Mulchandani, N.B. and Chadha, M.S., 1986. Insect feeding deterrents from the medicinal plant *Tylophora asthmatica. Entomol. Exp. Appl.*, 40: 99-101.
- Wondafrash, M., Getu, E and Terefe, G., 2012.
  Life-cycle Parameters of African bollworm, *Helicoverpa armigera* (Hubner)
  (Lepidoptera :Noctuidae) affected by Neem, *Azadirachta indica* (A. Juss) Extracts. *Agr. Sci. Res. J.*, 2(6): 335-345.
- Yuan, H., Zhu, M., Guo, W., Jin, L., Chen, W., Brunk, T.U., Zhao, M., 2011. Mustard seeds (*Sinapis alba Linn*) attenuate azoxymethaneinduced colon carcinogenesis. *Redox Report Commun. Free Radical Res.*, 16(1): 38-44.
- Yasui, H., Kato, A., Yazawa, M., 1998. Antifeedants to armyworms, Spodoptera litura and Pseudaletia separata, from bitter gourd leaves, Momordica charantia. J. Chem. Ecol., 4(5): 803-813.